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ORIGINAL RESEARCH

Haploinsufficiency of Endothelial Nitric Oxide Synthase Mitigates Beneficial Effects of Maternal Exercise on Fetal Heart Development During Pregestational Diabetes

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BACKGROUND: Pregestational diabetes (PGD) increases congenital heart defect (CHD) risk over 5-fold. Maternal exercise enhances eNOS (endothelial nitric oxide synthase) activity, benefiting embryos, though its causal role remains unclear. This study investigated the role of eNOS in maternal exercise-mediated protection of fetal heart development in a PGD mouse model.

METHODS: PGD was induced in eNOS$^{+/-}$ or wild-type female mice via streptozotocin before breeding with wild-type or eNOS$^{+/-}$ males. Pregnant females had access to a running wheel for voluntary exercise or remained sedentary. Fetuses were collected at embryonic day 18.5 for genotyping and CHD assessment. Embryonic day 12.5 hearts were analyzed for proliferation, apoptosis, oxidative stress, and eNOS protein levels.

RESULTS: Maternal exercise normalized litter size and mortality rates in offspring of diabetic eNOS$^{+/-}$ females but did not reduce CHD incidence in offspring of wild-type or eNOS$^{+/-}$ females with PGD. CHDs included septal defects, double outlet right ventricle, and valve defects. Exercise increased coronary artery density but not capillary density. Proliferation deficits at embryonic day 12.5 were restored by exercise, yet oxidative stress remained elevated. Maternal exercise in eNOS$^{+/-}$ dams during PGD did not significantly change eNOS protein or phosphorylation levels in both eNOS$^{+/+}$ and eNOS$^{+/-}$ fetal hearts. Offspring genotype did not affect CHD incidence, cell proliferation, apoptosis, or oxidative stress.

CONCLUSIONS: Maternal exercise does not prevent CHDs in PGD offspring of eNOS$^{+/-}$ mice. Its ability to mitigate PGD-induced oxidative stress is eNOS dependent and essential for improving heart morphology.

Key Words: congenital heart defects ■ embryonic heart development ■ endothelial nitric oxide synthase ■ maternal exercise ■ oxidative stress ■ pregestational diabetes

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Congenital heart defects (CHDs) are a leading cause of death in the first year of infant life and arise due to a variety of genetic, maternal, and environmental factors.[1,2] Pregestational diabetes (PGD) has been identified as an environmental factor that increases the risk of CHDs by more than 5 times.[1–3] The prevalence of CHDs continues to rise globally; although this can be attributed to improved

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CLINICAL PERSPECTIVE

What Is New?

- eNOS (endothelial nitric oxide synthase) is a critical mediator of the cardioprotective effects of maternal exercise during pregestational diabetes, and reduced eNOS expression abrogates the ability of maternal exercise to prevent congenital heart defects in diabetic pregnancies.
- Maternal exercise failed to normalize oxidative stress or eNOS signaling in fetuses of eNOS haploinsufficient dams, underscoring the necessity of intact eNOS function for exercise-mediated fetal cardiac benefits.

What Are the Clinical Implications?

- Maternal endothelial function and NO bioavailability are key determinants of fetal cardiac outcomes in diabetic pregnancies, suggesting that enhancing eNOS activity or NO signaling through pharmacologic or lifestyle approaches could improve fetal heart development in women with pregestational diabetes.

Nonstandard Abbreviations and Acronyms

4-HNE	4-hydroxynonenal
αSMA	α-smooth muscle actin
CHD	congenital heart defect
eNOS	endothelial nitric oxide synthase
eNOS$^{+/-}$	eNOS heterozygous
eNOS$^{+/+}$	wild-type
PGD	pregestational diabetes
pHH3	phosphorylated histone H3
ROS	reactive oxygen species

diagnostic screening for asymptomatic conditions, it is also correlated with an increase in diabetes prevalence in women of reproductive age.[4,5] The elevated blood glucose levels associated with PGD interfere with fetal heart development by increasing the production of reactive oxygen species (ROS) and oxidative stress, resulting in abnormalities in cellular apoptosis, proliferation, epithelial-to-mesenchymal transition, and abnormal placental function, ultimately contributing to alterations in development.[6–9]

We have shown that maternal exercise lowers CHD incidence and improves coronary vasculature development in offspring exposed to PGD.[10] Specifically, maternal exercise mitigated diabetes-induced changes to myocardial cell proliferation, epithelial-to-mesenchymal transition, and cardiac gene expression. Most notably, maternal exercise significantly lowered oxidative stress levels in fetal hearts as demonstrated by a reduction in ROS levels.[10] However, the molecular mechanisms responsible for stimulating these beneficial changes remain to be elucidated.

eNOS (endothelial nitric oxide synthase) is expressed in the developing heart as early as embryonic day (E)8.5.[11] Nitric oxide (NO) produced by eNOS plays a crucial role in embryonic development, particularly in the formation and maturation of the heart.[12–14] NO produced by eNOS contributes to processes essential to heart development including promoting cell survival, cell differentiation, epithelial-to-mesenchymal transition, and proliferation. These mechanisms are vital for regulating cardiac septation, valve formation, and the development of the coronary vasculature.[11,15–18] Furthermore, eNOS-derived NO plays a pivotal role in the fine-tuning of cardiovascular morphogenesis, ensuring the proper structural and functional integration of the fetal heart. Disruption in NO signaling pathways during this critical period can lead to heart defects, highlighting the importance of eNOS in early cardiac development.[11,15]

Exercise has been shown to induce eNOS expression and activity in the vascular endothelium in both nondiabetic and diabetic individuals.[19] Maternal exercise increases eNOS phosphorylation in the fetal heart of offspring exposed to PGD.[10] However, the causal relationship between eNOS function and the benefits of maternal exercise on the developing heart in PGD has not been established. In this study, we examined the hypothesis that eNOS haploinsufficiency would prevent the rescuing effects of maternal exercise on heart development in offspring exposed to PGD.

METHODS

All data and supporting materials have been provided with the published article.

Animals

Mice were used in this study in keeping with the Canadian Council on Animal Care and Use Animal Subcommittee guidelines of Western University, Canada (Protocol #2020–128). Animal suffering and number were minimized in this study. The animals were provided with standard chow and water ad libitum and housed in a 12-hour light/dark cycle. Wild-type (eNOS$^{+/+}$) and eNOS homozygous knockout (eNOS$^{-/-}$, *Nos3^{tm1Unc}*, strain #002684) C57BL/6 male and female mice were purchased from Jackson Laboratory (Bar Harbor, ME, USA). An eNOS$^{+/-}$ line was generated by crossing eNOS$^{-/-}$ females with wild-type (WT) males at 8 weeks old. eNOS$^{+/-}$ offspring were used in all experiments going forward.

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Experimental breeding procedures consisted of crossing eNOS$^{+/-}$ females with WT males to produce offspring in the same litter that were WT or eNOS$^{+/-}$. To determine the impact of maternal eNOS$^{+/-}$, an opposing cross was used where WT females were bred with eNOS$^{+/-}$ males to maintain the same litter composition. An eNOS$^{+/-}$ line was used to avoid the spontaneous cardiac defects associated with eNOS$^{-/-}$ lines, such as cardiac septation and coronary artery defects.[11,20] Mice were randomly assigned to different experimental groups.

Induction of Pregestational Diabetes and Voluntary Exercise

At 8 weeks of age, WT and eNOS$^{+/-}$ female mice (n=13 and n=5, respectively) were injected intraperitoneally with streptozotocin (Sigma-Aldrich) dissolved in 10 mM sodium citrate for 3 consecutive days at a dose of 75 mg/kg.[10] An additional group of eNOS$^{+/-}$ females served as a control and were left untreated (n=5). Blood glucose after 4 hours of fasting was measured 1 week following the third injection via the tail clip method using the OneTouch Verio2 glucose meter (LifeScan). Mice with blood glucose levels ≥11mmol/L were considered diabetic.

Once diabetes was confirmed, a subset of diabetic WT and eNOS$^{+/-}$ mice (n=5 and n=6, respectively) were housed in individual cages with access to a voluntary free-spinning running wheel (31.92 cm circumference, Columbus Instruments) for a 1-week acclimatization period.[10] Running wheel revolutions were recorded over 24-hour periods using Multi-Device Channel Interface Software (Columbus Instruments). Daily running distance was calculated by multiplying the number of revolutions by the circumference of the running wheel (Table S1). Females that traveled less than 2 km on average per day were excluded from the study. Healthy control eNOS$^{+/-}$ females and diabetic eNOS$^{+/-}$ females were placed in individual cages without a running wheel. Following acclimatization or a 1-week nonexercising period, females were mated overnight in cages without a running wheel with males ranging from 8 to 16 weeks of age. The presence of a vaginal plug indicated E0.5. Four-hour fasting blood glucose and body weight were assessed at E0.5 to E18.5 during gestation. There was a total of 8 groups of offspring. Three groups of eNOS$^{+/-}$ dams crossed with WT males gave rise to control WT and eNOS$^{+/-}$, PGD eNOS$^{+/+}$ and eNOS$^{+/-}$, and PGD+exercise WT and eNOS$^{+/-}$offspring. The seventh and eighth offspring groups were WT and eNOS$^{+/-}$ from WT dams with PGD+exercise crossed with eNOS$^{+/-}$ males.

Genotyping

Tails and upper limbs were collected to determine fetal genotype. DNA was extracted by incubation with 50 mM sodium hydroxide at 95 °C for 3 hours and subsequently neutralized using 1 M Tris–HCl. End point polymerase chain reaction was performed using DreamTaq Green PCR Master Mix (K1081, Thermo Fisher Scientific, Mississauga, ON). The eNOS$^{+/-}$ genotype was identified using common primer oIMR9358 (5′ CTT GTC CCC TAG GCA CCT CT 3′), eNOS knockout primer oIMR8963 (5′ AAT TCG CCA ATG ACA AGA CG 3′), and WT primer oIMR9357 (5′ AGG GGA ACA AGC CCA GTA GT 3′). Polymerase chain reaction products were amplified using a touchdown protocol (see Table S2 for protocol). The eNOS$^{+/-}$ genotype is indicated by the presence of both an eNOS knockout fragment at 300 bp and the WT fragment at 337 bp, where the WT genotype displays only a 337 fragment (Figure S1). The polymerase chain reaction products were separated using gel electrophoresis on a 1.7% agarose gel. The proportion of WT to eNOS$^{+/-}$ offspring in each maternal condition can be seen in Figure S2.

Cardiac Morphology Assessments

Offspring collected at E18.5 via cesarean section were dissected to isolate the thoracic region of the body and fixed overnight in 4% paraformaldehyde at 4 °C. The tissue samples were dehydrated using a series of ethanol washes of increasing concentrations (75%, 80%, 95%, 100%), followed by 2 xylene washes before paraffin embedding. Embedded tissues were cut into 5 μm thick transverse serial sections using a Leica RM2255 microtome (Leica Biosystems) beginning at the level of the aortic arch until the apex. To visualize heart morphology, serial sections were stained with hematoxylin and eosin (Sigma-Aldrich; 1% eosin in ethyl alcohol VWR) following deparaffinization using xylene (100%; VWR) and serial ethanol rehydration (100% x2, 95%, 75%). Heart morphology was assessed on a section-by-section basis, and the type of CHD was diagnosed by 2 or more blinded investigators using a light microscope (Zeiss Observer D1).

Immunohistochemical Assessments of Coronary Vasculature

Immunohistochemistry was performed to assess coronary artery and capillary development. Serial sections of E18.5 fetuses were dewaxed and rehydrated using xylene and ethanol. Antigen retrieval was performed in 0.01 M sodium citrate (pH 6.0) for 12 minutes at 94 °C in a microwave (BP-111, Microwave Research & Applications). Anti-αSMA (α-smooth muscle actin) primary antibody 1A4 (A5228, Sigma-Aldrich) was used to identify the smooth muscle component of the coronary arteries at room temperature overnight in a humidity chamber. Biotinylated horse anti-mouse IgG was applied to the tissue for 1 hour at room temperature. Lectin staining was performed in E18.5 heart sections

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using biotinylated griffonia simplicifolia lectin-1 (1:50 dilution, Vector Laboratories) to assess capillary density. Color signals for αSMA and lectin were developed using VECTASTAIN Elite ABC-HRP Kit, Peroxidase (Standard) (PK-6100) followed by 3–3′ di-aminobenzidine tetrahydrochloride (DAB, Sigma-Aldrich). Hematoxylin was used as a counterstain. Quantification of vessel density was done using Zen 3.4 Software by Zeiss. Researchers were blind to the offspring's condition.

Immunofluorescence

Embryonic 12.5 hearts were collected and frozen at −80 °C and subsequently embedded in Optimal Cutting Temperature Embedding Medium (Tissue-Tek, Sakura Finetek) at −20 °C. Fetal hearts were then sectioned into 8 μm thick sections using the Leica CM 1950 Cryostat (Leica Biosystems). Cryosections were washed in PBS and fixed in 100% methanol for 10 minutes at −20 °C. Cell proliferation, apoptosis, lipid peroxidation, and phospho-eNOS were assessed using rabbit anti-phosphorylated histone H3 (pHH3; 1:100; Cell Signaling #9701), rabbit anti-CC3 (cleaved-caspase-3; 1:100; Cell Signaling #9661), goat anti-4-hydroxynonenal (4-HNE; 1:1000; Applied Biological Materials), and rabbit anti-phospho-eNOS (Ser1177) (1:100, ThermoFisher Cat.# PA5-97371), respectively. Primary antibodies were incubated overnight at room temperature, followed by goat-anti-rabbit IgG (H+L) Cy-3 conjugated or donkey-anti-goat IgG (H+L) Cy-3 conjugated antibody (1:1500 dilution; Jackson Laboratories) for 1 hour at room temperature. Slides were then counterstained with DAPI. As a measure of oxidative stress, superoxide levels were quantified using dihydroethidine (Invitrogen Life Technologies). Sections were incubated in 2 μM of dihydroethidine for 30 minutes at 37 °C in a light-protected humidity chamber and counterstained with nuclear DAPI. All fluorescence signals were imaged using a Zeiss Observer D1 fluorescence microscope, and quantification was done using Zen 3.9 Software (Zeiss Germany) or ImageJ (Java 1.8 for PC). The number of proliferating and apoptotic cells was quantified by counting cells positive for pHH3 and CC3, respectively, in Zen 3.9 at 20× magnification. The total cell number was quantified using ImageJ to calculate the ratio of proliferating cells to total cells. The rate of apoptotic cells was determined by comparing the number of apoptotic cells to the myocardial area (μm^2). Lipid peroxidation, superoxide, and phospho-eNOS levels were quantified by densitometric analysis, with measurements obtained at 200× (3 images/heart, 15 locations) and 400× magnification (6 images/heart, 12 locations), respectively.[10,21,22]

eNOS Protein Levels in E12.5 Hearts

Frozen E12.5 and adult hearts were used for eNOS protein expression assessments via Western blotting. Hearts were homogenized in RIPA buffer (150mM NaCl, 1.0% TritonX-100, 0.5% Na deoxycholate, 0.1% SDS, 50 mM Tris pH 8.0, 1 mM EDTA). Tissue lysates were collected after centrifugation at 10 000 *g* (4 °C) for 10 minutes. The Lowry assay (Bio-Rad) was used to assess total protein concentrations and load equal amounts of proteins into a 10% PAGE. Samples were transferred to a PVDF membrane (Millipore). Membranes were incubated with mouse anti-eNOS antibody (1:1000, BD Biosciences) overnight at 4 °C with gentle agitation, followed by mouse anti-α-actinin antibody (Sigma #A7811, 1:2500 dilution) as a loading control. The eNOS signals were visualized using IRDye 800CW Goat anti-mouse IgG secondary antibody and IRDye 680RD goat anti-mouse IgD secondary antibody. Fluorescence signals were visualized using the Odyssey CLx and quantified using Image Studio (Version 3.1, LI-COR Biosciences).

Statistical Analysis

Data were summarized as mean±SEM. An unpaired Student *t* test was performed when comparing 2 groups. One-way ANOVA followed by Tukey's multiple comparison test was used to assess statistical significance between 3 or more groups. The χ^2 test was used to assess CHD incidence, whereas Fisher's exact test was applied for 2 × 2 contingency tables when the expected cell frequencies were <5. Sample size was calculated using Cohen's formula for contingency tables. For 40% 50% effect with α=0.05 and power = 0.8, the calculated sample size was 25 to 39 fetuses per group. Data were analyzed using GraphPad Prism, version 9.0 software. Statistical significance was indicated by a *P* value <0.05.

RESULTS

Body Weight, Fasting Blood Glucose, and Running Distance During Gestation in PGD Dams

Before streptozotocin injection, dams in all 4 maternal conditions underwent a 4-hour fast to establish baseline blood glucose values (Figure S3A). Dams with a baseline fasting blood glucose value of >9.0 mmol/L were excluded from the study (n=5). $eNOS^{+/-}$ and $eNOS^{+/+}$ dams were found to have similar basal fasting blood glucose levels. One week poststreptozotocin injection, both $eNOS^{+/+}$ and $eNOS^{+/-}$ mice had significantly elevated fasting blood glucose values compared with control $eNOS^{+/-}$ dams (Figure S3B).

Fasting blood glucose was measured at E0.5, E11.5, and E18.5 before fetal collection. Diabetic dams, independent of their genotype and exercise status, demonstrated elevated fasting blood glucose levels compared with control throughout gestation (Figure 1A).

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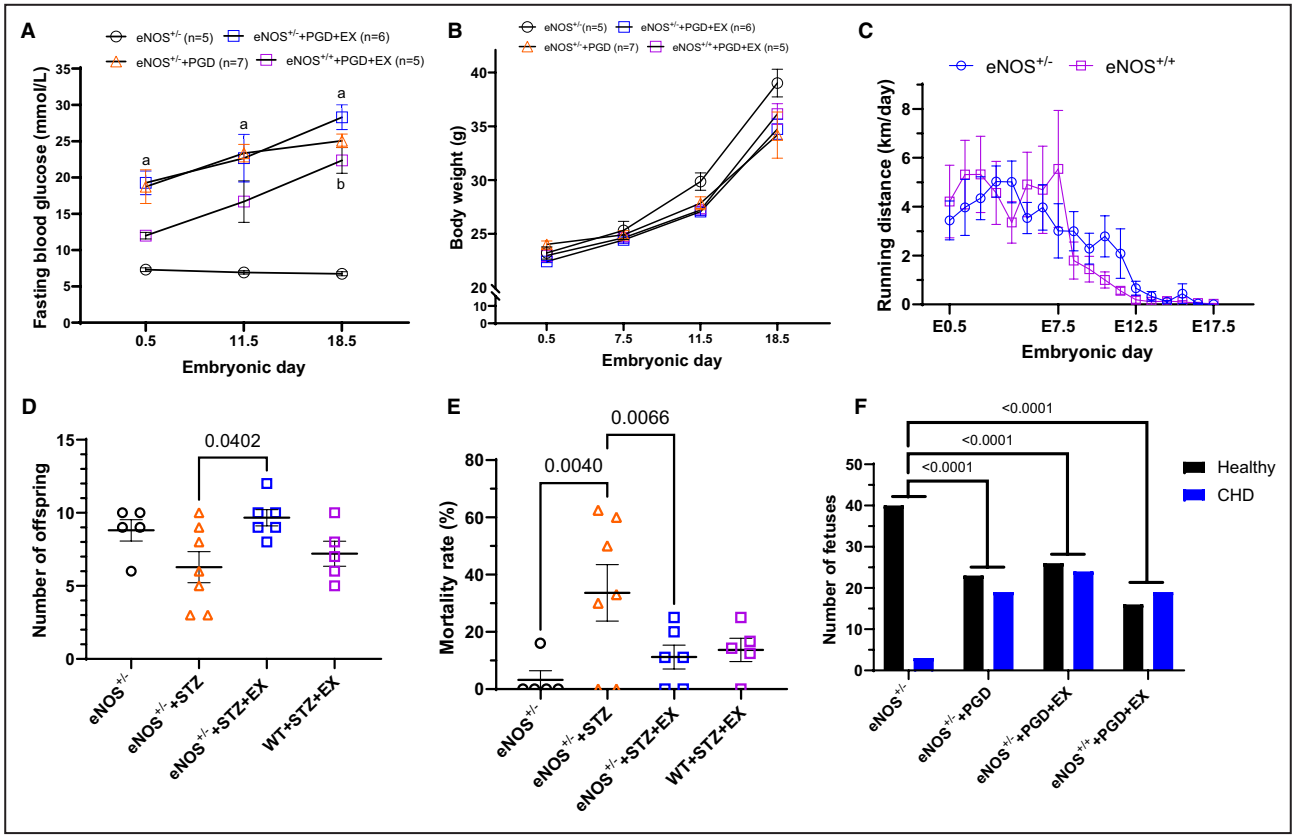


Figure 1. Effects of maternal exercise on blood glucose, body weight, litter size, offspring mortality rate, and congenital heart defects in the offspring of pregestational diabetes.

A, Fasting blood glucose levels in eNOS$^{+/-}$ dams with or without PGD or exercise at E0.5 to 18.5. **B**, Maternal body weight changes during gestation in eNOS$^{+/-}$ dams with or without PGD or exercise. **C**, Running distance of eNOS$^{+/+}$ and eNOS$^{+/-}$ dams with PGD during gestation. **D**, Litter size of eNOS$^{+/-}$ dams with or without PGD or exercise at E18.5. **E**, Mortality rate of fetuses of eNOS$^{+/-}$ dams with or without PGD /or exercise at E18.5. **F**, Number of CHD and healthy fetuses of eNOS$^{+/-}$ dams with or without PGD or exercise at E18.5. The numbers are a combination of both eNOS$^{+/+}$ and eNOS$^{+/-}$ fetuses in each maternal condition. Data represent mean±SEM and were analyzed using 2-way ANOVA followed by Tukey's test (**A**–**E**). a indicates $P<0.0001$ for PGD dams (with or without exercise) vs eNOS$^{+/-}$ control dams (**A**). b indicates $P<0.0001$ for eNOS$^{+/+}$+PGD+exercise vs eNOS$^{+/-}$+PGD+exercise dams (**A**). Fetal CHD numbers were analyzed using a χ^2 test. +/– indicates heterozygous; CHD, congenital heart defect; E, embryonic day; eNOS, endothelial nitric oxide synthase; EX, exercise; PGD, pregestational diabetes; STZ, streptozotocin; and WT, wild-type.

Additionally, fasting blood glucose was significantly elevated as gestation progressed in dams with PGD. Maternal exercise did not improve fasting blood glucose in diabetic eNOS$^{+/+}$ and eNOS$^{+/-}$ dams (Figure 1A). Body weight measurements were completed on E0.5, E7.5, E11.5, and E18.5 to ensure dam progression through gestation (Figure 1B). PGD and maternal exercise did not significantly alter maternal body weight at any time point throughout gestation.

Between E0.5 and E10.5, diabetic dams traveled ~2 to 6 kilometers (km) daily (Figure 1C). Following E10.5, running distance for both eNOS$^{+/+}$ and eNOS$^{+/-}$ genotypes steeply declined due to pregnancy weight gain. The maternal genotype did not alter the running distance throughout gestation.

To collect fetuses for assessments of cellular proliferation, apoptosis, oxidative stress, and eNOS protein levels at E12.5, a separate cohort of eNOS$^{+/-}$ dams with or without PGD/maternal exercise was studied, and the maternal fasting glucose, body weight, and running distance are shown in Figure S4.

Voluntary Maternal Exercise Normalized Litter Size and Reduced Mortality Rate in eNOS$^{+/-}$ Dams With PGD

Litter size and mortality rate (% dead) were tabulated at E18.5 during cesarean fetal collection. PGD status affected litter size in eNOS$^{+/-}$ dams (Figure 1D). Control dams produced litters with a mean size of 8.8±0.7, whereas PGD reduced the mean litter size to 6.3±1.0. Maternal exercise in eNOS$^{+/-}$ dams significantly elevated litter size to a mean of 9.7±0.6 compared with eNOS$^{+/-}$ dams with PGD alone. eNOS$^{+/+}$ dams bred with eNOS$^{+/-}$ males did not experience a significant increase in litter size despite maternal exercise (7.2±0.9).

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Table 1. Mortality Incidence by Genotype in Embryonic Day 18.5 Fetuses From eNOS$^{+/+}$ or eNOS$^{+/-}$ Diabetic Dams With or Without Maternal Exercise

Maternal condition	eNOS$^{+/-}$ (5 litters)		eNOS$^{+/-}$+PGD (7 litters)		eNOS$^{+/-}$+PGD+exercise (6 litters)		eNOS$^{+/+}$+PGD+exercise (5 litters)	
Offspring number	n=44		n=42		n=48		n=35	
Offspring Genotype	eNOS$^{+/+}$ (n=24)	eNOS$^{+/-}$ (n=20)	eNOS$^{+/+}$ (n=21)	eNOS$^{+/-}$ (n=21)	eNOS$^{+/+}$ (n=28)	eNOS$^{+/-}$ (n=22)	eNOS$^{+/+}$ (n=21)	eNOS$^{+/-}$ (n=14)
Alive	23 (96%)	19 (95%)	12 (57%)	19 (90%)	25 (89%)	18 (82%)	18 (86%)	13 (93%)
Dead	1 (4%)	1 (5%)	9 (43%)	2 (10%)	3 (11%)	4 (18%)	3 (14%)	1 (7%)
P values		>0.9999		0.0325		0.6843		0.6350

Data were analyzed using Fisher's exact test. *P* values are comparisons of eNOS$^{+/-}$ vs eNOS$^{+/+}$ littermate offspring within each group. The number of litters is equivalent to the number of dams used in each condition. +/– indicates heterozygous; eNOS, endothelial nitric oxide synthase; and PGD, pregestational diabetes.

Additionally, the fetal mortality rate was significantly elevated by PGD compared with control (Figure 1E). Maternal exercise in eNOS$^{+/-}$ dams with PGD significantly reduced the mortality rate (Figure 1E). The same trend was observed in fetuses from exercising eNOS$^{+/+}$ dams with PGD, suggesting maternal exercise promotes fetal survival until E18.5 following PGD exposure regardless of maternal genotype.

eNOS$^{+/-}$ fetuses from eNOS$^{+/-}$ dams with PGD experienced a lower mortality incidence than their eNOS$^{+/+}$ counterparts as shown in Table 1. Fetal genotype did not influence mortality risk in fetuses from eNOS$^{+/-}$ control, eNOS$^{+/-}$+PGD+exercise, or eNOS$^{+/+}$+PGD+exercise dams. These data suggest that eNOS heterozygous insufficiency may be protective against PGD exposure alone.

Voluntary Maternal Exercise Does Not Reduce CHD Incidence in Fetuses of eNOS$^{+/+}$ or eNOS$^{+/-}$ Dams With PGD

The incidence of CHDs in E18.5 fetuses exposed to PGD with or without maternal exercise was significantly elevated compared with control (Figure 1F). Additionally, CHD incidence was not impacted by maternal genotypes as fetuses exposed to PGD, maternal exercise, and differing maternal genotypes (eNOS$^{+/+}$ or eNOS$^{+/-}$) had similar CHD incidences. Therefore, independently of maternal genotype, fetal eNOS haploinsufficiency prevented the rescuing effects of maternal exercise on heart development, which we previously demonstrated.[10]

The most diagnosed defects in eNOS$^{+/+}$ and eNOS$^{+/-}$ fetuses exposed to PGD alone (Table 2) were atrial septal defect (43% and 48%, respectively), ventricular septal defect (14% and 14%), ventricular hypertrophy (14% and 19%), and thickening of both the pulmonary (20% and 14%) and aortic valves (10% and 14%) (Figure 2A through 2P). Other defects observed in this population at lower incidence were atrioventricular septal defect (Figure 2G), double outlet right ventricle (Figure 2M), bicuspid aortic valve (Figure 2N), and cardiac rupture/hemopericardium (Figure 3A through 3E). The incidence of these defects was not lowered in eNOS$^{+/+}$ or eNOS$^{+/-}$ fetuses exposed to voluntary maternal exercise, as atrial septal defect incidence remained around 50%, regardless of maternal or fetal genotype (Table 2). Notably, the incidence of ventricular hypertrophy was found to be significantly elevated in eNOS$^{+/+}$ fetuses of eNOS$^{+/+}$ dams with PGD and maternal exercise (29%) compared with eNOS$^{+/+}$ fetuses of eNOS$^{+/-}$ dams with PGD and exercise (4%). Ultimately, fetal eNOS heterozygosity did not significantly change the risk of developing any type of CHDs when fetuses were exposed to PGD (Table 2).

Fetal and Maternal eNOS Haploinsufficiency Coupled With PGD and Maternal Exercise Introduced Craniofacial Defects

No fetuses with craniofacial defects were collected from eNOS$^{+/-}$ control or eNOS$^{+/+}$+PGD+exercise dams (Figure 4A). Fetuses exposed to PGD in eNOS$^{+/-}$ mothers with and without maternal exercise had 3 and 5 cases of craniofacial defects, respectively (Figure 4A). An array of craniofacial defects was observed, including cleft lip, encephalocele, and exencephaly (Figure 4C and 4E); however, the majority of fetuses had healthy snout morphology (Figure 4B and 4D). A bilateral cleft lip was observed in our study as the skin failed to close on both sides of the snout (Figure 4C). Encephalocele occurs when the brain and meninges grow through a hole in the fetus' skull due to a failure of the neural tube to close (Figure 4C). Exencephaly also presents as a protrusion of the brain outside of the skull; however, it is not covered in meninges (Figure 4E). There was no statistical significance in any subgroup comparisons due to low incidences. As PGD with maternal exercise in eNOS$^{+/+}$ dams did not produce any craniofacial defects, the data suggest that maternal eNOS$^{+/-}$ and PGD may contribute to a higher risk of craniofacial defects.

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Table 2. Types of CHDs in Fetuses of eNOS$^{+/+}$ or eNOS$^{+/-}$ Dams With PGD, With or Without Maternal Exercise

Maternal condition	eNOS$^{+/-}$ (5 litters)		eNOS$^{+/-}$+PGD (7 litters)		eNOS$^{+/-}$+PGD+exercise (6 litters)		eNOS$^{+/+}$+PGD+exercise (5 litters)	
Offspring genotype	eNOS$^{+/+}$ (n=24)	eNOS$^{+/-}$ (n=20)	eNOS$^{+/+}$ (n=21)	eNOS$^{+/-}$ (n=21)	eNOS$^{+/+}$ (n=28)	eNOS$^{+/-}$ (n=22)	eNOS$^{+/+}$ (n=21)	eNOS$^{+/-}$ (n=14)
Normal	23 (92%)	18 (90%)	12 (57%)	11 (52%)	15 (54%)	11 (50%)	9 (43%)	7 (50%)
Abnormal	1 (4%)	2 (10%)	9 (43%)*	10 (48%)*	13 (46%)*	11 (50%)*	12 (57%)*	7 (50%)*
Atrial septal defect	1 (4%)	2 (10%)	9 (43%)*	9 (43%)	8 (29%)*	6 (27%)	10 (48%)*	6 (43%)
Ventricular septal defect	0	0	3 (14%)	3 (14%)	5 (18%)	2 (9%)	2 (10%)	0
Atrioventricular septal defect	0	0	0	0	1 (4%)	0	0	0
Double outlet right ventricle	0	0	1 (5%)	0	2 (7%)	1 (5%)	3 (14%)	1 (7%)
Ventricular hypertrophy	0	0	3 (14%)	4 (19%)	1 (4%)	1 (5%)	6 (29%)*,†	3 (21%)
Cardiac rupture/ hemopericardium	0	0	0	0	0	1 (5%)	0	0
Bicuspid aortic valve	0	2 (10%)	0	3 (14%)	2 (7%)	0	2 (10%)	0
Aortic valve thickening	0	0	2 (10%)	3 (14%)	1 (3.5%)	1 (5%)	0	3 (21%)
Pulmonary valve thickening	0	0	4 (20%)	3 (14%)	1 (3.5%)	1 (5%)	0	3 (21%)
Hypoplastic aorta	0	0	0	1 (5%)	0	0	0	0

Data were analyzed using chi-square or Fisher's exact test. Comparisons were made between offspring of the same genotype from eNOS$^{+/-}$ control dams and experimental groups (*P<0.05), and between offspring of the same genotype from eNOS$^{+/-}$+PGD+exercise dams vs eNOS$^{+/+}$+PGD+exercise dams (†P<0.05). The number of litters is equivalent to the number of dams used in each condition. +/– indicates heterozygous; CHD, congenital heart defect; eNOS, endothelial nitric oxide synthase; and PGD, pregestational diabetes.

Fetal genotype alone did not increase the risk of developing a craniofacial defect as evidenced by eNOS$^{+/+}$ and eNOS$^{+/-}$ from control dams displaying no incidences of craniofacial defects (Table 3). Fetuses exposed to PGD alone in eNOS$^{+/-}$ dams displayed no trend in craniofacial defect development correlated with fetal genotype. Additionally, PGD with maternal exercise in eNOS$^{+/+}$ dams displayed no fetal genotype dichotomy in the incidence of craniofacial defects (Table 3).

Maternal Exercise Increased Coronary Artery Abundance in eNOS$^{+/+}$ and eNOS$^{+/-}$ Fetuses of eNOS$^{+/-}$ Dams With PGD

The smooth muscle cells of the coronary arteries were labeled with αSMA in E18.5 hearts to assess for coronary malformations (Figure 5A through 5D and 5D'). PGD did not significantly affect coronary artery abundance, as eNOS$^{+/+}$ and eNOS$^{+/-}$ fetuses exposed to eNOS$^{+/-}$ dams with PGD displayed similar coronary artery abundance to myocardial area ratios (Figure 5E). Maternal exercise in eNOS$^{+/-}$ dams with PGD significantly increased fetal coronary artery abundance to myocardial area ratios compared with eNOS$^{+/-}$ control and eNOS$^{+/-}$ dams with PGD conditions (Figure 5E and 5F). However, the same trend was not observed in fetuses from exercising eNOS$^{+/+}$ dams with PGD displaying coronary artery abnormalities that were reflective of fetuses from eNOS$^{+/-}$ control and eNOS$^{+/-}$ with PGD.

The benefits of maternal exercise on coronary artery development were observed in eNOS$^{+/+}$ fetuses but not in eNOS$^{+/-}$ fetuses (Figure 5F and 5G). However, there was no statistical difference in coronary artery abundance to myocardial area between eNOS$^{+/+}$ and eNOS$^{+/-}$ fetuses in any maternal conditions (Figure S5).

Capillary Density Was Not Affected by Maternal Exercise or PGD in eNOS$^{+/+}$ or eNOS$^{+/-}$ Fetuses

Capillaries were visualized using biotinylated lectin-1 and capillary density was quantified using 3 images for the right ventricular myocardium, interventricular septum, and left ventricular myocardium taken at the 4-chamber level (Figure 6A through 6D). Capillary density was normalized to the area of the myocardium and was not significantly altered by changes in maternal genotype, PGD, or maternal exercise (Figure 6E through 6G). Additionally, there were no significant differences in capillary density relative to the myocardial area between eNOS$^{+/+}$ and eNOS$^{+/-}$ fetuses in each maternal condition (Figure S6). However, it should be noted that capillary density in eNOS$^{+/-}$ fetuses tended to be lower than their eNOS$^{+/+}$ counterparts.

Maternal Exercise Normalizes Cellular Proliferation During PGD in E12.5 eNOS$^{+/+}$ and eNOS$^{+/-}$ Hearts

Cellular proliferation was assessed using pHH3 immunofluorescence staining in E12.5 fetal heart

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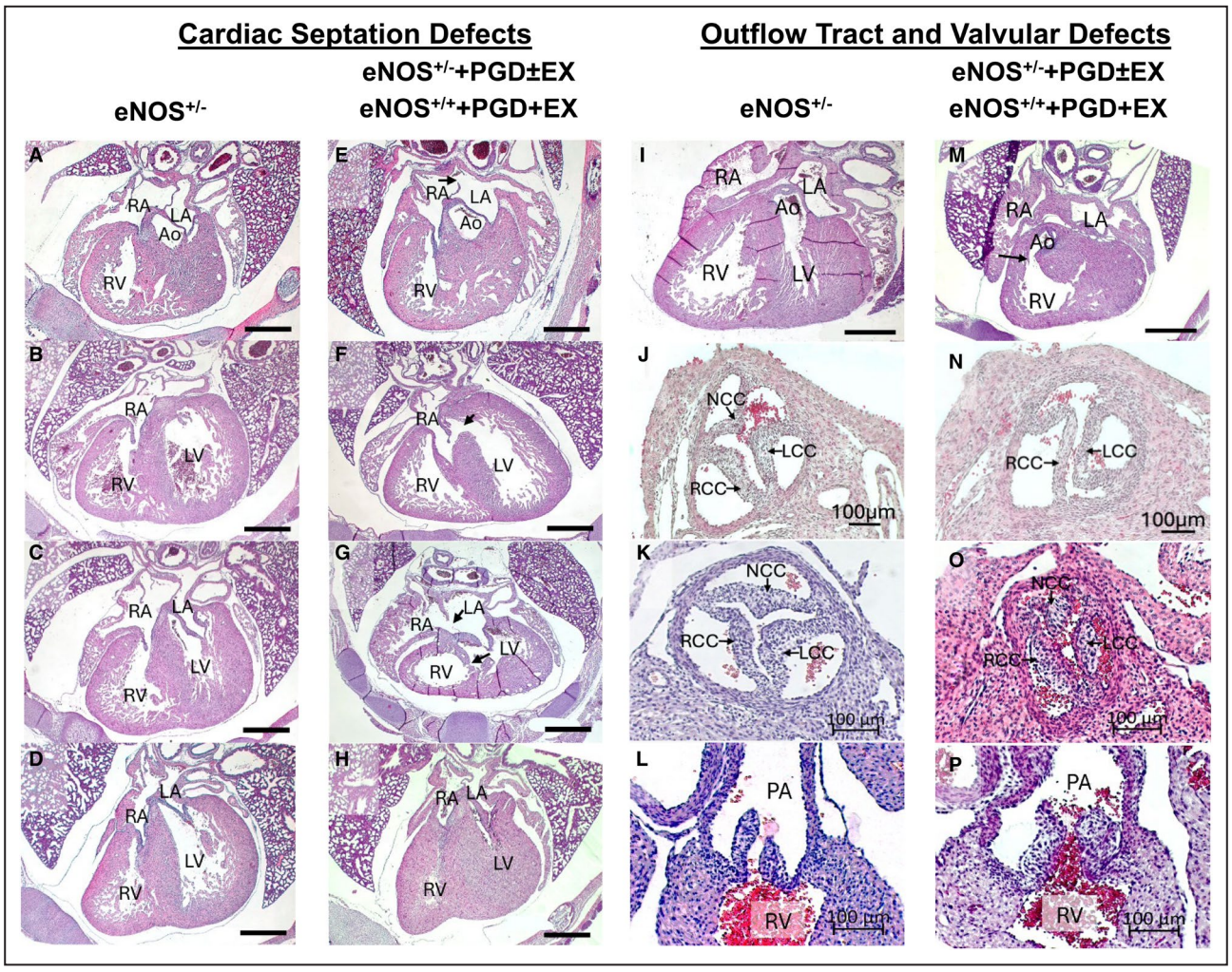


Figure 2. Representative images of cardiac septation defects, myocardial hypertrophy, outflow tract and valve defects in E18.5 eNOS$^{+/+}$ and eNOS$^{+/-}$ offspring exposed to PGD and maternal exercise.

A–**C**, Normal cardiac septation morphology from eNOS$^{+/-}$ control dams; **D**, Normal myocardial wall thickness; **E**, ASD type 1; **F**, Perimembranous VSD; **G**, AVSD with 1 common valve; **H**, Myocardial hypertrophy; **I**, Normal right ventricle and aorta morphology; **J**, **K**, Normal aorta morphology (tricuspid); **L**, Normal aorta connected to LV;, **M**, Aorta connected to RV as part of DORV (position more than 50% above RV); **N**, BAV; **O**, thickened aortic leaflets; **P**, thickened pulmonary valve leaflets. Scale bar is 500 µm in heart images and 100 µm in valve images. +/– indicates heterozygous; Ao indicates aorta; E, embryonic day; eNOS, endothelial nitric oxide synthase; EX, exercise; LA, left atrium; LCC, left coronary cusp; LV, left ventricle; NCC, noncoronary cusp; PA, pulmonary artery; PGD, pregestational diabetes; RA, right atrium; RCC, right coronary cusp; and RV, right ventricle.

cryosections of eNOS$^{+/-}$ control dams (Figure 7A through 7B). The ratio of proliferating cells to total cells was quantified (Figure 7C). Basal rates of proliferation were not significantly affected by pup genotype.

PGD in eNOS$^{+/-}$ dams significantly reduced cellular proliferation in both eNOS$^{+/+}$ and eNOS$^{+/-}$ hearts at E12.5 (Figure 7D through 7I). Maternal exercise in eNOS$^{+/-}$ dams returned cellular proliferation in eNOS$^{+/+}$ and eNOS$^{+/-}$ fetal hearts to control levels (Figure 7D through 7I). Additionally, differences in proliferation between eNOS$^{+/+}$ and eNOS$^{+/-}$ fetal hearts following PGD exposure with or without maternal exercise were assessed. No significant differences in cellular proliferation were found between the 2 genotypes in either maternal condition (Figures S7A and S7B).

E12.5 eNOS$^{+/-}$ Hearts Display Increased Rates of Cellular Apoptosis Compared With eNOS$^{+/+}$ Hearts

Cellular apoptosis was assessed using CC3 immunostaining in E12.5 hearts. Representative images are shown in Figure 8A and 8B. Quantitative analysis showed that eNOS$^{+/-}$ hearts have a greater number of apoptotic cells per mm^2 than eNOS$^{+/+}$ hearts

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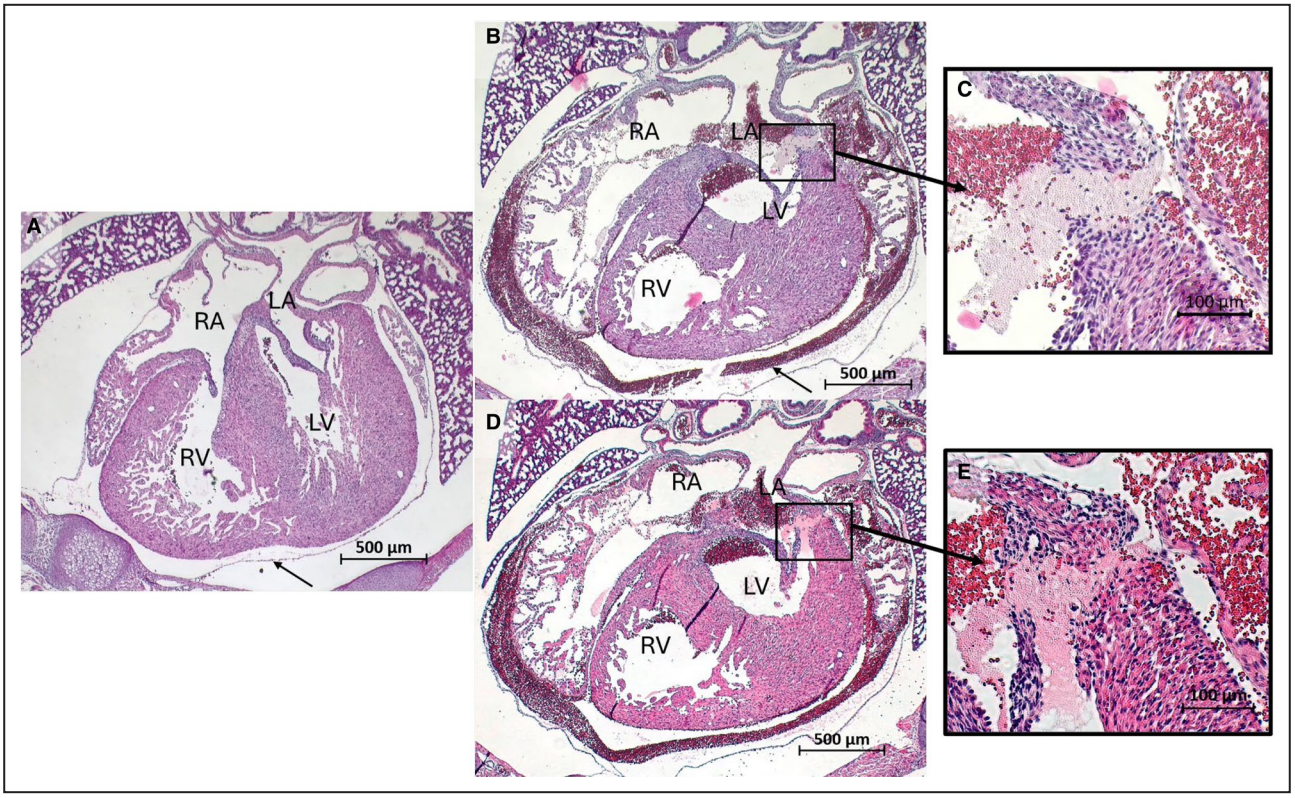


Figure 3. Cardiac rupture and hemopericardium CHD observed in an E18.5 eNOS$^{+/-}$ offspring of an eNOS$^{+/-}$ diabetic dam and maternal exercise.

A, Normal pericardium morphology represented by the arrow. **B** and **D**, Cardiac rupture at atrioventricular junction and blood-filled pericardium (hemopericardium) represented by the arrow. Scale bar represents 500 µm. **C** and **E** are enlarged areas of (**B**) and (**D**), respectively, showing rupture sites and thrombus formation. Scale bar represents 100 µm. **D** and **E** are 25 µm lower than images in (**B**) and (**C**). CHD indicates coronary heart defect; E, embryonic day; eNOS, endothelial nitric oxide synthase; LA, left atrium; LV, left ventricle; RA, right atrium; and RV, right ventricle.

(Figure 8C), suggesting that fetal eNOS heterozygosity increases basal apoptosis in E12.5 hearts.

Rates of Cellular Apoptosis Were Not Significantly Altered by PGD With or Without Maternal Exercise in E12.5 eNOS$^{+/+}$ and eNOS$^{+/-}$ Hearts

Fetal exposure to PGD did not significantly increase apoptosis rates in E12.5 hearts compared with control (Figure 8D through 8I). Additionally, maternal exercise in PGD dams did not alter apoptosis in eNOS$^{+/-}$ fetuses or combined eNOS$^{+/+}$ and eNOS$^{+/-}$ fetuses (Figure 8G and 8I) but significantly increased apoptosis in eNOS$^{+/+}$ fetuses (Figure 8H).

Differences in apoptosis were investigated between eNOS$^{+/+}$ and eNOS$^{+/-}$ E12.5 hearts exposed to PGD with and without exercise (Figure S8). The ratio of apoptotic cells to myocardial area (mm^2) was not significantly different between the 2 fetal genotypes. eNOS$^{+/+}$ mice exposed to PGD had a mean ratio of 51.0±19.3 apoptotic cells/mm^2 compared with a mean of 35.6±12.0 apoptotic cells/mm^2 in eNOS$^{+/-}$ hearts. The same is true in eNOS$^{+/+}$ and eNOS$^{+/-}$ mice exposed to PGD with maternal exercise, as they displayed means of 46.7±8.8 apoptotic cells/mm^2 and 49.0±10.6 apoptotic cells/mm^2, respectively. PGD exposure did not significantly alter apoptosis in eNOS$^{+/-}$ fetuses (Figure 8C).

The Impact of Fetal eNOS Heterozygosity, PGD, and Maternal Exercise on Markers of Oxidative Stress

Two markers of oxidative stress were evaluated: dihydroethidine and 4-HNE. Dihydroethidine is a probe for superoxide levels in tissues, whereas 4-HNE is a product of lipid peroxidation visualized using immunostaining. Fluorescence intensity was sampled 5 times at 3 separate locations in E12.5 hearts at a constant exposure (150 ms for 4-HNE, 90 ms for dihydroethidine) in each fetal group. Basal superoxide levels were similar between eNOS$^{+/+}$ and eNOS$^{+/-}$ fetal hearts (Figure 9A through 9C). PGD without and with maternal exercise significantly increased superoxide levels in E12.5 fetal hearts (Figure 10A through 10D). The same effect was observed in eNOS$^{+/+}$ and eNOS$^{+/-}$ fetuses when analyzed

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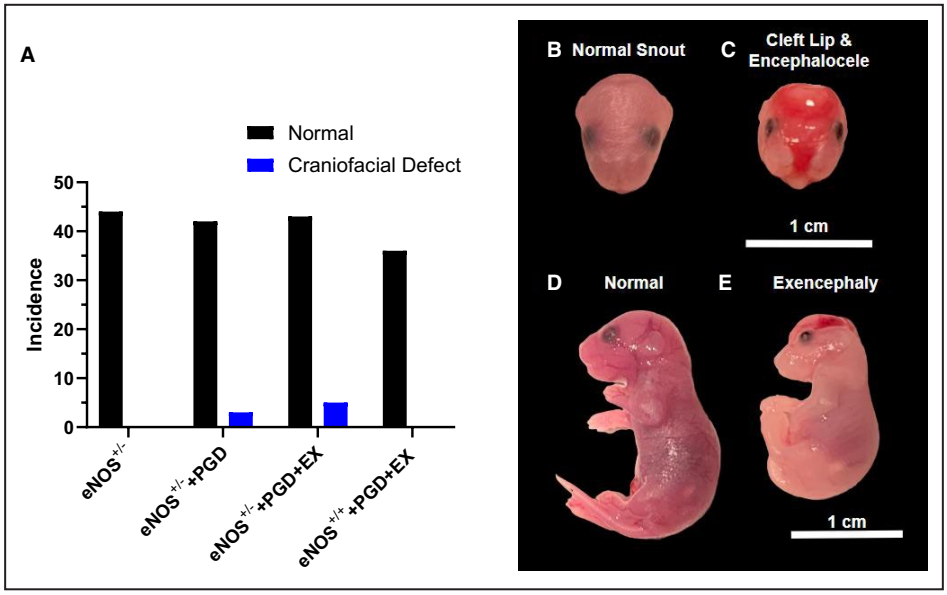


Figure 4. Craniofacial defect incidence and types observed in E18.5 eNOS$^{+/+}$ and eNOS$^{+/-}$ exposed to eNOS$^{+/-}$ dams with PGD with and without maternal exercise.
A, Incidence of craniofacial defects in offspring exposed to eNOS$^{+/-}$ control dams (n=44), eNOS$^{+/-}$+PGD (n=42), eNOS$^{+/-}$+PGD+exercise (n=48), and eNOS$^{+/+}$+PGD+exercise (n=36). eNOS$^{+/-}$ dams with PGD and maternal exercise produced offspring with the highest risk of craniofacial defect development. **B**, Normal facial and snout morphology. **C**, Cleft lip: failure of the skin to close over the mouth, Encephalocele: brain matter begins to grow outside the skull. **D**, The normal brain is encapsulated by meninges, skull and skin. **E**, Exencephaly: the brain is located entirely outside of the skull. There was no statistical significance by Fisher's exact test (P=0.0714). +/– indicates heterozygous; E, embryonic day; eNOS, endothelial nitric oxide synthase; EX, exercise; and PGD, pregestational diabetes.

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separately (Figure 10E through 10F). Additionally, no differences in superoxide levels were found between eNOS$^{+/+}$ and eNOS$^{+/-}$ fetuses exposed to PGD or PGD with maternal exercise (Figure S9).Lipid peroxidation was not significantly different between eNOS$^{+/+}$ and eNOS$^{+/-}$ hearts in the control condition (Figure 9D through 9F). Exposure to PGD with and without maternal exercise increased oxidative stress in the form of lipid peroxidation in eNOS$^{+/-}$ fetuses but not in eNOS$^{+/+}$ fetuses (Figure 10G through 10L). Lipid peroxidation levels were not significantly different between eNOS$^{+/+}$ and eNOS$^{+/-}$ hearts from PGD dams with and without exercise (Figure S10A and S10B).

To summarize, eNOS heterozygosity did not increase oxidative stress on its own. PGD without or with maternal exercise increases oxidative stress in the fetal heart. The observed increase in oxidative stress was not dependent on fetal eNOS heterozygosity.

eNOS Protein Levels Are Unaffected by Maternal Exercise During PGD in E12.5 Hearts

The eNOS$^{+/-}$ mouse model was validated by Western blot analysis assessing eNOS protein levels in control eNOS$^{+/+}$, eNOS$^{+/-}$, and eNOS$^{-/-}$ E12.5 hearts (Figure 11A). Protein levels were quantified using densitometry which revealed reduced eNOS protein levels in eNOS$^{+/-}$ hearts compared with eNOS$^{+/+}$ hearts and undetectable expression in eNOS$^{-/-}$ hearts (Figure 11B). Although eNOS$^{+/-}$ animals are deficient in 1 eNOS allele, protein levels in eNOS$^{+/-}$ hearts were found to be 60% of eNOS$^{+/+}$ rather than the expected 50%.

Similarly, eNOS protein levels in eNOS$^{+/-}$ fetal hearts of eNOS$^{+/-}$ dams exposed to PGD or PGD and maternal exercise were approximately 60% of the eNOS protein in the eNOS$^{+/+}$ hearts (Figure 11C and 11D). Maternal exercise in eNOS$^{+/-}$ dams during PGD did not significantly change eNOS expression in both eNOS$^{+/+}$ and eNOS$^{+/-}$ fetal hearts (Figure 11C and 11D).

Levels of eNOS serine 1177 phosphorylation were assessed using immunofluorescence in E12.5 hearts from eNOS$^{+/+}$ and eNOS$^{+/-}$ fetuses of nonexercising and exercising eNOS$^{+/-}$ dams with PGD (Figure 11E). eNOS phosphorylation was not affected by fetal genotype, as eNOS$^{+/+}$ and eNOS$^{+/-}$ hearts of the same condition displayed similar phosphorylated eNOS levels (Figure 11F). Furthermore, maternal exercise did not affect phospho-eNOS levels in the E12.5 hearts of eNOS$^{+/+}$ or eNOS$^{+/-}$ fetuses (Figure 11F).

Table 3. Craniofacial Defect Type and Incidence in Embryonic Day 18.5 eNOS$^{+/+}$ and eNOS$^{+/-}$ Fetuses Exposed to Diabetic Dams Without or With Maternal Exercise

Maternal condition	eNOS$^{+/-}$ (5 litters)		eNOS$^{+/-}$+PGD (7 litters)		eNOS$^{+/-}$+PGD+exercise (6 litters)		eNOS$^{+/+}$+PGD+exercise (5 litters)	
Offspring genotype	eNOS$^{+/+}$ (n=24)	eNOS$^{+/-}$ (n=20)	eNOS$^{+/+}$ (n=21)	eNOS$^{+/-}$ (n=21)	eNOS$^{+/+}$ (n=28)	eNOS$^{+/-}$ (n=22)	eNOS$^{+/+}$ (n=21)	eNOS$^{+/-}$ (n=14)
Normal	24 (100%)	20 (100%)	20 (95%)	19 (90%)	27 (96%)	18 (82%)	21 (100%)	14 (100%)
Abnormal	0	0	1 (5%)	2 (10%)	1 (4%)	4 (18%)	0	0
Exencephaly	0	0	1 (5%)	0	1 (4%)	4 (18%)	0	0
Encephalocele	0	0	0	1 (5%)	0	0	0	0
Cleft lip	0	0	0	1 (5%)	0	0	0	0

Data were analyzed using Fisher's exact test P=0.0714. The number of litters is equivalent to the number of dams used in each condition. +/– indicates heterozygous; eNOS, endothelial nitric oxide synthase; and PGD, pregestational diabetes.

DISCUSSION

Currently, 16.7 million women of reproductive age (20–49 years) were living with diabetes in 2021.[23] Risk factors for adverse fetal outcomes, such as PGD, can be reduced through effective interventions like glycemic control during pregnancy.[3] However, the incidence of CHDs related to maternal diabetes has only slightly decreased by 6% from 1997 to 2011 compared with earlier years, indicating a need for improved preventative strategies.[3] As diabetes prevalence continues to rise, establishing the mechanisms responsible for the benefits of maternal exercise is imperative to improve pre- and antenatal interventions. We aimed to establish a link between eNOS and the benefits of maternal exercise in mitigating CHDs in PGD.

This study demonstrated that maternal exercise did not reduce CHD incidence caused by PGD in an eNOS haploinsufficient environment. However, in WT (eNOS$^{+/+}$) mice, we have shown that maternal exercise significantly decreased PGD-induced CHD incidence from 59.5% to 25%.[10] Our findings show that eNOS haploinsufficiency limits the protective effects of maternal exercise during PGD. Interestingly, CHD incidence was similar in fetuses of eNOS$^{+/+}$ and eNOS$^{+/-}$ dams with PGD and exercise, indicating that maternal eNOS may not be the primary mediator of exercise benefits. Additionally, fetal eNOS heterozygosity did not increase the risk of CHD development during PGD regardless of maternal exercise (Table 2). Hoffman et al. established criteria to classify the severity of CHDs according to the complexity of the intervention required for lifetime management of the defect.[24] According to these criteria, atrial septal defect, ventricular septal defect, and valve defects (leaflet thickening, bicuspid aortic valve) are less severe than more complex defects like double outlet right ventricle and atrioventricular septal defect. Previous research showed that maternal exercise during PGD reduced the incidence of more severe CHDs, like double outlet right ventricle and atrioventricular septal defect.[10] However, the current study found no difference in the incidence of more severe to less severe defects across maternal conditions (Table S3). Furthermore, although prior studies have reported additional severe defects like hypoplastic heart syndrome,[10] these were not observed in this study. Differences in blood glucose levels between study cohorts throughout gestation may explain these discrepancies. Additionally, the fetal genotype did not influence the risk of developing CHDs, reinforcing the conclusion that fetal eNOS alone does not independently drive the benefits of maternal exercise.

Cardiac and vascular defects arise when the normal steps of cardiogenesis are impeded. E12.5 hearts exposed to PGD showed fewer pHH3 immunostained cells, indicating fewer cells undergoing proliferation. Maternal exercise restored cellular proliferation impaired by PGD to control levels in our study, regardless of fetal genotype. We have previously shown that PGD impairs cellular proliferation in fetal hearts, and maternal exercise restores cellular proliferation to control levels.[10] The improved cellular proliferation was attributed to increased expression of *Cyclin D1*, a marker of cell proliferation, and cardiogenic transcription factor *Gata4* that is critical for driving proliferation at key stages of cardiac development, such as septation.[10,25,26] In the present study, eNOS haploinsufficiency did not impede the maternal exercise benefits to cellular proliferation. Notably, maternal exercise has been shown to normalize the dysregulated micro-RNAs enriched in cell proliferation.[27] Our data suggest that maternal exercise may rescue cell proliferation in the fetal heart via eNOS independent signaling pathways, such as micro-RNA expression networks.

Maternal exercise during PGD did not alter eNOS protein levels in E12.5 fetal hearts. This aligns with findings in adult male eNOS$^{+/-}$ mice, where exercise did not change eNOS mRNA or protein levels in blood vessels, though eNOS expression was increased in the left ventricular myocardium.[28] In WT fetal hearts, maternal exercise during PGD did not significantly affect eNOS protein levels but did increase eNOS

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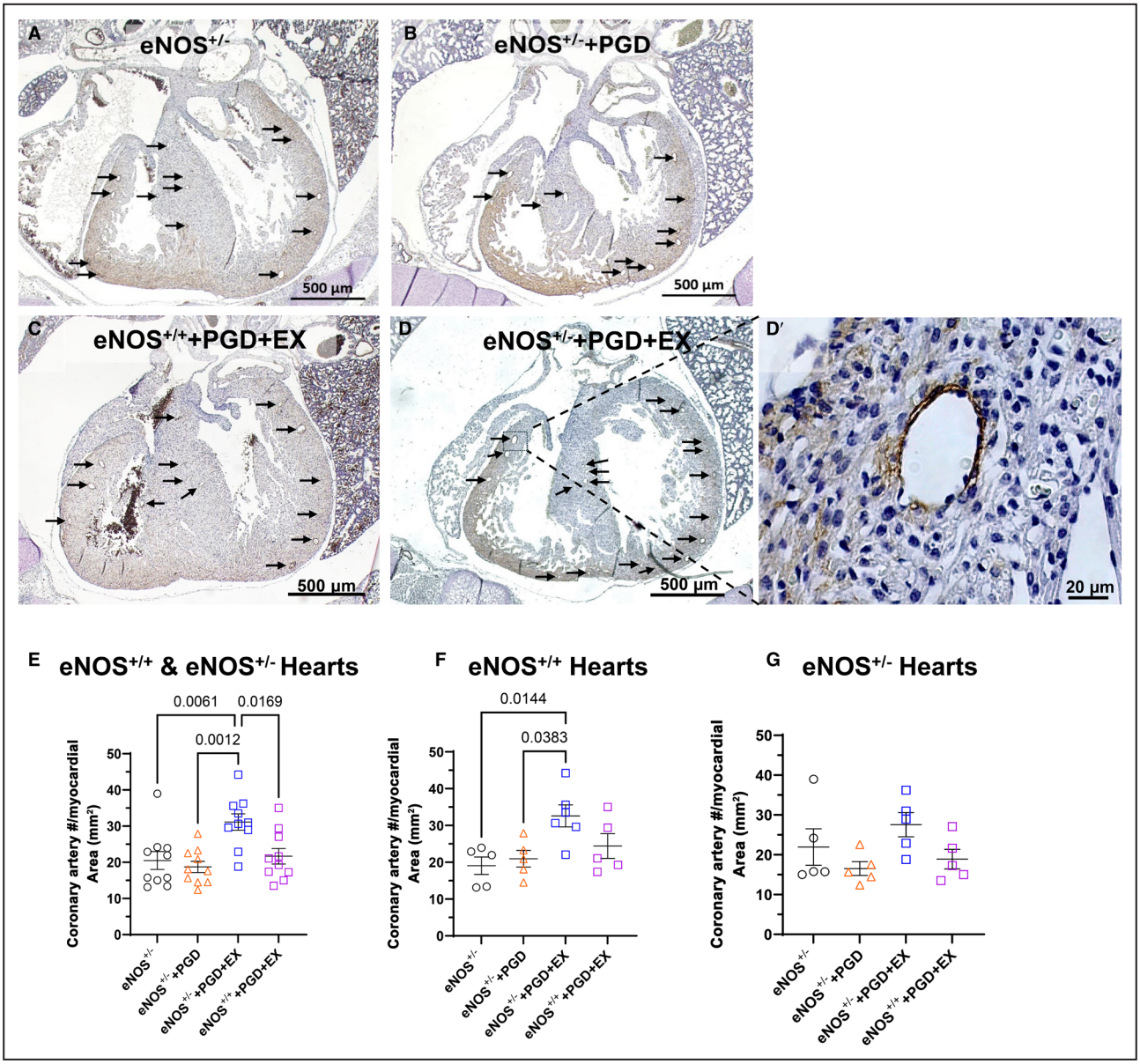


Figure 5. Maternal exercise in eNOS$^{+/-}$ dams with PGD improves coronary artery abundance in E18.5 eNOS$^{+/+}$ and eNOS$^{+/-}$ hearts.

A–D, Representative images of coronary arteries in E18.5 fetal hearts. Sections are immunostained with α-SMA to label coronary arteries as indicated by arrows. **D'** shows a coronary artery stained with α-SMA in the RV free wall of the heart. **E**, Average number of coronary arteries counted in 3 sections for eNOS$^{+/+}$ and eNOS$^{+/-}$ fetuses combined (n=10 hearts per group). **F**, Average coronary artery abundance in eNOS$^{+/+}$ fetuses only (n=5 hearts per group). **G**, The average coronary artery abundance in eNOS$^{+/-}$ fetuses (n=5 hearts per group). Vessels were deemed coronary arteries with a diameter 15–60 µm. Data represent means±SEM and were analyzed using 1-way ANOVA followed by Tukey's test. +/– indicates heterozygous; α-SMA, α-smooth muscle actin; E, embryonic day; eNOS, endothelial nitric oxide synthase; EX, exercise; PGD, pregestational diabetes; and RV, right ventricle.

phosphorylation.[10] These data suggest that the protective effects of maternal exercise during PGD are not driven by an overall increase in eNOS protein expression. Notably, eNOS phosphorylation at serine 1177 increases its activity. In the present study, maternal exercise did not increase eNOS phosphorylation in E12.5 hearts of either eNOS$^{+/+}$ or eNOS$^{+/-}$ fetuses from diabetic eNOS$^{+/-}$ dams. The low levels of eNOS phosphorylation in these fetal groups are consistent with their high incidences of CHDs with and without maternal exercise. The results strongly support a critical role of eNOS in mitigating CHD risk during PGD by maternal exercise.

A major contributor to CHD pathogenesis is oxidative stress.[9,29,30] Our study found that oxidative stress levels were elevated in E12.5 fetal hearts exposed to

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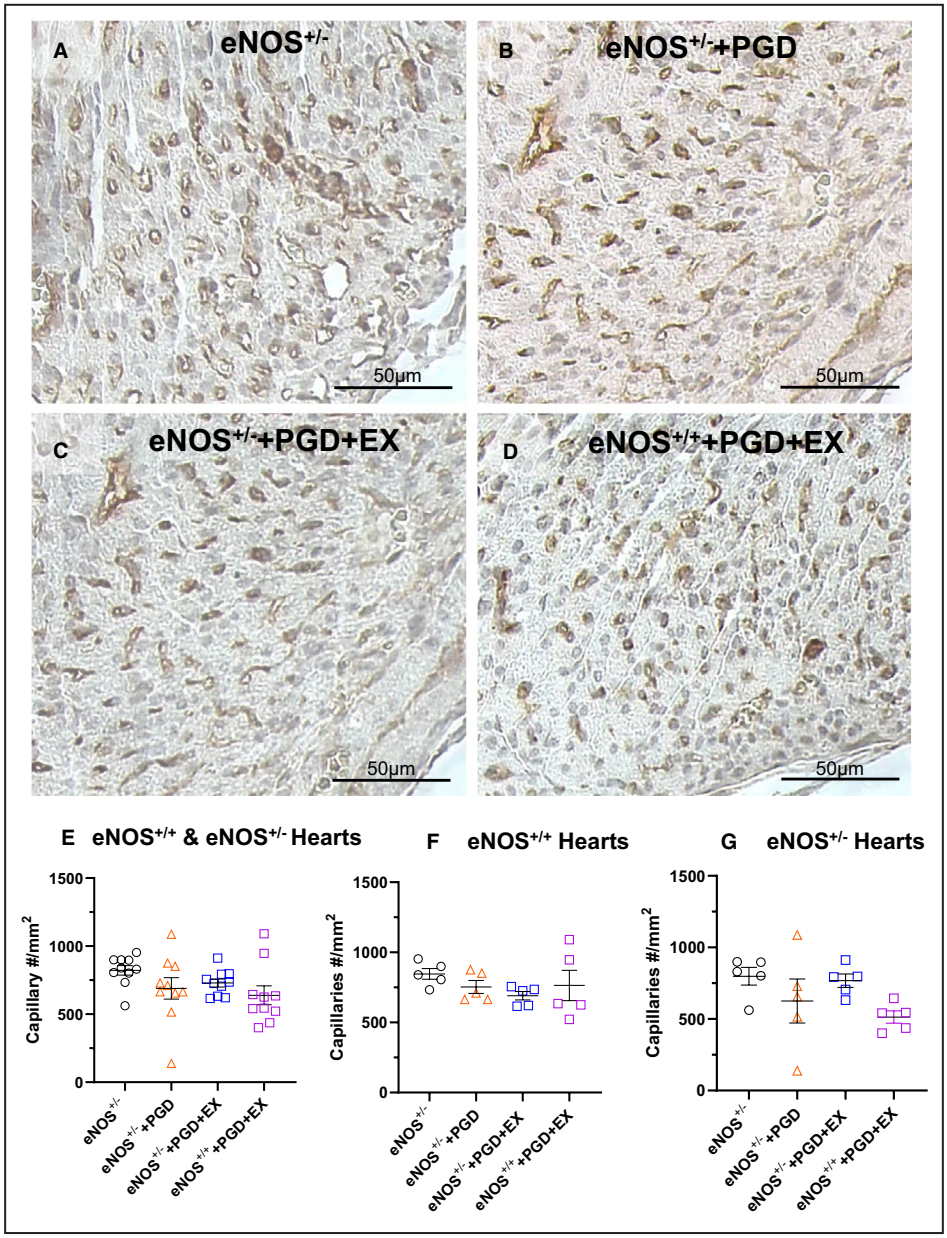


Figure 6. Maternal exercise in eNOS$^{+/+}$ and eNOS$^{+/-}$ dams with PGD does not alter capillary density in E18.5 eNOS$^{+/+}$ and eNOS$^{+/-}$ hearts.
A–D, Representative images of ventricular capillaries at the 4-chamber level of E18.5 fetal hearts. Sections are immunostained with lectin-1 to label endothelial cells. **E**, Average number of capillaries counted in 3 sections per ventricular structure for eNOS$^{+/+}$ and eNOS$^{+/-}$ fetuses combined (n=10 hearts per group). **F**, Average capillary density in eNOS$^{+/+}$ fetuses only (n=5 hearts per group). **G**, Average capillary density in eNOS$^{+/-}$ fetuses only (n=5 hearts per group). Data represent means±SEM and were analyzed using 1-way ANOVA followed by Tukey's test. +/– indicates heterozygous; E, embryonic day; eNOS, endothelial nitric oxide synthase; EX, exercise; and PGD, pregestational diabetes.

PGD, consistent with previous research.[7,10,31] This increase was evident through higher superoxide and lipid peroxidation levels. However, maternal exercise did not reduce oxidative stress in E12.5 eNOS$^{+/+}$ or eNOS$^{+/-}$ hearts of fetuses from eNOS$^{+/-}$ dams during PGD. Previously, we demonstrated that maternal exercise lowers superoxide and lipid peroxidation levels in eNOS$^{+/+}$ fetal hearts during PGD.[10] This discrepancy in our current findings may be due to reduced NO availability caused by eNOS heterozygosity, which compromises its antioxidant properties. NO can efficiently transfer from maternal to fetal circulation via

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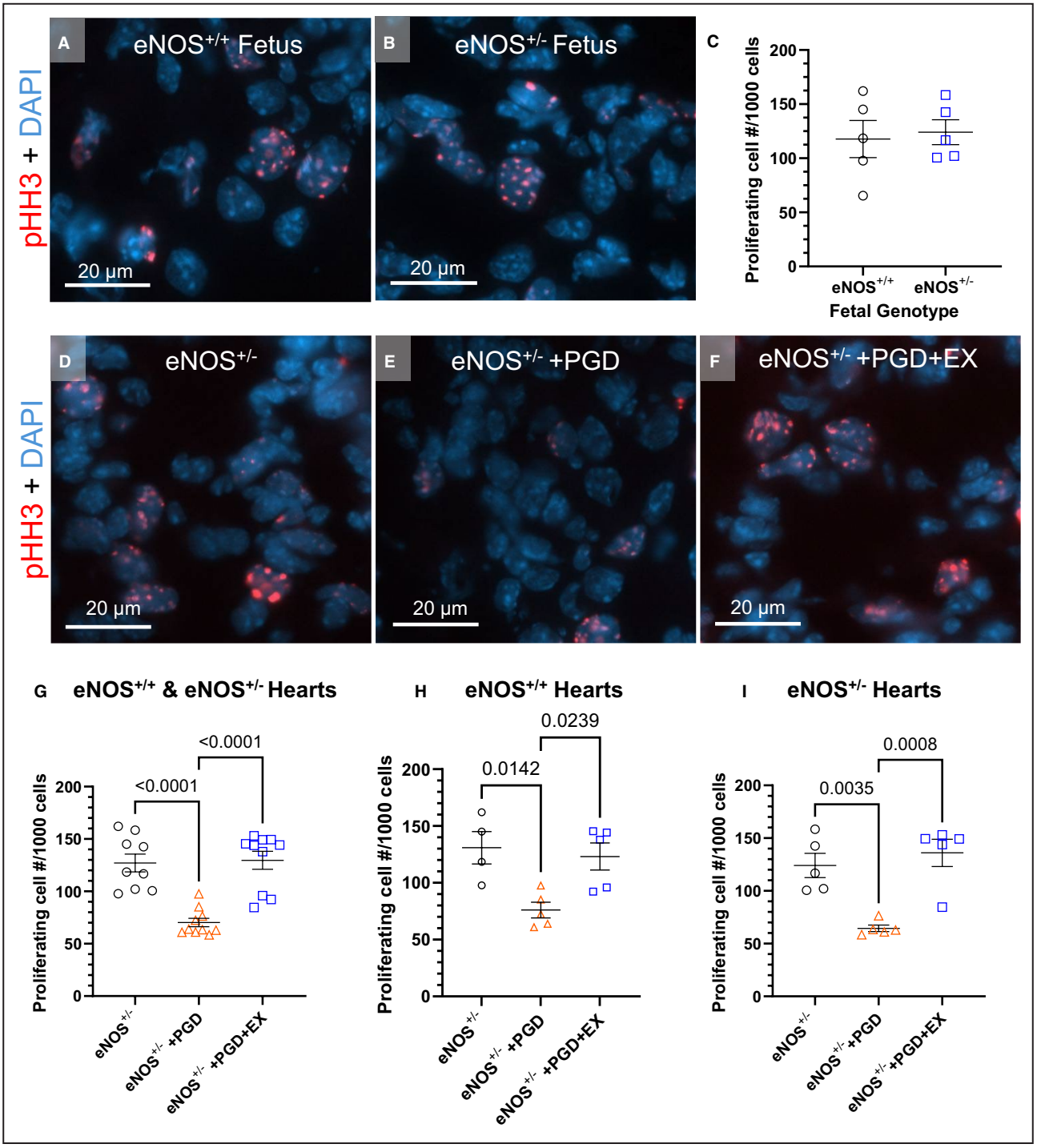


Figure 7. Maternal exercise increased cellular proliferation in E12.5 eNOS$^{+/+}$ and eNOS$^{+/-}$ hearts exposed to PGD.
A, **B**, Representative images of pHH3 immunofluorescence in eNOS$^{+/+}$ and eNOS$^{+/-}$ hearts of E12.5 fetuses. **C**, Rates of cellular proliferation per 1000 cells were not significantly different between eNOS$^{+/+}$ and eNOS$^{+/-}$ fetuses. **D–F**, Representative images of pHH3 immunofluorescence in E12.5 fetal hearts from eNOS$^{+/-}$ control, eNOS$^{+/-}$+PGD, and eNOS$^{+/-}$+PGD+exercise dams. **G**, Rates of proliferation per 1000 cells were reduced by PGD and rescued by maternal exercise in eNOS$^{+/+}$ and eNOS$^{+/-}$ fetal hearts. The same trend is observed in eNOS$^{+/+}$ hearts (**H**) and eNOS$^{+/-}$ hearts (**I**). Data represent mean±SEM of n=4 to 10 hearts per group and were analyzed using unpaired Student's *t* test (**C**) or 1-way ANOVA followed by Tukey's test (**G–I**). +/– indicates heterozygous; E, embryonic day; eNOS, endothelial nitric oxide synthase; EX, exercise; PGD, pregestational diabetes; and pHH3, phosphorylated histone H3.

the placenta; a shared pool of NO likely exists regardless of fetal genotype. Exercise has been shown to increase NO production in WT adults but not in eNOS$^{+/-}$ animals.[19,28] NO mitigates Fenton-mediated oxidative stress through several mechanisms, including direct scavenging, inhibition of peroxide reactions,

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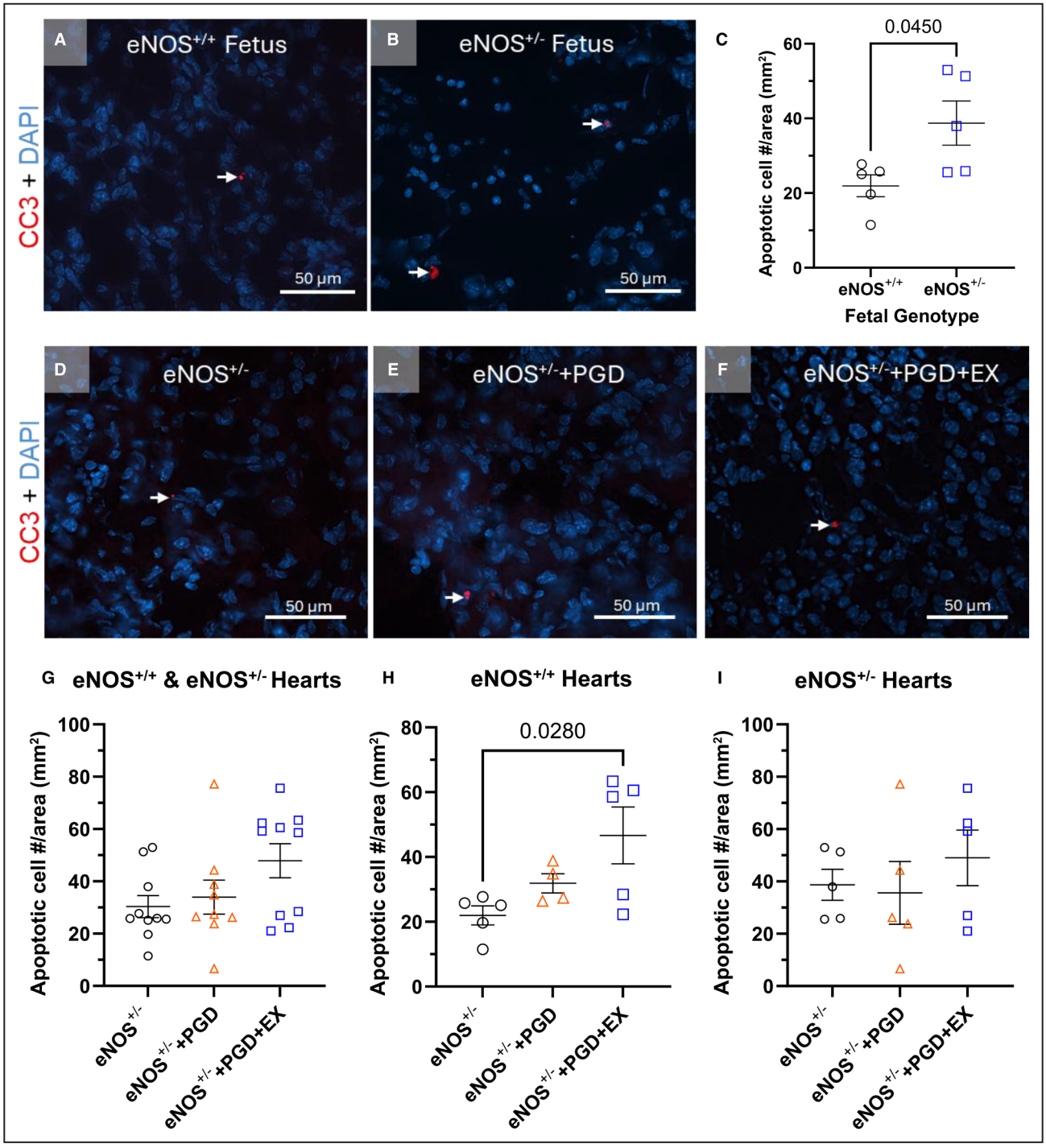


Figure 8. Rates of apoptosis in E12.5 fetal hearts from eNOS$^{+/-}$ dams without and with PGD and maternal exercise.
A, **B**, Representative images of cleaved caspase-3 immunostaining in E12.5 eNOS$^{+/+}$ and eNOS$^{+/-}$ hearts. **C**, Ratio of apoptotic cells to heart area in fetal eNOS$^{+/+}$ and eNOS$^{+/-}$ hearts of eNOS$^{+/-}$ dams. **D**–**F**, Representative images of cleaved caspase-3 immunostaining in E12.5 eNOS$^{+/+}$ and eNOS$^{+/-}$ hearts from eNOS$^{+/-}$ control, eNOS$^{+/-}$+PGD, and eNOS$^{+/-}$+PGD+exercise dams. **G**, The ratio of apoptotic cells to heart area of eNOS$^{+/+}$ and eNOS$^{+/-}$ fetal hearts. **H**, eNOS$^{+/+}$ fetuses of diabetic dams with exercise showed elevated rates of apoptosis. **I**, The ratio of apoptotic cells to heart area of eNOS$^{+/-}$ fetal hearts. Data represent mean±SEM of n=4 to 10 hearts per group and were analyzed using unpaired Student's *t* test (**C**) or 1-way ANOVA followed Tukey's test (**F**). +/– indicates heterozygous; CC3, cleaved caspase-3; E, embryonic day; eNOS, endothelial nitric oxide synthase; EX, exercise; and PGD, pregestational diabetes.

and neutralization of superoxide. For example, NO can bind to ferrous iron in softer ligand fields, such as heme complexes, forming a metal-nitrosyl complex that prevents peroxide-iron interactions and ROS production. In addition, NO reacts with superoxide to form peroxynitrite, which is rapidly converted to nitrate,

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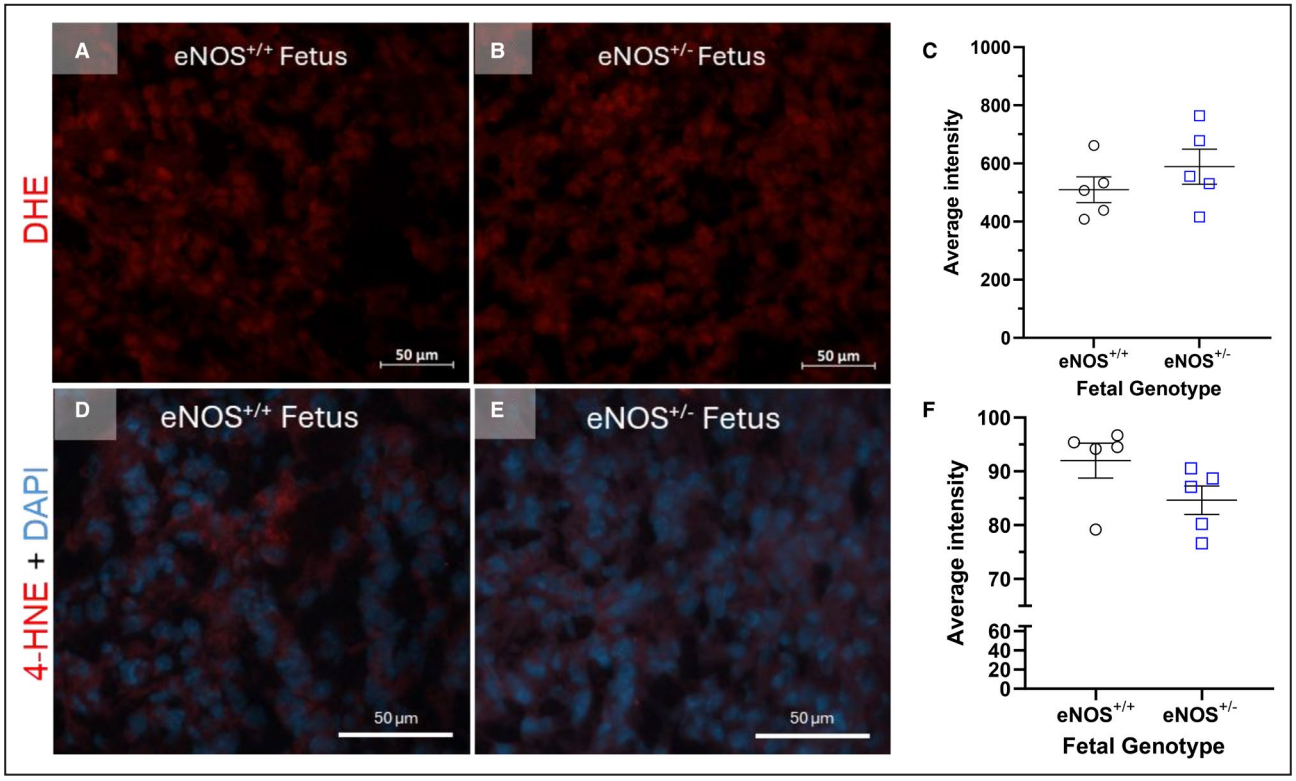


Figure 9. Oxidative stress levels were similar in E12.5 eNOS$^{+/+}$ and eNOS$^{+/-}$ hearts.
A, **B**, Representative images of DHE staining in eNOS$^{+/+}$ (**A**) and eNOS$^{+/-}$ (**B**) fetal hearts of eNOS$^{+/-}$ dams. **C**, DHE fluorescence intensity of eNOS$^{+/+}$ and eNOS$^{+/-}$ fetal hearts. **D**, **E**, Representative images of 4-HNE immunostaining with nuclear DAPI in eNOS$^{+/+}$ (**D**) and eNOS$^{+/-}$ (**E**) fetal hearts of eNOS$^{+/-}$ dams. **F**, 4-HNE fluorescence intensity in eNOS$^{+/+}$ and eNOS$^{+/-}$ fetal hearts. Data represent mean±SEM of n=5 hearts per group and were analyzed using unpaired Student's *t* test. +/– indicates heterozygous; 4-HNE, 4-hydroxynonenal; DHE, dihydroethidine; E, embryonic day; eNOS, endothelial nitric oxide synthase; EX, exercise; and PGD, pregestational diabetes.

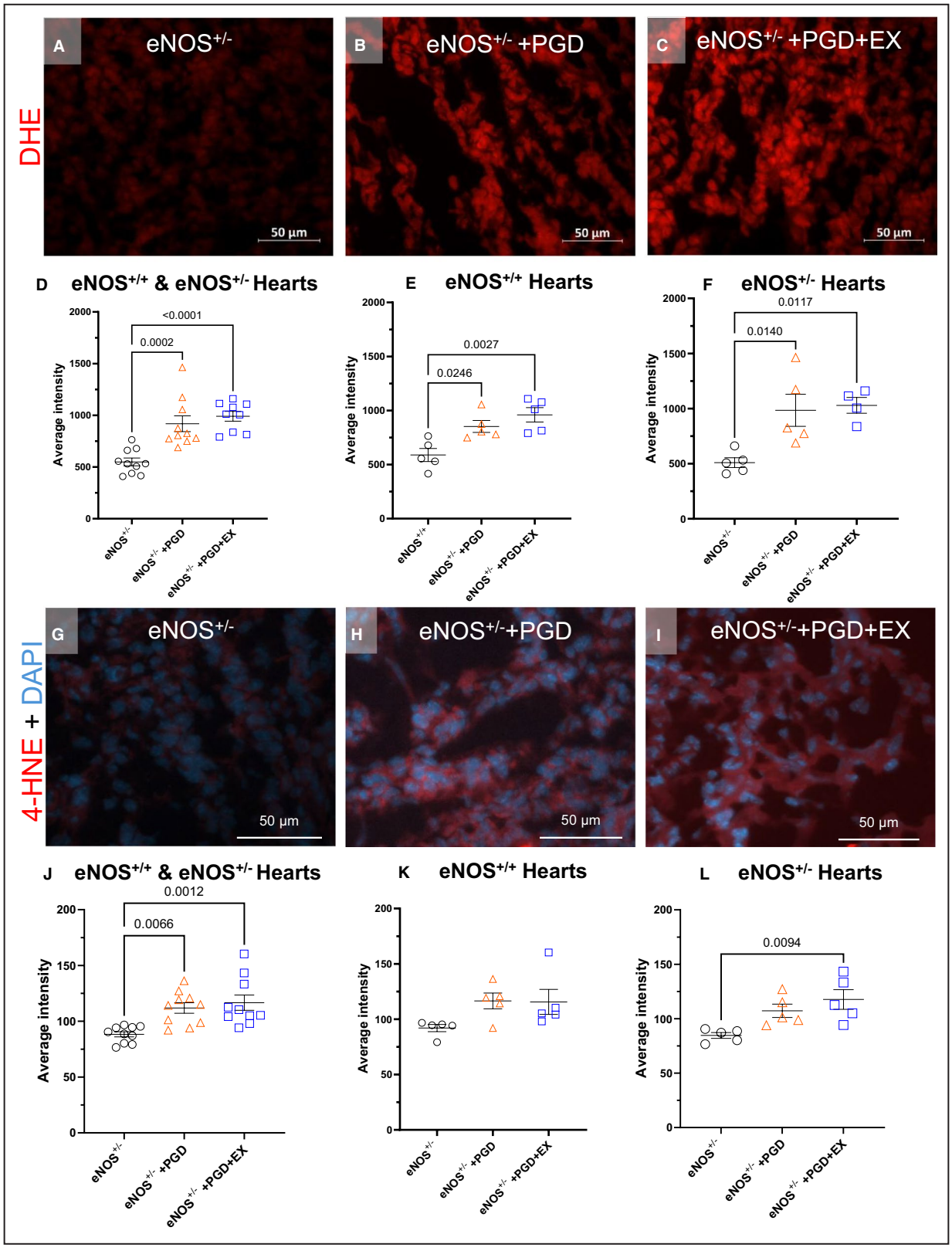
reducing ferric iron (Fe^{3+}) and halting ROS formation.[32] In the present study, a lower NO level in eNOS$^{+/-}$ dams may have been insufficient to counteract ROS production induced by PGD.[6,33]

In adults, exercise increases ROS production, which paradoxically enhances redox capacity by upregulating antioxidant enzymes such as SOD (superoxide dismutase), catalase, and glutathione peroxidase in cardiac and skeletal muscles. This adaptive response strengthens antioxidant defenses, protecting against ischemia–reperfusion injury.[34,35] Additionally, eNOS-derived NO plays a crucial role in downstream signaling, influencing gene expression.[36] Fukai et al. reported that exercise-induced eNOS activation correlates with increased SOD expression in adult vasculature, leading to a subsequent reduction in ROS.[37] Recent studies in humans have revealed that a specific isoform of SOD, SOD3, is upregulated by maternal exercise in a type 2 diabetes model in the placenta, which subsequently leads to activation of the AMPK/TET pathway in the fetal liver.[38] Furthermore, maternal exercise induces epigenetic modifications in utero, leading to long-term improvements in fetal liver function.[38] These findings highlight the need for further research into crosstalk between the placenta and fetal organs to better understand how maternal exercise promotes fetal heart development.

eNOS-derived NO regulates uteroplacental perfusion, endothelial function, and shear-stress signaling.[15,39]

Figure 10. Oxidative stress was elevated by PGD and unchanged by maternal exercise in E12.5 eNOS$^{+/+}$ and eNOS$^{+/-}$ hearts.
A–**C**, Representative images of DHE staining in fetal hearts exposed to eNOS$^{+/-}$ dams (**A**), eNOS$^{+/-}$ dams with PGD (**B**), and eNOS$^{+/-}$ dams with PGD and maternal exercise (**C**). **D**, DHE fluorescence intensity in eNOS$^{+/+}$ and eNOS$^{+/-}$ fetal hearts. **E**, DHE fluorescence intensity in eNOS$^{+/+}$ fetal hearts. **F**, DHE fluorescence intensity in eNOS$^{+/-}$ fetal hearts. **G**–**I**, Representative images of 4-HNE immunostaining and DAPI nuclear stain in fetal hearts exposed to eNOS$^{+/-}$ dams (**G**), eNOS$^{+/-}$ dams with PGD (**H**), and eNOS$^{+/-}$ dams with PGD and maternal exercise (**I**). **J**, 4-HNE fluorescence intensity in eNOS$^{+/+}$ and eNOS$^{+/-}$ fetal hearts. **K**, 4-HNE fluorescence intensity in eNOS$^{+/+}$ fetal hearts. **L**, 4-HNE fluorescence intensity in eNOS$^{+/-}$ fetal hearts. Data represent mean±SEM of n=5 to 10 hearts per group and were analyzed using 1-way ANOVA followed by Tukey's test. +/– indicates heterozygous; 4-HNE, 4-hydroxynonenal; DHE, dihydroethidine; E, embryonic day; eNOS, endothelial nitric oxide synthase; EX, exercise; and PGD, pregestational diabetes.

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Studies have shown that uteroplacental blood flow is significantly reduced in eNOS$^{+/-}$ mice.[40] Systemic NO levels, as assessed by nitrate/nitrite, are not significantly different between eNOS$^{-/-}$ and WT mice.[41] To our knowledge, there is still no report on circulating NO levels in eNOS$^{+/-}$ mice. Because systemic NO levels are maintained in eNOS$^{-/-}$ mice, it is unlikely that they are significantly affected in eNOS$^{+/-}$ mice. Whether maternal

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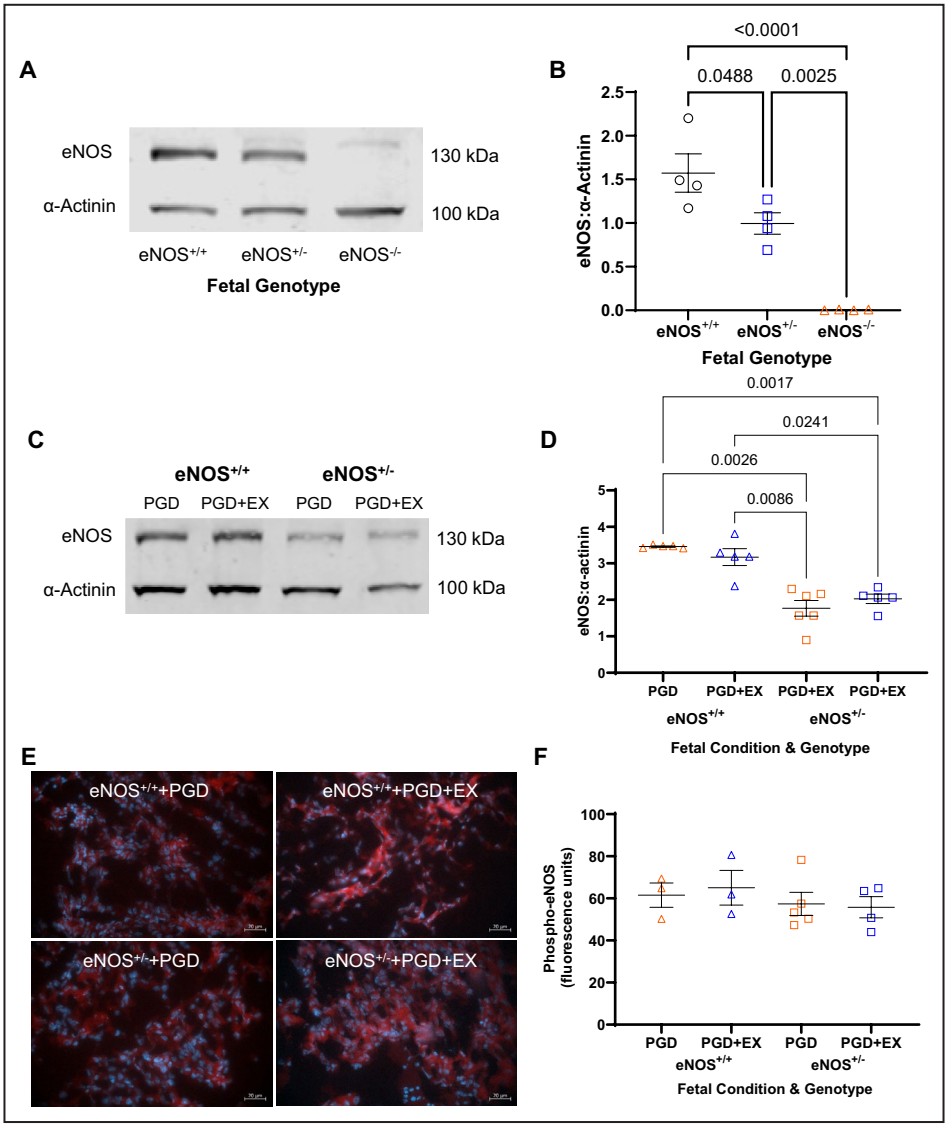


Figure 11. eNOS protein levels in E12.5 fetal hearts of control dams and dams exposed to PGD and maternal exercise.

A, Representative Western blot depicting eNOS protein levels in eNOS$^{+/+}$, eNOS$^{+/-}$, and eNOS$^{-/-}$ E12.5 hearts of control dams. **B**, Densitometry quantification of the ratio of eNOS to α-actinin. **C**, Representative protein levels from eNOS$^{+/+}$ or eNOS$^{+/-}$ fetal hearts of dams exposed to PGD without and with maternal exercise. **D**, Densitometry quantification of the ratio of eNOS to α-actinin. **E** and **F**, Immunostaining and quantification of eNOS serine 1177 phosphorylation. Data represent mean±SEM of n=3 to 6 hearts per group and were analyzed using 1-way ANOVA followed by Tukey's test. +/– indicates heterozygous; E, embryonic day; eNOS, endothelial nitric oxide synthase; EX, exercise; and PGD, pregestational diabetes.

exercise can increase uteroplacental blood flow and systemic NO levels in eNOS$^{+/-}$ dams with PGD requires further investigation.

PGD perturbs endothelial function and NO signaling, and increases vulnerability to CHDs during early organogenesis.[15,39] In our murine model, eNOS is identified as a requisite for the cardioprotective effects of maternal exercise, which augments shear stress and restores NO tone.[39] As this pathway is conserved in humans, these findings support early initiation of moderate physical activity and point to pharmacological enhancement of eNOS activity or NO bioavailability as potential strategies to reduce CHD risk in pregestational diabetes.

CONCLUSIONS

In summary, maternal exercise during PGD in eNOS$^{+/-}$ dams does not alleviate PGD-induced oxidative stress or the incidence of CHDs in fetuses. Our findings

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suggest that while maternal exercise can influence fetal cardiac oxidative stress in utero, this effect depends on eNOS. Because eNOS haploinsufficiency diminishes the benefits of maternal exercise on fetal heart development, our study highlights a critical role of eNOS in mitigating CHD risk during PGD by maternal exercise.

ARTICLE INFORMATION

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Acknowledgments

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Disclosures

None.

Supplemental Material

Data S1.
Data S2.

REFERENCES

1. Yamagishi H. Clinical developmental cardiology for understanding etiology of congenital heart disease. *J Clin Med*. 2022;11:2381. doi: 10.3390/jcm11092381
2. Yasuhara J, Garg V. Genetics of congenital heart disease: a narrative review of recent advances and clinical implications. *Transl Pediatr*. 2021;10:2366–2386. doi: 10.21037/tp-21-297
3. Øyen N, Diaz LJ, Leirgul E, Boyd HA, Priest J, Mathiesen ER, Quertermous T, Wohlfahrt J, Melbye M. Prepregnancy diabetes and offspring risk of congenital heart disease. *Circulation*. 2016;133:2243–2253. doi: 10.1161/CIRCULATIONAHA.115.017465
4. International Diabetes Federation. Diabetes Facts & figures [Internet]. 2024 Accessed Jun 10, 2024. https://idf.org/about-diabetes/diabetes-facts-figures/.
5. Liu Y, Chen S, Zühlke L, Black GC, Choy M, Li N, Keavney BD. Global birth prevalence of congenital heart defects 1970–2017: updated systematic review and meta-analysis of 260 studies. *Int J Epidemiol*. 2019;48:455–463. doi: 10.1093/ije/dyz009
6. Basu M, Garg V. Maternal hyperglycemia and fetal cardiac development: clinical impact and underlying mechanisms. *Birth Defects Res*. 2018;110:1504–1516. doi: 10.1002/bdr2.1435
7. Moazzen H, Lu X, Ma NL, Velenosi TJ, Urquhart BL, Wisse LJ, Gittenberger-de Groot AC, Feng Q. N-Acetylcysteine prevents congenital heart defects induced by pregestational diabetes. *Cardiovasc Diabetol*. 2014;13:46. doi: 10.1186/1475-2840-13-46
8. Morgan SC, Relaix F, Sandell LL, Loeken MR. Oxidative stress during diabetic pregnancy disrupts cardiac neural crest migration and causes outflow tract defects. *Birth Defects Res A Clin Mol Teratol*. 2008;82:453–463. doi: 10.1002/bdra.20457
9. Wang F, Fisher SA, Zhong J, Wu Y, Yang P. Superoxide dismutase 1 in vivo ameliorates maternal diabetes-induced apoptosis and heart defects through restoration of impaired Wnt signaling. *Circ Cardiovasc Genet*. 2015;8:665–676. doi: 10.1161/CIRCGENETICS.115.001138
10. Saiyin T, Engineer A, Greco ER, Kim MY, Lu X, Jones DL, Feng Q. Maternal voluntary exercise mitigates oxidative stress and incidence of congenital heart defects in pre-gestational diabetes. *J Cell Mol Med*. 2019;23:5553–5565. doi: 10.1111/jcmm.14439
11. Feng Q, Song W, Lu X, Hamilton JA, Lei M, Peng T, Yee S-P. Development of heart failure and congenital septal defects in mice lacking endothelial nitric oxide synthase. *Circulation*. 2002;106:873–879. doi: 10.1161/01.CIR.0000024114.82981.EA
12. Teichert A-M, Scott JA, Robb GB, Zhou Y-Q, Zhu S-N, Lem M, Keightley A, Steer BM, Schuh AC, Adamson SL, et al. Endothelial nitric oxide synthase gene expression during murine embryogenesis: commencement of expression in the embryo occurs with the establishment of a unidirectional circulatory system. *Circ Res*. 2008;103:24–33. doi: 10.1161/CIRCRESAHA.107.168567
13. Wei C-M, Jiang S-W, Lust JA, Daly RC, McGregor CGA. Genetic expression of endothelial nitric oxide synthase in human atrial myocardium. *Mayo Clin Proc*. 1996;71:346–350. doi: 10.4065/71.4.346
14. Feron O, Belhassen L, Kobzik L, Smith TW, Kelly RA, Michel T. Endothelial nitric oxide synthase targeting to Caveolae: specific interactions with caveolin isoforms in cardiac myocytes and endothelial cells. *J Biol Chem*. 1996;271:22810–22814. doi: 10.1074/jbc.271.37.22810
15. Liu Y, Feng Q. NOing the heart: role of nitric oxide synthase-3 in heart development. *Differentiation*. 2012;84:54–61. doi: 10.1016/j.diff.2012.04.004
16. Peterson JC, Chughtai M, Wisse LJ, de Gittenberger- Groot AC, Feng Q, Goumans M-JTH, VanMunsteren JC, Jongbloed MRM, DeRuiter MC. Bicuspid aortic valve formation: Nos3 mutation leads to abnormal lineage patterning of neural crest cells and the second heart field. *Dis Model Mech*. 2018;11:dmm034637. doi: 10.1242/dmm.034637
17. Peterson JC, Wisse LJ, Wirokromo V, van Herwaarden T, Smits AM, de Gittenberger- Groot AC, Goumans M-JTH, VanMunsteren JC, Jongbloed MRM, DeRuiter MC. Disturbed nitric oxide signalling gives rise to congenital bicuspid aortic valve and aortopathy. *Dis Model Mech*. 2020;13:dmm044990. doi: 10.1242/dmm.044990
18. Lee TC, Zhao YD, Courtman DW, Stewart DJ. Abnormal aortic valve development in mice lacking endothelial nitric oxide synthase. *Circulation*. 2000;101:2345–2348. doi: 10.1161/01.CIR.101.20.2345
19. Arefirad T, Seif E, Sepidarkish M, Mohammadian Khonsari N, Mousavifar SA, Yazdani S, Rahimi F, Einollahi F, Heshmati J, Qorbani M. Effect of exercise training on nitric oxide and nitrate/nitrite (NOx) production: a systematic review and meta-analysis. *Front Physiol*. 2022;13:953912. doi: 10.3389/fphys.2022.953912
20. Liu Y, Lu X, Xiang F-L, Poelmann RE, Gittenberger-de Groot AC, Robbins J, Feng Q. Nitric oxide synthase-3 deficiency results in hypoplastic coronary arteries and postnatal myocardial infarction. *Eur Heart J*. 2014;35:920–931. doi: 10.1093/eurheartj/ehs306
21. Moazzen H, Lu X, Liu M, Feng Q. Pregestational diabetes induces fetal coronary artery malformation via reactive oxygen species signaling. *Diabetes*. 2015;64:1431–1443. doi: 10.2337/db14-0190
22. Engineer A, Saiyin T, Lu X, Kucey AS, Urquhart BL, Drysdale TA, Norozi K, Feng Q. Sapropterin treatment prevents congenital heart defects induced by Pregestational diabetes mellitus in mice. *J Am Heart Assoc*. 2018;7:e009624. doi: 10.1161/JAHA.118.009624
23. International Diabetes Foundation. IDF Diabetes Atlas 2021 | IDF Diabetes Atlas [Internet]. 2021 Accessed Dec 5, 2024. https://diabetesatlas.org/atlas/tenth-edition/.
24. Hoffman JIE, Kaplan S, Liberthson RR. Prevalence of congenital heart disease. *Am Heart J*. 2004;147:425–439. doi: 10.1016/j.ahj.2003.05.003
25. Zhou L, Liu J, Xiang M, Olson P, Guzzetta A, Zhang K, Moskowitz IP, Xie L. Gata4 potentiates second heart field proliferation and

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hedgehog signaling for cardiac septation. *Proc Natl Acad Sci USA*. 2017;114:E1422–E1431. doi: 10.1073/pnas.1605137114
26. Yang K, Hitomi M, Stacey DW. Variations in cyclin D1 levels through the cell cycle determine the proliferative fate of a cell. *Cell Div*. 2006;1:32. doi: 10.1186/1747-1028-1-32
27. Ye F, Lu X, van Neck R, Jones DL, Feng Q. Novel circRNA-miRNA-mRNA networks regulated by maternal exercise in fetal hearts of pregestational diabetes. *Life Sci*. 2023;314:121308. doi: 10.1016/j.lfs.2022.121308
28. Kojda G, Cheng YC, Burchfield J, Harrison DG. Dysfunctional regulation of endothelial nitric oxide synthase (eNOS) expression in response to exercise in mice lacking one eNOS gene. *Circulation*. 2001;103:2839–2844. doi: 10.1161/01.CIR.103.23.2839
29. Sivan E, Lee YC, Wu YK, Reece EA. Free radical scavenging enzymes in fetal dysmorphogenesis among offspring of diabetic rats. *Teratology*. 1997;56:343–349. doi: 10.1002/(SICI)1096-9926(199712)56:6<343::AID-TERA1>3.0.CO;2-X
30. Laforgia N, Di Mauro A, Favia Guarnieri G, Varvara D, De Cosmo L, Panza R, Capozza M, Baldassarre ME, Resta N. The role of oxidative stress in the Pathomechanism of congenital malformations. *Oxidative Med Cell Longev*. 2018;2018:7404082. doi: 10.1155/2018/7404082
31. Moazzen H, Wu Y, Engineer A, Lu X, Aulakh S, Feng Q. NOX2 is critical to endocardial to mesenchymal transition and heart development. *Oxidative Med Cell Longev*. 2020;2020:1679045. doi: 10.1155/2020/1679045
32. Wink D, Miranda K, Espey M, Pluta R, Hewett S, Colton C, Vitek M, Feelisch M, Grisham M. Mechanisms of the antioxidant effects of nitric oxide. *Antioxid Redox Signal*. 2001;3:203–213. doi: 10.1089/152308601300185179
33. Basu M, Zhu J-Y, LaHaye S, Majumdar U, Jiao K, Han Z, Garg V. Epigenetic mechanisms underlying maternal diabetes-associated risk of congenital heart disease. *JCI Insight*. 2017;2:e95085. doi: 10.1172/jci.insight.95085
34. Wang F, Wang X, Liu Y, Zhang Z. Effects of exercise-induced ROS on the pathophysiological functions of skeletal muscle. *Oxidative Med Cell Longev*. 2021;2021:3846122. doi: 10.1155/2021/3846122
35. Königstein K, Dipla K, Zafeiridis A. Training the vessels: molecular and clinical effects of exercise on vascular health—a narrative review. *Cells*. 2023;12:2544. doi: 10.3390/cells12212544
36. Hofseth LJ, Robles AI, Espey MG, Harris CC. Nitric oxide is a signaling molecule that regulates gene expression. *Methods Enzymol*. 2005;396:326–340. doi: 10.1016/S0076-6879(05)96027-8
37. Fukai T, Siegfried MR, Ushio-Fukai M, Cheng Y, Kojda G, Harrison DG. Regulation of the vascular extracellular superoxide dismutase by nitric oxide and exercise training. *J Clin Invest*. 2000;105:1631–1639. doi: 10.1172/JCI9551
38. Kusuyama J, Alves-Wagner AB, Conlin RH, Makarewicz NS, Albertson BG, Prince NB, Kobayashi S, Kozuka C, Møller M, Bjerre M, et al. Placental superoxide dismutase 3 mediates benefits of maternal exercise on offspring health. *Cell Metab*. 2021;33:939–956.e8. doi: 10.1016/j.cmet.2021.03.004
39. Engineer A, Saiyin T, Greco ER, Feng Q. Say NO to ROS: their roles in embryonic heart development and pathogenesis of congenital heart defects in maternal diabetes. *Antioxidants*. 2019;8:436. doi: 10.3390/antiox8100436
40. Kulandavelu S, Whiteley KJ, Qu D, Mu J, Bainbridge SA, Adamson SL. Endothelial nitric oxide synthase deficiency reduces uterine blood flow, spiral artery elongation, and placental oxygenation in pregnant mice. *Hypertension*. 2012;60:231–238. doi: 10.1161/HYPERTENSIONAHA.111.187559
41. Peleli M, Zollbrecht C, Montenegro MF, Hezel M, Zhong J, Persson EG, Holmdahl R, Weitzberg E, Lundberg JO, Carlström M. Enhanced XOR activity in eNOS-deficient mice: effects on the nitrate-nitrite-NO pathway and ROS homeostasis. *Free Radic Biol Med*. 2016;99:472–484. doi: 10.1016/j.freeradbiomed.2016.09.004

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