Meakin Lab.

Modification Form for Permit BIO-SP-[illegible]

Permit Holder: Susan Meakin

Approved Personnel
(Please stroke out any personnel to be removed)

~~Jupinder Bains~~
~~Andrew Lu~~
Jennifer Forsyth
~~Jennifer Gerasimoff~~
Chunhui Li
~~Kim Brookes~~
~~Todd Hryciw~~
~~Sara LeMay~~
James MacDonald

Additional Personnel
(Please list additional personnel here)

Alfonso Dietrich
Asghar Talebian
Ian Grant
~~[illegible]~~

* *PLEASE ATTACH A MATERIAL SAFETY DATA SHEET OR EQUIVALENT FOR NEW BIOHAZARDS.*
** *PLEASE ATTACH A BRIEF DESCRIPTION OF THE WORK THAT EXPLAINS THE BIOHAZARDS USED AND HOW THEY WILL BE USED.*

Classification: 2

Date of last Biohazardous Agents Registry Form: May 3, 2007

Signature of Permit Holder: Susan Meakin

BioSafety Officer(s): _____

Chair,Biohazards Subcommittee: _____

	Please stroke out any approved Biohazards to be removed below	Write additional Biohazards for approval below. *
Approved Microorganisms	Yeast, E.coli containing plasmids	
Approved Cells	Human (established), rodent (established, primary), NHP (established), Hek 293, 293T cells, SY5Y, IMR32, SK-N-AS, CHP-212, BE(2)-C, SK-N-DZ, SK-N-MC, SK-N-F1, Daoy, Daoy-TrkA/TrkB/TrkC (wt and	U87MG. UW426 U341 U373
Approved Use of Human Source Material		
Approved GMO	SV 40 Large, T antigen, pCMX, pCDNA3.1, pEGFP, FRS2, FRS3, ShcB, ShcC, AKT, Fyn, NR2B, STEP isoforms, RasFrfl, Nesca, Axin, beta-catenin, FGFRs, GST fusion vectors, pAS, pACT, pGad, pAd-Easy,	
Approved use of Animals		
Approved Toxin(s)		

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** *PLEASE ATTACH A BRIEF DESCRIPTION OF THE WORK THAT EXPLAINS THE BIOHAZARDS USED AND HOW THEY WILL BE USED.*

Classification: 2

Date of last Biohazardous Agents Registry Form: May 3, 2007

Signature of Permit Holder: Susan Meaney

BioSafety Officer(s): _____

Chair,Biohazards Subcommittee: _____

The research in my lab addresses the mechanisms activated by the receptor tyrosine kinases termed Trk in the developing and mature nervous systems. The Trk family includes 3 members, TrkA (activated by nerve growth factor), TrkB (activated by brain-derived neurotrophic factor) and TrkC (activated by neurotrophin 3). My research addresses (1) mechanisms of how TrkA can activate cell death in tumors of the nervous system such as medulloblastomas and glioblastomas, (2) mechanisms where by TrkB facilitates long-term potentiation and memory, via ShcC, RasGrf1 and the NMDA receptors, (3) mechanisms of neuronal survival and neuronal process formation, via Nesca, RasGrf1 and FRS3 and (4) mechanisms of how FRS3 facilitates cortical neuron development and migration during cortical neurogenesis. We address these questions using a variety of molecular and cell biology approaches including transgenic mice, primary neuron cultures, knock down studies using knock-out mice or loss of expression via siRNA or shRNA approaches as well as transfection and over-expression approaches using plasmids or adenoviral expression in both established and primary neuronal cell culture.

Meakin
Adenovirus list.

pAd- TRICA

- TRICB

- Sh Pinder

~~[redacted]~~

-DN Pinder

- WT Pinder

- DN cdc 42

- CA cdc 42

- WT cdc 42

- PKC alpha

- PKC zeta

- JNIC 1

- JNIC 2

- DN JNIC1

- DN JNIC2

- GFP NESCA

- GFP

- Mono RED

MEAKIN LAB PLASMIDS

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1	HB101			
2	DH1			
3	pSV2gpt	HB101	amp	Mulligan & Berg
4	pSVogpt	HB101	amp	S.Meakin
5	pSVOCAT	HB101	amp	Gorman& Howard
6	pSVOCAT(Xho1)	HB101	amp	S.Meakin
7	RSVCAT	HB101	amp	Gorman& Howard
8	pUC9-HMT11a	HB101	amp	D.Hamer
9	pSGM1	HB101	amp	S.Meakin
10	pδcr17	HB101	amp	J.Piatigorsky
11	pRSV?gal	HB101	amp	J.Piatigorsky
12	pUC9	HB101	amp	J.Piatigorsky
13	pKK232-8	HB101	amp	Pharmacia
14	pSV2ACAT	HB101	amp	Kadesch&Berg
15	pSVOATCAT	HB101	amp	Meakin & Lok
16	pSTK7	HB101	amp	M.L.Breitman
17	p91023	HB101	tet	R.Kaufman
18	pM21CR3	HB101	tet	J.Piatigorsky
19	pGem-3	HB101	amp	
20	pSV0ATCAT(Xho1)	HB101	amp	S.Meakin
21	pUC9-γ2βgal	HB101	amp	S.Meakin
22	pSV2CAT	HB101	amp	Gorman& Howard
23	pGem3'γ2βgal1 Œn	HB101	amp	S.Meakin
24	pSGM1X	HB101	amp	S.Meakin
25	pUC-γ2βgal Œn	HB101	amp	S.Meakin
26	pUC-γ2Cat Œn	HB101	amp	S.Meakin
27	KM 392	HB101	amp	
28	pNGFR.1(in PUC 9) p75	HB101	amp	M.Radeke
29	pSV2NEO	HB101	amp	Mulligan&Berg
30	pBAP-NEO	HB101	amp	L.Kedes
31	pUC19	HB101	amp	
32	p19-2GR1(-ve) 5'end rNGFRgene	HB101	amp	S.Meakin
33	p19-2GR(+ve)	HB101	amp	S.Meakin
34	p19-2Ia1(2-1) aka p19-E2(+)	HB101	amp	S.Meakin
35	p19-2Ia2(2-3) aka p19-E2(-)	HB101	amp	S.Meakin
36	p19-21b(4-1) aka p19-E3	HB101	amp	S.Meakin
37	PGem-NGF(+)	HB101	amp	J.Jongstra-Bilen
38	PGem-NGF(-)	HB101	amp	J.Jongstra-Bilen
39	pXP1	HB101	amp	Steve Nordeen
40	pXP2	HB101	amp	Steve Nordeen
41	pH1-3	HB101	amp	Moses Chao
42	pH3	HB101	amp	John Darby
43	pCHTK	HB101	amp	M.Wigler
44	pC-fos(human)-1	HB101	amp	T.Curran
45	pC-fos(rat)-1	HB101	amp	T.Curran
46	pNB-1	HB101	amp	M.Schwab

	NAME	HOST Str	ANTIBIOTIC	SOURCE
47	see 85	HB101	amp	
48	PRβA-1	HB101	amp	L.Kedes
49	PSVC-myc1(mouse)	HB101	amp	Land et al. Nature 304:596
50	pUC9-42A	HB101	amp	Piotr Masiakowski
51	pUC9-42C	HB101	amp	Piotr Masiakowski
52	pE3-1	HB101	amp	S.Meakin
53	pE3-2	HB101	amp	S.Meakin
54	pE3-3	HB101	amp	S.Meakin
55	pE3-4	HB101	amp	S.Meakin
56	pE1-11	HB101	amp	S.Meakin
57	pE1-10	HB101	amp	S.Meakin
58	pE1-9	HB101	amp	S.Meakin
59	pE1-8	HB101	amp	S.Meakin
60	pE1-7	HB101	amp	S.Meakin
61	pE1-6	HB101	amp	S.Meakin
62	pE1-5	HB101	amp	S.Meakin
63	pE1-4	HB101	amp	S.Meakin
64	pE1-3	HB101	amp	S.Meakin
65	pE1-2	HB101	amp	S.Meakin
66	pE1-1	HB101	amp	S.Meakin
67	pB51	HB101	amp	M.Chao
68	pEF1	HB101	amp	M.Chao
69	PDGF	HB101	amp	A.Ulrich
70	pCNGFR(in pCDL1) p75	HB101	amp	M.Radeke
71	PCDL1-NGF	HB101	amp	
72	pE1B(-)	HB101	amp	S.Meakin
73	pE1B(+)	HB101	amp	S.Meakin
74	p2G(-) ŒB	HB101	amp	S.Meakin
75	M106/p3 (p3 plasmid containing amber mutated amp and tet resist. elements—requires a Sup F containing plasmid to confer amp, tet)	HB101	amp tet	B.Seed
76	MoIL2	HB101	amp	Ken-Ichi Arai
77	MoIL2R	HB101	amp	Ken-Ichi Arai
78	HuIL2	HB101	amp	Ken-Ichi Arai
79	HuIL2R	HB101	amp	Ken-Ichi Arai
80	pE2SB	HB101	amp	S.Meakin
81	pE3XB	HB101	amp	S.Meakin
82	CDM8	HB101	amp&tet (inM106/p3)	Brian Seed
83	p19-MoIL2R	HB101	amp	S.Meakin
84	pCH3	HB101	amp	M.Radeke
85	pOD32	HB101	tet	P.Coffno
86	pG43-1.2C3	HB101	amp	P.Skene
87	pMmH4-Hpa	HB101	amp	
88	PCDL-SRα296	HB101	amp	DNAX
89	pN1.8	HB101	amp	D.Baltimore
90	pC4	HB101	amp	Felice Tirone
91	pBJ5	HB101	amp	DNAX
92	pCAL-8Mβ	HB101	amp	Ueli Suter

	NAME	HOST Str	ANTIBIOTIC	SOURCE
94	p5-1(IL2R exoIII clones #94-#122)	HB101	amp	S.Meakin
95	p5-2	HB101	amp	S.Meakin
96	p5-3	HB101	amp	S.Meakin
97	p5-4	HB101	amp	S.Meakin
98	p5-5	HB101	amp	S.Meakin
99	p5-6	HB101	amp	S.Meakin
100	p5-7	HB101	amp	S.Meakin
101	p5-8	HB101	amp	S.Meakin
102	p5-9	HB101	amp	S.Meakin
103	p5-10	HB101	amp	S.Meakin
104	p5-12	HB101	amp	S.Meakin
105	p5-13	HB101	amp	S.Meakin
106	p5-14	HB101	amp	S.Meakin
107	p5-15	HB101	amp	S.Meakin
108	p5-17	HB101	amp	S.Meakin
109	p5-19	HB101	amp	S.Meakin
110	p5-21	HB101	amp	S.Meakin
111	p5-22	HB101	amp	S.Meakin
112	p6-1	HB101	amp	S.Meakin
113	p6-2	HB101	amp	S.Meakin
114	p6-7	HB101	amp	S.Meakin
115	p6-8	HB101	amp	S.Meakin
116	p6-9	HB101	amp	S.Meakin
117	p6-14	HB101	amp	S.Meakin
118	p6-17	HB101	amp	S.Meakin
119	p6-18	HB101	amp	S.Meakin
120	p6-21	HB101	amp	S.Meakin
121	p6-23	HB101	amp	S.Meakin
122	p6-24	HB101	amp	S.Meakin
123	pILNGFR 1(-)	HB101	amp	S.Meakin
124	pILNGFR 7(-)	HB101	amp	S.Meakin
125	pCAL-NGFR	HB101	amp	S.Meakin
126	pILNGFR 1(+)	HB101	amp	S.Meakin
127	pILNGFR 7(+)	HB101	amp	S.Meakin
128	pILNGFR 14(+)	HB101	amp	S.Meakin
129	PNRSB	HB101	amp	S.Meakin
130	PRSVneo	HB101	amp	Gorman& Howard
131	pSP651B15	HB101	amp	Danielsen et al DNA 7:261 '88
132	pMT-2	HB101	amp	R.Kaufman MCB 9:946 '89
133	pMT-ILN 1(+) *clone 1-5*	HB101	amp	S.Meakin
134	pMT-ILN 7(+) *clone 7-3*	HB101	amp	S.Meakin
135	pMT-ILN 14(+) *clone 14-2*	HB101	amp	S.Meakin
136	pBJ-ILN 1(+)	HB101	amp	S.Meakin
137	pBJ-ILN 7(+) *clone 7-6*	HB101	amp	S.Meakin
138	pBJ-ILN 14(+) *clone 14-1*	HB101	amp	S.Meakin
139	pMT-ILN 1(-) *clone 1-3*	HB101	amp	S.Meakin
140	pMT-ILN 7(-) *clone 7-7*	HB101	amp	S.Meakin

	NAME	HOST Str	ANTIBIOTIC	SOURCE
141	pMT-ILN 14(-) *clone 14-6*	HB101	amp	S.Meakin
142	pMT-NGFR(+) *clone MTN-1(rat)* p75	HB101	amp	S.Meakin
143	pMT-NGFR(-) *clone MTN-2(rat)*	HB101	amp	S.Meakin
144	pMT-IL2R(+) *clone MT1R-1(mouse)*	HB101	amp	S.Meakin
145	pMT-IL2R(-) *clone MT1R-6(mouse)*	HB101	amp	S.Meakin
146	pCD-NGFR(+) *clone #4*	M106/p3	Sup F- amp & tet (7.5)	S.Meakin
147	pCD-NGFR(-) *clone #5*	M106/p3	Sup F in M106/p3	S.Meakin
158	PTK1 *clone 1A-1*	HB101	amp	S.Meakin
159	PTK2 *clone 5A-4*	HB101	amp	S.Meakin
160	PTK3 *clone 7-1*	HB101	amp	S.Meakin
161	PTK4 *clone 7-3*	HB101	amp	S.Meakin
162	PTK5 *clone 7-4*	HB101	amp	S.Meakin
163	PTK6 *clone 8B-1 rat Neu*	HB101	amp	S.Meakin
164	PTK7 *clone 8B-2 rat FIK*	HB101	amp	S.Meakin
165	PTK8 *clone 8B-3*	HB101	amp	S.Meakin
166	PTK9 *clone 2-9 elk*	HB101	amp	S.Meakin
167	PTK10 *clone 6-6*	HB101	amp	S.Meakin
168	PTK11 *clone 12-1*	HB101	amp	S.Meakin
170	PTK12 *clone 1A-7*	HB101	amp	S.Meakin
171	PTK13 *clone 1A-9*	HB101	amp	S.Meakin
172	PTK14 *clone 1A-10*	HB101	amp	S.Meakin
173	PTK15 *clone 2-13*	HB101	amp	S.Meakin
174	PTK16 *clone 2-16*	HB101	amp	S.Meakin
175	PTK17 *clone 5A-8*	HB101	amp	S.Meakin
176	PTK18 *clone 6-14*	HB101	amp	S.Meakin
177	PTK19 *clone 7-10*	HB101	amp	S.Meakin
178	PTK20 *clone 7-11*	HB101	amp	S.Meakin
179	PTK21 *clone 7-13*	HB101	amp	S.Meakin
180	PTK22 *clone 8-2*	HB101	amp	S.Meakin
181	PTK23 *clone 11-7*	HB101	amp	S.Meakin
182	PTK24 *clone 8-3*	HB101	amp	S.Meakin
183	PTK25 *clone 8-6*	HB101	amp	S.Meakin
184	PTK26 *clone 12-6*	HB101	amp	S.Meakin
185	pSP72	HB101	amp	Pharmacia
186	pSP72-TRK 5-2	HB101	amp	S.Meakin
187	TRK 5-7 "	HB101	amp (in pSP72)	S.Meakin
188	TRK3-9 PCR of oligo 12 & 13	HB101	Amp (in pSP72)	S.Meakin
189	TRK 3-12 "	HB101	Amp (in pSP72)	S.Meakin
191	PC12 TRK clones 12-1(+)	HB101	Sup F (in CDM8)	S.Meakin
192	" 14-1(+)	HB101	Sup F (in CDM8)	S.Meakin
193	" 17-2(-) TRK(-)	HB101	Sup F (in CDM8)	S.Meakin
194	pCD17-2(+)	HB101	Sup F (in CDM8)	S.Meakin
195	pCD14-1(-)	HB101	Sup F (in CDM8)	S.Meakin
196	SP-TRK6 (in Xba 1 site) TrkA full length	HB101	Amp (in pSP72)	S.Meakin
197	SP-TRK7 (in Xba 1 site)	HB101	Amp (in pSP72)	S.Meakin

	NAME	HOST Str	ANTIBIOTIC	SOURCE
198	CD-TRK(+) *clone T-6*	HB101	Sup F (in CDM8)	S.Meakin
199	CD-NGFRG(+) (in Xho 1 site)	HB101	Sup F (in CDM8)	S.Meakin
200	CD-NGFRG(-) (")	HB101	Sup F (in CDM8)	S.Meakin
201	TRK clone 6-1	HB101	Sup F (in CDM8)	S.Meakin
202	TRK clone 11-1	HB101	Sup F (in CDM8)	S.Meakin
203	TRK clone 11-2	HB101	Sup F (in CDM8)	S.Meakin
204	TRK clone 17-2(+)	HB101	Sup F (in CDM8)	S.Meakin
205	TRK clone 22-1	HB101	Sup F (in CDM8)	S.Meakin
206	pJFE14	HB101	amp	P.Barker
207	PCMX	HB101	amp	Regeneron
208	p139 (in Xba 1 site)	HB101	Amp (in pCMX)	P.Barker
209	p146 (in Xba 1 site)	HB101	amp	P.Barker
210	pBJ-NGF	HB101	Amp (in BJ5)	C.Drinkwater/ Suter/ Barker
211	pCMX-TRK A(+) (in Xho site)	HB101	amp (in pCMX)	S.Meakin
212	pCMX-TRK A(-) (in Xho site)	HB101	amp (in pCMX)	S.Meakin
213	pCMX-TRK A(+) Nde 1	HB101	amp (in pCMX)	S.Meakin
214	pMEX	HB101	amp	Brian Stanton
215	pGEX-4T2	HB101	amp	Pharmacia
216	pMEXneo	HB101	amp, kan/G418	D.M. Zanca
217	pCMX-TRK A(+) S1 *clone 4*	HB101	amp	S. Meakin
218	pMEX-TRK A(-) *clone 4*	HB101		
219	pMEX-TRK A(+) *clone 5*	HB101		
220	pSP72-TRK A , Bgl II *clone 5*	HB101	amp	S.Meakin
221	HB101			Gibco BRL
222	pSP72-TRK A,Bgl II, *clone S2-1*	HB101	amp	J. Ping Yang
223	pCMX-TRK A T2 *clone Not 3*	HB101	amp	S.Meakin
224	pSP72-TRK A T1 *clone Stu 3*	HB101	amp	S.Meakin
225	pCMX-TRK A S2(+) *clone 9*	HB101	amp	J. Ping Yang
226	pCMX-TRK A S2(-) *clone 7*	HB101	amp	J. Ping Yang
227	pMEXNEO-TRK A(+) *clone 6*	HB101	amp/G418	S.Meakin
228	pCMX-HA-TRK A T2(+) *clone 6*	HB101	amp	S.Meakin
229	pCMX-HA-TRK A S2(+) *clone 1*	HB101	amp	S.Meakin
230	pCMX-HA-TRK A S1(+) *clone 1*	HB101	amp	S.Meakin
231	pCMX-XBA *clone 17*	HB101	amp	S.Meakin
232	pCMX-HA-TRK A T2(+) *clone 3*	HB101	amp	S.Meakin
233	pCMXHA-TRK A T1 *clone 4*	HB101	amp	S.Meakin
234	PCMX-NGF *clone 2*	HB101	amp	S.Meakin
235	PCMX-HA-TRK A S3(+) *clone 4*	HB101	amp	S.Meakin
236	PMEX-HA TRK A(+) *clone 7*	HB101	amp	S.Meakin
237	PCMX TRK B	HB101	amp	P. Barker
238	pBJ-BDNF	HB101	amp	P. Barker
239	pTK hygro	HB101	amp/hygromycin	C. Strathdee
240	PCMX TRKA Δ231-368 *clone 40-5*	HB101	amp	H. Schneider
241	PCMX TRKA Δ201-368 *clone 40-52*	HB101	amp	H. Schneider
242	PCMX TRKA Δ 263-368 *clone 30-3*	HB101	amp	H. Schneider
243	PCMX TRKA Δ322-368 *clone 30-5*	HB101	amp	H. Schneider
244	PCMX TRKA Δ 363-368 *clone 5-4*	HB101	amp	H. Schneider

	NAME	HOST Str	ANTIBIOTIC	SOURCE
245	PCMX TRKA Exo clone 20-7 Δ 328-368	HB101	amp	H. Schneider
246	PCMX-HA-TRK A S5(+) *clone 3*	HB101	amp	H. Schneider
247	pGBT9	HB101	amp	S. Fields
248	PGAD424	HB101	amp	S. Fields
249	PCMX-HA-TRK A S4(+) *clone 2F*	HB101	amp	A. Ho
250	pSP72-TRK A S7-4 * *PCR mistake*	HB101	amp	H. Schneider
251	PCMX HA-TRK A(+) Rat	HB101	amp	S. Meakin
252	PCMX TRK A Nae 1	HB101	amp	S. Meakin
253	pSP72-TRK A S6-3 * *PCR mistake*	HB101	amp	H. Schneider
254	pCMX TRK A Δ 35-58 (60-F)	HB101	amp	J. MacDonald
255	pCMX TRK A Δ 35-76	HB101	amp	J. MacDonald
256	pCMX TRK A Δ 35-102 (45-D)	HB101	amp	J. MacDonald
257	pCMX TRK A Δ 35-107 (40-8)	HB101	amp	J. MacDonald
258	pCMX TRK A Δ 35-114 (20-4)	HB101	amp	J. MacDonald
259	pCMX TRK A Δ 35-139 (60-8)	HB101	amp	J. MacDonald
260	pCMX TRK A Δ 35-150 (60-7)	HB101	amp	J. MacDonald
261	pCMX TRK A Δ 35-174 (60-1)	HB101	amp	J. MacDonald
262	pCMX TRK A Δ 35-290 (FN mutant) clone 1	HB101	amp	J. MacDonald
263	pCMX HA-TRK A S7 *clone 7*	HB101	amp	H. Schneider
264	pCMX HA-TRK A S8 *clone 4*	HB101	amp	H. Schneider
265	pCMX HA-TRK A S8/S9 *clone 1*	HB101	amp	H. Schneider
266	pCMX HA-TRK A S6 clone 4	HB101	amp	H. Schneider
267	pCMXTRK A Δ372-390 clone H1	HB101	amp	S. Meakin
268	pCMX MYC-TRK A S1 clone 5	HB101	amp	S. Meakin
269	pCMX MYC-TRK A clone 5	HB101	amp	S. Meakin
270	pcDNA TRK C clone 1	HB101	amp	S.Meakin/Amgen
271	pB61 rel est	HB101	amp	Amgen
272	pMEX TRK A S8/S9(-) clone 12	HB101	amp	S. Meakin
273	HB101			Gibco
274	HB101			Gibco
275	pAS-1	HB101	amp	C. Brandl
276	pACT I	HB101	amp	C. Brandl
277	pACT II	HB101	amp	C. Brandl
278	pMEX HATRK A Exo III clone 25-1	HB101	amp	H.Schneider
279	pMEX HATRK A Exo III clone 25-5	HB101	amp	H.Schneider
280	pMEX HATRK A S8/S Exo III clone 20-4	HB101	amp	H.Schneider
281	pADB	HB101	amp	C.Strathdee
282	pTKB	HB101	amp	C.Strathdee
283	pCMVBgal	HB101	amp	C.Strathdee
284	pMEX HATRK A Exo III clone 30-4	HB101	amp	H.Schneider
285				
286				
287	pAS-1 SHC full NOTE:re-transformed 2.1.98 by CJK	HB101	amp	Jane McGlade
288	pAS-1 SHC PTB (ShcA)	HB101	amp	Jane McGlade

	NAME	HOST Str	ANTIBIOTIC	SOURCE
289	pG SHC B15 pGem3-human SHC	HB101	amp	Jane McGlade
290	p448 IRS clone	HB101	amp	Thomas Gustafson
291	p390 IRS clone	HB101	amp	Thomas Gustafson
292	pAS-1 lerk 5 clone 3	HB101	amp	H.Schneider
293	pCMX MYC TRK A S3 clone 4	HB101	amp	H.Schneider
294	pGAD424-TRK A clone 4	HB101	amp	H.Schneider
295	pCMX 57 TRK A clone 1	HB101	amp	J. MacDonald
296	pGAD-IR-IRS-1 clone H	HB101	amp	E. Gryz
297	pGAD TRK A S1 clone 1	HB101	amp	H. Schneider
298	pGAD TRK A S3 clone 1	HB101	amp	H. Schneider
299	pGAD TRK A S8 clone 1	HB101	amp	S. Meakin
300	pAS-1 TRK A clone 8	HB101	amp	H. Schneider/S.M.
301	pGAD TRK A S1/S3 clone 3	HB101	amp	H. Schneider
302	pGEX-4T2/ TRKA 25-1 clone 1	HB101	amp	E.Gryz
303				
304	Rat chromograninA Pst 1 insert	HB101	amp	Munoz
305	pGAD-IRS-1 clone J	HB101	amp	E.Gryz
306	pGBT9-IR-IRS-1 clone 5	HB101	amp	E.Gryz
307	pGBT9-IRS-1 clone 8	HB101	amp	E.Gryz
308	pBPXp57TRK A(+) clone 2	HB101	amp	J.MacDonald
309	pCMX-HA TRK A S10 clone 17	HB101	amp	H.Schneider
310	pGEX4T2/TRKA 25-1 clone G	HB101	amp	E.Gryz/H.S.
311	pAS-1/IR-IRS-1 clone 1	HB101	amp	E.Gryz
312	pBPX-TRK A S8 clone 2	HB101	amp	S.Meakin
313	pGAD TRK A S1/S8 clone 1	HB101	amp	H.Schneider
314	pDBH (Dopamine β-hydroxylase)	HB101	amp	Richard Palmiter
315	pSK(+)	HB101	amp	Stratagene(C.S.)
316	pSK(-)	HB101	amp	Stratagene
317	pSKTRK A (+) clone A	HB101	amp	S.Meakin
318	pSKTRK A (-) clone 4	HB101	amp	S. Meakin
319	pGAPDH	HB101	tet	J. Pickering
320	pKS(+)	HB101	amp	Stratagene-M.Lebat
321	pKS(-)	HB101	amp	Stratagene-M.Lebat
322	pCMX-HA TRK A S1/S8 clone 1	HB101	amp	H.Schneider
323	pGUS 299	HB101	amp	T.Gustafson
324	pGUS 306	HB101	amp	T.Gustafson
325	pGUS 318	HB101	amp	T.Gustafson
326	pGUS 392	HB101	amp	T.Gustafson
327	p15γMAG-Fc chimera	HB101	amp	M.Tropak
328	pUChγ?1.1	HB101	amp	M.Tropak
329	pUChγ?1.2	HB101	amp	M.Tropak
330	pCMX-HA TRK A S15 clone 3	HB101	amp	E.Gryz
331	pCMX-HA TRK A S16 clone 3	HB101	amp	E.Gryz
332	pJDM(NGF1A)	HB101	amp	Jeff Milbrandt
333	pJDM(NGF1B)	HB101	amp	Jeff Milbrandt
334	pBS Tα1 tubulin promoter	HB101	amp	Freda Millar

	NAME	HOST Str	ANTIBIOTIC	SOURCE
335	pCMX TRK A S18 clone 1	HB101	amp	H. Schneider
336	pSPR TRK A S17 clone 1	HB101	amp	H. Schneider
337	pSK HA TRK A S11a clone 5#4	HB101	amp	H. Schneider
338	pGST SHC 1-209		amp	Jane McGlade
339	pGST SHC/SH2		amp	Jane McGlade
340	pECE-γ???Xho clone 2	HB101	amp	S.Meakin
341	pSK TRK A S12p clone 4p	HB101	amp	E.Gryz
342	pSK TRK A S12 clone 36	HB101	amp	E.Gryz
343	pSK TRK A S13a clone 3a	HB101	amp	E.Gryz
344	p CMX TRK A S19 clone 2	HB101	amp	E.Gryz
345	pGAD TRK A S11a clone 7	HB101	amp	H.Schneider
346				
347	pCMX HA TRK A S17 clone 1	HB101	amp	H.Schneider
348	pClone73 (Peripherin)	HB101	amp	Ed Ziff
349	pCMX HA TRK A S18 clone 1	HB101	amp	H.Schneider
350	pγ?X-HATRK A lower clone 1(+)	HB101	amp	H.S./S.Meakin
351	pCMXHA TRK A S12 clone 1	HB101	amp	E.Gryz
352	pCMXHA TRK A S12p clone 2	HB101	amp	E.Gryz
353	pCMXHA TRK A S13a clone 2	HB101	amp	E.Gryz
354	pCMXHA TRK A S19 clone 2	HB101	amp	E.Gryz
355	pTK HA TRK A clone 7	HB101	amp	H.Schneider
356	pCMXHA TRK A S1b clone 2	HB101	amp	H.Schneider
357	pBPX HA TRK A S15 clone 3	HB101	amp	E.Gryz
358	pCMXHA TRK A S13b clone 7	HB101	amp	E.Gryz
359	pGST-PTP1C	HB101	amp	K.Siminovitch
360	pUHD 15-1 *tet vector system*	HB101	amp	Bujard ZMBH
361	pUHD 15-1 neo	HB101	amp	Bujard ZMBH
362	pUHD 151-1	HB101	amp	Bujard ZMBH
363	pUHG 17-1	HB101	amp	Bujard ZMBH
364	pUHD 172-1neo	HB101	amp	Bujard ZMBH
365	pUHC 13-3	HB101	amp	Bujard ZMBH
366	pUHG 16-3	HB101	amp	Bujard ZMBH
367	pUHD 10-3	HB101	amp	Bujard ZMBH
368	pUHC 13-6	HB101	amp	Bujard ZMBH
369	pUCN-CAD(N-Cadherin in pUC18)	HB101	amp	M. Takeichi
370	pRSV-N-CAM	HB101	amp	Ted Lo
371	PC12-BR7(phosphatase) Bluescript	HB101	amp	Paul Lombroso
372	pCMXHA TRK A S10T1 clone 1	HB101	amp	H.Schneider
373	pUHD 10-3 MYC TRK A clone 2	HB101	amp	H.Schneider
374	pSP72BglII TRK A Xho 411 clone 1	HB101	amp	H.Schneider
375	pCMXHA TRK A Xho 411 clone 2	HB101	amp	H.Schneider
376	pUHD 10-3 p75 clone 3	HB101	amp	H.Schneider
377	pCMXHA TRK A S9 clone 21	HB101	amp	E.Gryz
378	pCMXHA TRK A S11b clone 2	HB101	amp	H.Schneider
379	pGEX 4T2/TRK A clone 2	HB101	amp	E.Gryz
380	pBPX HATRK A S9 clone 4	HB101	amp	E.Gryz
381	p γ?X TRK A Xho 411(-) clone A	HB101	amp	H.Schneider

	NAME	HOST Str	ANTIBIOTIC	SOURCE
382	pGAD TRK A S11b clone 3	HB101	amp	H.Schneider
383	pAS-1TRK A S11b clone 5	HB101	amp	H.Schneider
384	pCH126A2 clone 2	HB101	amp	Frank Lee
385	pGEX4T2 TRK A S15 clone 3	HB101	amp	E.Gryz
386	pGEX4T2 TRK A S15 clone 5	HB101	amp	E.Gryz
387	BL21(lambda DE3 strain)	HB101	amp	Ted Lo
388	pSK TRK S13(glu,glu) clone 10	HB101	amp	E.Gryz
389	p γ?X TRK Xho 411(+) clone CD5	HB101	amp	H.Schneider
390	pCMX HA TRK A LRM II	HB101	amp	J.MacDonald
391	pBPXHA TRK A S1 clone 2	HB101	amp	H.Schneider
392	pCh110	HB101	amp	Pharmacia
393	pCMX HA TRK A -Bgl I	HB101	amp	J.MacDonald
394	pCMX TRK A S15(QQ)	HB101	amp	E.Gryz
395	pBTM 116	HB101	amp	J.Verdi/S.Fields
396	pBPE	HB101	amp	J.MacDonald
397				
398	pGM2163	HB101	chloramphenicol resistant	NEB
399	pCh126A2	HB101	amp	Frank Lee
400	pCMXHA TRK A S13(gl,gl) clone16	HB101	amp	E.Gryz
401	pCMXHA TRK A S13 (gl,gl) clone13	HB101	amp	E.Gryz
402	pBSK TRK A-Nco I	HB101	amp	J.MacDonald
403	pBPE-NCad	HB101	amp	J.MacDonald
404	pCMX TRK C (rat)	HB101	amp	Phil Barker
405				
406	pUHD 10-3 TRK A S13(gl,gl) clone 3	HB101	amp	E.Gryz
407	pUHD 10-3 TRK A S13b clone 5	HB101	amp	E.Gryz
408	pUHD 10-3 TRK A S13a clone 16	HB101	amp	E.Gryz
409	pATG lac Z(Sal)2	HB101	amp	Qiurong Liu
410	pCDNA3 trk A *human*	HB101	amp,neo	U.Saragovi
411	pKS Bluescript-RN-gATA-4	DH5 α	amp	I. Skerjanc
412	pBluescript II SK¯NBx 2.5	DH5 α	amp	I. Skerjanc
413	pGK-MEF2C-(E15)	DH5 α	amp	I. Skerjanc
414	pEMC IIS-MyoD (probe)	DH5 α	amp	I. Skerjanc
415				
416	pCMXHATRK A S13c-4e(asp,asp)	HB101	amp	E.Gryz
417	pBPXHA TRK A S13 clone 1	HB101	amp	E.Gryz
418	pBPXHA TRK A S13a clone 8	HB101	amp	E.Gryz
419	pGL3 control(for luciferase assay)	HB101	amp	Promega
420	pUHD10-3 HA TRK A S13c clone 2	HB101	amp	E.Gryz
421	pBPXHA TRK A S13c clone 6	HB101	amp	E.Gryz
422	pBPXHA TRK A S13b clone 8	HB101	amp	E.Gryz
423	pAS-1 TRK A S1 clone 4	HB101	amp	E.Gryz
424	pBK/SK	HB101	kanamycin	C.Strathdee
425	pSP72 HA TRK A S17a clone 6	HB101	amp	H.Schneider
426	pSP72 HA TRK A S3a clone 8	HB101	amp	H.Schneider
427	pCMX TRK B S3 clone 14	HB101	amp	G.Z.
428	pCMXHA TRK A S13p57 clone 3	HB101	amp	H.Schneider

	NAME	HOST Str	ANTIBIOTIC	SOURCE
429	pCMXHA TRK A LRM 1	HB101	amp	J.MacDonald
430	pCMXTRK B S3 clone 14	HB101	amp	G.Z.
431	pCMXHA TRK A S17a clone 1	HB101	amp	H.Schneider
432	pCDNA 3 clone 1	HB101	amp	C.Strathdee
433	pCMXHA TRK A S3a clone 21	HB101	amp	H.Schneider
434	pBPXho Baculovirus vector	HB101	amp	J.MacDonald
435	pBPXLRM I TRK A	HB101	amp	J.MacDonald
436	pBPXLRM II TRK A	HB101	amp	J.MacDonald
437	pBPXLRM III TRK A	HB101	amp	J.MacDonald
438	pGexEG2 clone A	BL21	amp	E.Gryz
439	pJG4-5 Grip2 (SH2)	HB101	amp	Gustafson
440	pAcid SYP (SH2)	HB101	amp	Gustafson
441	pJG4-5 p85 acid blob	HB101	amp	Gustafson
442	pCMXHA TRK AS13d clone 2	HB101	amp	E.Gryz
443	pCMXHA TRK AS13e clone 3	HB101	amp	E.Gryz
444	pCMXHA TRK AS13f clone 1	HB101	amp	E.Gryz
445	pCMXHA TRK AS13g clone1	HB101	amp	E.Gryz
446	PLCX (-) MT-2	HB101	amp	McGlade
447	pUHD 10-3HA TRK AS13d clone4	HB101	amp	E.Gryz
448	pUHD 10-3HA TRK A S13eclone2	HB101	amp	E.Gryz
449	pUHD 10-3HA TRK A S13f clone2	HB101	amp	E.Gryz
450	pGex TRK A (from stock #302)	BL21	amp	E.Gryz
451	pGex SHC/PTB (from #338)	BL21	amp	McGlade
452	pGex SHC/SH2 (from #339)	BL21	amp	McGlade
453	pBPX HA TRK AS13d clone 3	HB101	amp	E.Gryz
454	pBPX HA TRK AS13e clone 2	HB101	amp	E.Gryz
455	pBPX HA TRK AS13f clone 4	HB101	amp	E.Gryz
456	pBPX HA TRK AS13g clone 2	HB101	amp	E.Gryz
457	pUHD 10-3HA TRK AS13g cloneC	HB101	amp	E.Gryz
458	pGEX 4T2 EG1 B	BL21	amp	E.Gryz
459	pUHD 10-3HA S1HA TRK A clone 1	HB101	amp	H.Schneider
460	pSP72 TRK B (1.3kb) clone 2	HB101	amp	G.Z.
461	pSP72 TRK B (3.0kb, (-)) clone 2	HB101	amp	G.Z.
462	pSP72 TRK B (3.0kb?, (+)) clone 6	HB101	amp	G.Z.
463	PBRX-HA-TrkA	HB101	Amp	James
464	pCMX HA TRK A S1d clone 1	HB101	amp	H.Schneider
465	DH10B			
466	pCol.luc.3	HB101	amp	
467	pAS-1 TRK A S3 clone 1	DH10B	amp	E.Gryz
468	pSP72 HA TRK B clone 2	HB101	amp	G.Z.
469	pSP72 HA TRK B clone 4	HB101	amp	G.Z.
470	pGex 4T2	BL21	amp	E.Gryz
471	pBS-SK(-) TRK C(Ki14) Rat	HB101	amp	P.Tsoulfas
472	pBS-SK(-) TRK C(Ki25) Rat	HB101	amp	P.Tsoulfas
473	pCMXHA trk B (rat) clone 1	HB101	amp	GZ
474	pBR322	HB101	amp,kan	Gibco
475	pAS EG1 clone a	HB101	amp	E.Gryz

	NAME	HOST Str	ANTIBIOTIC	SOURCE
476	pAS EG1 clone b	HB101	amp	E.Gryz
477	pRK172-p13 suc-1	BL21(DE3)	amp	D.Litchfield
478	pCMX HA trk B S3 clone 1	HB101	amp	GZ
479	pCMX HA trk B S3 clone 2	HB101	amp	GZ
480	pSP72 HA trk B (-) clone 1	HB101	amp	GZ
481	pCDM8 EG1 clone 13-1	M106/p3	supF/amp/tet	S.Meakin
482	pCDM8 EG1 clone 13-3	M106/p3	supF/amp/tet	S.Meakin
483	pCDM8 EG1 clone 13-4	M106/p3	supF/amp/tet	S.Meakin
484	pCDM8 EG1 clone 2-1	M106/p3	supF/amp/tet	S.Meakin
485	pCDM8 EG1 clone 3-1	M106/p3	supF/amp/tet	S.Meakin
486	pGEX 5X-3	HB101	amp	Pharmacia
487	pCMX HA trk B ShcF-Y S8	HB101	amp	GZ
488	pCMX HA trk B Xho	HB101	amp	GZ
489	pAS-1 BNTRK A(LRM) clone 3	HB101	amp	H.Schneider
490	pSP72 HA TRK B(+) clone 4	HB101	amp	GZ
491	pEGFP N1	HB101	kan	Clontech(C.S.)
492	pEGFP C1	HB101	kan	Clontech(C.S.)
493	pSP72HA TRK B S9 clone 1	HB101	amp	GZ
494	pCMXHA TRK A(2Eco R1, 1 Xho 1)	HB101	amp	GZ
495	pBPX HA γ??TRK A II	HB101	amp	J.MacDonald
496	pGEX TRK 128	HB101	amp	J.MacDonald
497	pCDM8 EG1 clone 7-1b,a	M106/p3	supF	S.Meakin
498	pCDM8 EG1 clone 7-1b,b	M106/p3	supF	S.Meakin
499	pCMXHA TRK B S9	HB101	amp	GZ
500	pGEX TRK 47	HB101	amp	J.MacDonald
501	pEGFPN1 HA TRK A XP clone 3	HB101	kan	H.Schneider
502	pCMXHA TRK B S8S9 clone 5	HB101	amp	GZ
503	pAS-1 p13 suc 1	HB101	amp	S.Meakin
504	pAS-1 HA TRK A LRM	HB101	amp	J.MacDonald
505	pGEX-1N	JM109	amp	D.Litchfield
506	pGEX-2T	HB101	amp	D.Litchfield
507	pGEX-3X	DH5?	amp	D.Litchfield
508	p γ?X HA TRK A LRM(-)	HB101	amp	S.Meakin
509	pCMX-cyclin D2(R1-Xho) clone3	HB101	amp	G.Z.
510	pCMXHA TRK A X411(2R,1X)	HB101	amp	G.Z.
511	pBTM116.p13 suc 1.	HB101	amp	E.Manto
512	pUHD10-3 HA TRK A (-) clone 1	HB101	amp	E.Gryz
513	pEGFPN1HA TRK A 60-7 clone 1	HB101	kan	H.Schneider
514	pGEX-1N trk 5	HB101	amp	J.MacDonald
515	pGEX-1N trk 59	HB101	amp	J.MacDonald
516	pAD-D2/R1/R1 clone 3	HB101	amp	G.Z.
517	pCMX HA TRK AB clone 4	HB101	amp	G.Z.
518	pCMX HA TRK BA clone 5	HB101	amp	G.Z.
519	pGEX-1N	BL21	amp	J.MacDonald
520	pGEX 1N trk 5	BL21	amp	J.MacDonald
521	pGEX 1N trk 59	BL21	amp	J.MacDonald
522	pBTM116 LRM HA TRK A	HB101	amp	J.MacDonald

	NAME	HOST Str	ANTIBIOTIC	SOURCE
523	p γ?LRM HA TRK (+) clone 32	HB101	amp	H.Schneider
524	pUHD 10-3 cyclin D2 clone 12	HB101	amp	GZ
525	pGEX 1N trk 86	BL21	amp	J.MacDonald
526	pUHD 10-3 HA TRKA(+)clone 2	HB101	amp	E.Gryz
527	pCMX HA TRK A S25 clone 14	HB101	amp	H.Schneider
528	pGEX-4T1 trk 47	BL21	amp	J.MacDonald
529	pGEX-4T1 trk52	BL21	amp	J.MacDonald
530	pGEX human Grb2 SH3 (N) 1- 58 a.a.	BL21	amp	M. Anafi/T. Pawson
531	pGEX human Grb2 SH3 (C) 159-217 a.a.	BL21	amp	M. Anafi
532	pGEX mouse Grb2 full length 2-217 a.a.	BL21	amp	M. Anafi
533	pGEX SH2 domain (PLCΩSH2)	BL21	amp	M. Anafi
534	pGEX mouse SOS tail	BL21	amp	M. Anafi
535	pcDNA3-HA Grb2	HB101	amp	M.Anafi
536	pGEX-4T1 trk 68	BL21	amp	J.MacDonald
537	pGEX-4T1 trk 76	BL21	amp	J.MacDonald
538	pGEX-4T1 trk 110	BL21	amp	J.MacDonald
539	pGEX-4T2 trk 23	BL21	amp	J.MacDonald
540	pGEX-4T2 trk 17	BL21	amp	J.MacDonald
541	pGEX-4T2 trk 105	HB101	amp	J.MacDonald
542	pHYB LEX ZEO TRK	HB101	zeocin	J.MacDonald
543	pcDNA3 HACrk II	HB101	amp	M.Anafi
544	pCMX-CYCLIN D2(R1/R1) +	HB101	amp	GZ
545	pCMX-CYCLIN D2(R1/R1) -	HB101	amp	GZ
547	pWWP.LUC	HB101	tetracyclin/chloramphen.	B.Vogelstein
548	pGEX4T2(back up)	BL21	amp	E.Gryz
549	pGEXSHC-PTB(back up) -- ShcA	BL21	amp	J.McGlade
550	pGEX EG1(back up)	BL21	amp	E.Gryz
551	pBPX γHA LRM TRK A(-) clone 4	HB101	amp	H.Schneider
552	pBPX γHA LRM TRK A(-) clone27	HB101	amp	H.Schneider
553	p53/T1198	BL21	amp	K.Huppi
554	p107(1-4E)	BL21	amp	K.Huppi
555	pGEM3Z-mo MYC antisense	BL21	amp	K.Huppi
556	pGEM4Zp21-9C	HB101	amp	Erhardt/Pitman
557	p3'SS	HB101	amp	Erhardt/Pitman
558	pOPRSVICAT(p21)	HB101	amp	Erhardt/Pitman
559	pBPX HA TRK A S17 clone 5	HB101	amp	B.Napier
560	pBABE-hygro	HB101		Morgenstern(Verdi)
561	pBABE-hygro	HB101	amp	Morgenstern(Verdi)
562	pBABE-bleo	HB101		Morgenstern(Verdi)
563	pBABE-puro	HB101		Morgenstern(Verdi)
564	pLXSHD	HB101		Morgenstern(Verdi)
565	pLHDCX	HB101	amp	Morgenstern(Verdi)
566	pBPXHA TRK A SIIb clone 5	HB101	amp	B.Napier
567	pGEX 5X-3 clone 73	BL21	amp	J.MacDonald

	NAME	ST Str	TIBIOTIC	SOURCE
568	pCMX-p21 (+) (R1/R1)	101	p	GZ
569	pCMX-p21 (-) (R1/R1)	101	p	GZ
570	pUMD10-3-p21	101	p	GZ
571	pBABE-hygro-p21	101	p	GZ
572	pRSV-cyclin D1 (mouse)	101	mp	C.J. Sherr
573	pSRα mSV p19 tk CD8	101	mp	C.J. Sherr
574	pBPX HA S3	15α	mp	James
575	trk 18-3 pGEX-1N	21	p	mes
576	trk 118-2 pGEX-1N	21	p	mes
577	trk 78(A) pGEX-1N	21	p	mes
578	trk 4 pGEX-1N	21	p	mes
579	trk 16 pGEX-1N	21	p	mes
580	trk 124 pGEX-4T1	21	p	mes
581	trk 107 pGEX-4T1	21	p	mes
582	trk 38 pGEX2T	21	mp	Manto
583	trk 100 pGEX-4T1	21	mp	Manto
584	cyclin D2 pGEX-5X-3	21	p	lacDonald
585	trk43 pGEX-4T1	21	mp	lacDonald
586	trk 111 pGEX-4T1	21	p	Manto
587	trk 136 pGEX-4T1	21	p	Manto
588	trk 33 pGEX-4T1	21	p	Manto
589	trk 32 pGEX-4T1	21	p	Manto
590	trk 72 pGEX-4T1	21	p	Manto
591	trk 105 pGEX-1N	21	p	Manto
592	phyblexA Zeo	101	cin	rogen
593	pBPX HA S8	101	p	es
594	pCMX-p19 (+)	101	p	
595	pCMX-p19 (-)	101	p	
596	pUHD10-3-p19 (+)	101	p	
597	pUHD10-3-p19 (-)	101	p	Z
598	pCMX-cyclinD1 (+)/R1 clone 5	101	p	
599	pCMX-cyclinD1 (-)/R1 clone 8	101	p	
600	pBSSK EG-1	101	p	s
601	pBSSK EG-2	101	p	s
602	pGEX-abl	101	amp	S. erguson
603	pGEX-crk N-SH3	101	amp	S. Ferguson
604	pGEX-crk full length	101	p	S. Ferguson
606	pUHD10-3-D1 (+)	101		
607	pBABE-hygro-D2 (+)	101	p	
608	pBABE-hygro-D2 (-)	101	p	
609	pBSSK- TRK Jux	101	p	eakin
610	pAS-1- **FRS2** (mutant)	101	p	eakin
611	pGEX-Grb2 SH3(N) 1-58 a.a. (backup) (human)	21	amp	T. Pawson
612	pGEX-Grb2 SH3 (C) 159-217 a.a. (backup) (human)	21		T. Pawson

	NAME	HOST Str	TIBIOTIC	OURCE
613	pGEX-Grb2 SH2 (backup) (human)	BL21	)	T. Pawson
614	pGEX-Grb2 full 2-217 a.a. (backup) (mouse)	BL21	)	T. Pawson
615	pGEX-HA- **FRS2** (mutant)	BL21	)	S. Meakin
616	pAS-2.1	HB101	)	S. Meakin
617	pAS-2.1- **FRS2** (1.5 kb) (Nco-Sal) (mutant)	HB101	)	S. Meakin
618	pcDNA3-HA- **FRS2** (1.5 kb) (Nhe-Sal/Xho) (mutant)	HB101		S. Meakin
619	pGST-Rb-(terminus) a.a. 792-928	BL21	)	E. Harlow MCB 142:2077-2086
620	pCMV-p16	HB101	)	Ed Harlow
621	pcdk4-dN	HB101	)	Ed Harlow Science, 262: 2050-2054 1993
623	plexhybzeo-LRM	HB101	in	James
624	pGAD-trk17	HB101		Verdi/James
625	pGEX4T1-trk128	HB101		James
626	pcDNA-trk86	HB101		James
627	pAS-2.1-LRM	HB101	)	James
628	SCS 110 (dam-,dcm-)			Stathdee / Stratagene
629	pGAD10-GRB2 (mouse)	HB101	)	CLK
630	pGAD424-p13	HB101	)	CLK
631	pBPX-HA-TRKB	HB101		CLK
632	pCMX-HA-TRKC (rat)	HB101		S. Meakin
633	pCR2.1-SOM1 (new mutant)	HB101		J. Verdi
634	pGADtrk5-II	HB101		James
635	pGADtrk86-II	HB101	)	James
636	pGEX-trk128	BL21		James
637	pcDNAtrk47	HB101	)	James
638	pcDNAtrk52	HB101		James
639	pAS-2.1-TRKA	HB101		S. Meakin
640	pAS-2.1- **FRS2** (PTB)	HB101		CLK
641	pBPX-HA-TRKC (rat)	HB101		CLK
642	pAS-2.1-SHC (full)	HB101		CLK
643	pCR2.1- **FRS2**			CLK—sequenced—this one is actually full length and has stop codon (SL)
644	pBPX-HA-TRKC (#5)	HB101		CLK
645	pGST-PTPIC			F. Siminovitch
646	pGAD424-SHCA (full)	HB101		CLK
647	pGEX4T1-SHC (full)	BL21		CLK
648	pGEX4T2-trk70	HB101		James
649	pGEX4T2- **FRS2**	BL21		CLK
650	pGEX4T2- **FRS2** (PTB)	BL21		CLK
651	pAS-1- **FRS2**	HB101		CLK
652	pAS-2.1- **FRS2**	HB101		CLK
653	pGEX-4T1-trk63	BL21		James
655	PGV16-RasGrf	HB101		Lowy
656	pGEX-5X3-peripherin	BL21		James
657	pGEX-RBD Ras Binding Domain	GC		Holloway

	NAME	HO	Str	IBIOTIC	URCE
658	pGEX2T-PLCγ1 (N+C) (SH2)	JM1			son
659	pGEX2T-Syp NC SH2	DH5			son
660	pBABE-IRES Ap-puro	HB1	1		erdi
661	pGEX2T-Syp NC SH2	BL2			son
662	pGEX2T-PLCγ1 (N+C) (SH2)	BL2			son
663	GST-abl (SH3)				i Fiore
664	GST-yes (SH3)				i Fiore
665	GST-nckl (SH3)				i Fiore
666	pAS. 1-S9 (TrkA)	H	1		s
667	pBABE-AP-puro-p21 (-) C1	HI	1		
668	pBABE-AP-puro-p21 (+) C2	HB	1		
669	GST-GAP (SH3)	DH			son
670	GST-p85 (SH3)	JM1			son
671	GST-V-SRC (SH3)	JM1	1		on
672	myc-GRF-pBABE-puro	HB1	1		kson 273:1782-1787
673	pCRScript- **FRS2** (carboxy term)	D			
674	pBABE-IRES-AP-puro-D1 (+)	D			
675	pcDNA3.1-myc-His- **FRS2** (pure)	H	1		missing last 5 aa
676	pMIAORI (βgal)	H	1		erdi
677	pGFP-N2- **FRS2**	H	1		sequenced – missing last 5 aa
678	pGAD424-HA- **FRS2**	H	1		
679	pGAD424-peripherin	HB1	1		
680	DH5α				
681	pGAD424-peripherin	H	1		
682	pAS1-S8 (TRKA)	D			
683	pLEN-FGFR	D			di
684	pEGFP-N1-numb1	D			
685	pEGFP-N1-numb3	D			
686	pBACPAK8-GST(4T2)	D			
687	pCRScript- **FRS2**-5'Race (human)	D			
688	pCRScript- **FRS2**-3'Race (human)	DH5			
689	PTR	D			i
690	pCRET-1	D			di
691	pMCRET	D			di
692	pcDNA3-**HA-FRS2** (HA on 5' end)	D			
694	pBPX-HA-TRKA S8/S9	X	gold		
695	pRc-**cSRC** (human) ?vSRC	D			guson/D. Fujita
696	pCRScript	D			gene
697	pAS2.1- **FRS2** (c-terminus)	D			
698	pGEX4T1- **FRS2** (c-terminus)	D			
699					
700					
701	pEGFP-N2- **FRS2** (full length)	D			–Change last 5 aa
702	pEGFP-N1- **FRS2** (PTB) (Bam site)	D			
703	pIRES-EGFP- **FRS2**	D			missing last 5 aa
704	pBACPAK8-GST(4T2)-trk5	D			

	NAME	[illegible] Str	[illegible]IBIOTIC	[illegible]RCE
705	pBACPAK8-GST(4T2)-trk86	DH[illegible]		[illegible]M
706	pBACPAK8-GST(4T2)-trk52	DH[illegible]		
707	pCRScript- **FRS2** Interactor #5	DH[illegible]		
708	pSE380-trkA T1 (3' Nco-Xba fragment)	DH[illegible]		
709	pBACPAK8- **FRS2**-myc	DH5[illegible]		
710	pBACPAK8-GST(4T2)- **FRS2**	DH5[illegible]	[illegible]p	
711	pAS2.1-trkA T1	DH[illegible]		
712	GST-crk SH3 (N)	DH[illegible]		[illegible]n/M. Gold
713	GST-crk SH2	DH[illegible]		[illegible]n/M. Gold
714	pCR2.1-**FRS3** (human)	DH[illegible]		
715	pUHD- **FRS2** -EGFP	DH[illegible]		[illegible]nissing last 5 aa
716	pUHD- **FRS2** (PTB)-EGFP	DH[illegible]		
717	pUHD- **FRS2**-myc	DH5[illegible]		
718	pGEX4T1-**FRS3** (human)	DH5[illegible]		
719	pBS Tα1- **FRS2**	DH[illegible]		
720	pBS Tα1-NECDIN	DH[illegible]		
721	pBS Tα1-MAGY	DH[illegible]		
722	pINTRON2-NECDIN	DH[illegible]		
723	pCR2.1-trk105-5' Race (rat)	DH[illegible]		
	The next ones are Gina's yeast scree[illegible]es for [illegible]s bait			
725	pACT2-mouse3 Human chr7 cosmid	[illegible]1		
726	pACT2-mouse5 Ribosomal protein	[illegible]1		
727	pACT2-mouse10 Novel	[illegible]1		
728	pACT2-mouse12 Ribosomal protein	[illegible]1		
729	pACT2-mouse14 Ribosomal protein	[illegible]1		
730	pACT2-mouse22 GADIO	[illegible]1		
731	pACT2-mouse28	[illegible]1		
732	pACT2-HFB3 Alu Repeats	[illegible]		
733	pACT2-HFB6 Soares mRNA	[illegible]		
734	pACT2-HFB10 GADIO	[illegible]1		
735	pACT2-HFB12 Soares Transcr. Factor	[illegible]		
736	pACT2-Mam32	[illegible]		
737	pACT2-Mam36 ATP Synthase	[illegible]1		
738	pACT2-Mam60	[illegible]1		
739	pACT2-Mam65	[illegible]1		
740	pACT2-Mam67 Zfp 7	[illegible]1		
741	pACT2-Mam75 Hum. RNA helicase p68	[illegible]2		
742	pACT2-Mam80 P1-3 Kinase	[illegible]1		
743	pACT2-Mam81 ATP Synthase	[illegible]1		
744	pACT2-Mam86 SH3-Domain interact. prot.	[illegible]		
745	pACT2-Mam87 Nebulin	[illegible]		
746	pACT2-Mam95 Kox3 Zincfinger protein	[illegible]1		
747	pACT2-Mam97	[illegible]3101		

	NAME	T Str	BIOTIC	RCE
748	pACT2-Mam98Zingfinger protein ZFp6			
749	pACT2-Mam103 GT4-2 Homolog			
750	pACT2-Mam116 N-Rap			
751	pACT2-Mam119 Chr 16 Zincfinger prot.			
752	pACT2-Mam122			
753	pACT2-Mam130			
754	pACT2-Mam133			
755	pACT2-Mam140 Human Bac clone chr7			
756	pACT2-Mam150			
757	pACT2-Mam156 Novel			
758	pACT2-Mam157			
759	pACT2-Mam164 Human chr7			
760	pACT2-Mam170 Zincfinger protein			
761	pACT2-Mam171			
762	pACT2-Mam174			
763	pACT2-Mam187			
764	pACT2-Mam212 Synapse - Assoc.protein			
765	pACT2-Mam213			
766	pACT2-Mam219			
767	pACT2-Mam224			
768	pACT2-Mam249 CyclinG1 interact. prot.			
769	pACT2-Mam269			
770	pACT2-Mam282 Dead box protein p72			
771	pACT2-mouse1 -Nesca			
	we now resume with our regularly scheduled clones			
772	pEBG			borg
773	pEBG-**RasGRF1**			borg
774	pGEX 2T **-crkl** (see 94/85 pubs from lab)			roffen
775	**nestin-c**			li
776	nestin-H-tk g11 TKZ (flipped)			li
777	pBridge- **FRS2**			
778	pINTRON2- **FRS2**			
779	pBS-**trk 105** (full)			
780	pBS-**MAGY** (5' UTR-400 bp)			
781	pCR2.1-**numb (PRR+)**			
782	pBridge- **FRS2-lexA-trkA**			
783	pCR2.1-**numb (PRR-)**			
784	pBS Tα1-**MAGY** (new start site)			
785	pINTRON2-**MAGY** (new start site			
786	pINTRON2- **FRS2**-nestin-C			
787	pINTRON2-**MAGY**-nestin-C			

	NAME	HOST Str	ANTIBIOTIC	SOURCE
788	pINTRON2-**NECDIN**-nestin-C	DH5α	amp	CJK
789	pGEX4T2-**MAM80** (PI3 kinase)	DH5α	amp	CJK
790	pCR2.1-**trk105 3' race (rat)**	DH5α	amp	CJK
791	pCR2.1- **FRS2-Y^{306}A mutant**	DH5α	amp	CJK
792	pCR2.1- **FRS2-Y^{349}A mutant**	XL10-gold	amp	CJK
793	pCR2.1- **FRS2-Y^{436}A mutant**	DH5α	amp	CJK
794	pCR2.1- **FRS2-Y^{471}A mutant**	DH5α	amp	CJK
795	pCR2.1-**FRS2-Y^{392}A** mutant	XL10-gold	amp	CJK
796	pGEX4T2	DH5α	amp	CJK
797	pNSE-lacZ	DH5α	amp	P. Danielson
798	pCR2.1-**FRS2-Y^{196}A** mutant	DH5α	amp	CJK
799	pBS-**NUMB (PRR+)**	DH5α	amp	CJK
800	pBS-**NUMB (PRR-)**	DH5α	amp	CJK
801	pBS-**trk105 5' race (rat)**	DH5α	amp	CJK
802	pBPX-HA-**TrkA S8/S9/Y^{679}A S27**	XL10-gold	amp	CJK
803	pIRES-EGFP-**FRS2-Y^{306}A**	XL10-gold	amp	CJK
804	pIRES-EGFP-**FRS2-Y^{436}A**	XL10-gold	amp	CJK
805	pIRES-EGFP-**FRS2-Y^{471}A**	XL10-gold	amp	CJK
806	cβS-FLAG	XL10-gold	amp	M. Cole
807	pBPX-HA-**trkA-Y^{679}A**	XL10-gold	amp	JM
808	pIRES-EGFP-**FRS2-Y^{196}A**	XL10-gold	amp	CJK
809	pIRES-EGFP-**FRS2-Y^{349}A**	XL10-gold	amp	CJK
810	pIRES-EGFP-**FRS2-Y^{392}A**	XL10-gold	amp	CJK
811	pBPX-HA-**TrkA T1**	XL10-gold	amp	CJK
812	pGEX4T2-**Mam282**	XL10-gold	amp	JM
813	pGEX4T2-**Mam156**	XL10-gold	amp	JM
814	pGEX4T2-**mouse1**	XL10-gold	amp	JM
815	pEBG-**trk47**	XL10-gold	amp	JM
816	pEBG-**trk86**	XL10-gold	amp	JM
817	pEBG-**trk5**	XL10-gold	amp	JM
818	pGEX-4T2 **Mam157**	XL10-gold	amp	JM
819	pGEX-4T2 **Mam171**	XL10-gold	amp	JM
820	pEBG **Peripherin**	XL10-gold	amp	CJK
821	pGEMT-**Rat-5'SNT Race**#2	XL10-gold	amp	CJK
822	pGEMT-**Rat-5'SNT Race**#3	XL10-gold	amp	CJK
823	pGEMT-Mouse-**5'FRS2 Race**#2	XL10-gold	amp	CJK **(in situ probe)**
824	pGEMT-Mouse-**5'FRS2 Race**#3	XL10-gold	amp	CJK
825	p38LoxpNeo (Frs2 KO vector 5' and 3' arms)	DH10	amp	Genome Systems
826	pBELO-BAC-**Som1** #1850	DH10	Chlorampen.	Genome Systems
827	pGEX4T1-**MAGY**	XL10-gold	amp	JV
828	Lox-Neo (lox-lox)	XL10-gold	amp	Bryce Vissel (Salk)
829	pEBG-**ras GRF $L^{263}Q$ mutant**	XL10-gold	amp	CJK
830	pBSTα1-**NUMB1 (++)**	XL10-gold	amp	CJK
831	pBSTα1-**NUMB3 (+-)**	XL10-gold	amp	CJK
832	pGEX4T2 **Mam122**	XL10-gold	amp	JM
833	pGEX4T1	XL10-gold	amp	CJK -- *NOT PRESENT??*
834	pGEX4T3	XL10-gold	amp	CJK

	NAME	HOST Str	ANTIBIOTIC	SOURCE
835	pEBG **Trk52**	XL10-gold	amp	JM
836	pBAKPAK8	XL10-gold	amp	Invitrogen
837	**SCG10** (pan neuronal marker - linearize w BamHI, transribe w SP6 for antisense)	XL10-gold	amp	JV/Andy Groves (Caltech)
838	pGEX4T2-**MAGY** (5'Bam band)	XL10-gold	amp	CJK
839	pZero-**MAGY** A14	XL10-gold	Zeocin	CJK
840	XL10-GOLD (competent cells-RKB)	XL10-gold	Chloramphen.	Stratagene
841	pAS2.1-**TrkB**(IC) RI/Sal	XL10-gold	amp	CJK
842	pBSTα1-Intron 2 XbaI	XL10-gold	amp	CJK
843	RET.INS.AP	XL10-gold	amp	D. Anderson, Caltech
844	RET.TK.AP	XL10-gold	amp	D. Anderson, Caltech
845	pEBG-**rasGRF cluster mutant** (Bam/Not)	XL10-gold	amp	CJK/P. Anborgh
846	pBPX-HA-TrkA S8/S9/Y^{679}**A**/Y^{683}**D**/Y^{684}**E (S30)**	XL10-gold	amp	JM
847	p38LoxpNeo-**MAGY** 1st arm Sac/RI	XL10-gold	amp	CJK
848	pGEXT-human **EG1**pcr (3' RACE3-REV2)	XL10-gold	amp	CJK
849	p38Loxpeo-**PITS** Knockout	XL10-gold	amp	CJK
850	pBABE-IRES-AP-PURO-**PITS** (RI)	XL10-gold	amp	CJK
851	pGEX4T2 **Mam65**	XL10-gold	amp	JM
852	pGEXT-human **EG1** 5' Race	XL10-gold	amp	CJK
853	TrkA KO (pJM4-3)	XL10-gold	amp	JV
854	TrkA 3' probe (129-105)	XL10-gold	amp	JV
855	TrkA 5' probe (pMM2)	XL10-gold	amp	JV
856				
857	pEBG-3	XL10-gold	amp	JM
858	pX^2Z-1 (lox-lox)	XL10-gold	amp	JV/MJ
859	pX^2Z-2(lox-lox)	XL10-gold	amp	JV/MJ
860	pX^2Z-3(lox-lox)	XL10-gold	amp	JV/MJ
861	plox-ex (lox-lox)	XL10-gold	amp	JV/MJ
862	pX^2Z-2-Sac1335 B-RI-RI-H (6848 bp) LoxP (cloned into Bgl II/H) contains mFRS-2 exons 1-4 <<pFLOX-1>>	XL10-Gold	amp	MJ (K.O.)
863	pZero Sac 1335 (**FRS-2** exons 1-4 Sac 1 fragments)	XL10-Gold	Zeocin	Genome Systems
864	pCDNA-HA-ShcB Y296A	XL10-Gold	amp	RKB
865	pCDNA-HA-ShcB Y296A & Y296A	XL10-Gold	amp	RKB
866	pCDNA-HA-ShcB Y296A,Y296A, Y372A	XL10-Gold	amp	RKB
867	pBS(SK)-Numb1 (++;$^{PTB/PRR}$) full length ~ 2.8 kb	XL10-Gold	amp	JV
868	pBS(SK)-Numb3 (+-;$^{PTB/PRR}$) full length ~ 2.8 kb	XL10-Gold	amp	JV
869	pBS(SK)-Numb12 (-+;$^{PTB/PRR}$) full length ~ 2.8 kb	XL10-Gold	amp	JV
870	pBS(SK)-Numb13 (--;$^{PTB/PRR}$) full length ~ 2.8 kb	XL10-Gold	amp	JV
871	pEBG-3Mouse1	XL10-gold	amp	JM

	NAME	HOST Str	ANTIBIOTIC	SOURCE
872	pBABE-puro-MAPKKH	XL10-gold	amp	Ferguson
873	pEFP-Ras615A	XL10-gold	amp	Ferguson
874	pBPX-HA-TrkAS8S9 $Y^{683}D:Y^{684}E$ **S32**	XL10-gold	amp	JM
875	pEBG-Mam 282	XL10-gold	amp	JM
876	pBABE-IRES-AP-PURO-**FRS2** + EGFP	XL10-Gold	amp	CJK –sequenced – missing last 5 aa
877	pBABE-IRES-AP-PURO-**FRS2 (PTB-Bam site)** + EGFP	XL10-Gold	amp	CJK
878	pBPX-HA-TrkA: $Y^{679}A:Y^{683}D:Y^{684}E$ **(S28)**	XL10-Gold	amp	JM
879	pEBG-Mam 156	XL10-gold	amp	JM
880	pEBG-Mam 171	XL10-gold	amp	JM
881	pEBG-Mam 122	XL10-gold	amp	JM
882	pEBG-Mam 103	XL10-Gold	amp	JM
883	pEBG-Mam 140	XL10-Gold	amp	JM
884	pCMX-HA-TrkA $Y^{679}A$ = S26	XL10-gold	amp	CJK
885				
886	pX^2Z-2 - Sac1335 B-H (3378 bp) LoxP (cloned Into BglII/H) contains **mFRS-2 exon 1** **<< pFLOX-2>>**	XL10-Gold	amp	MJ
887	pEBG-Mam 60	XL10-gold	amp	JM
888	pEBG-Mam 157	XL10-gold	amp	JM
889	pSE280	XL10-gold	amp	Invitrogen
890	p38LoxPNeo	XL10-gold	amp	Genome Systems
891	pX^2Z-2 - Sac1335 RI-H (2264 bp) LoxP	SCS110	amp	MJ
892	pBS-mouse-5'SNT Race	XL10-gold	amp	MJ
893	pSE280-2A	XL10-gold	amp	MJ
894	pBPX-HA-TrkA S9 (Alanine)	XL10-gold	amp	CJK
895	pSE280-1B-L (floxed neo)	XL10-gold	amp	MJ
896	pSE280-IB-R (floxed neo)	XL10-gold	amp	MJ
897	pEGFP-N2-PITS	XL10-gold	amp	CJK
898	pZero-C9 (FRS2 exons 2-5; RI fragment)	XL10-gold	amp	Genome Systems
899	----------	Stable-2	-------	Stratagene
900	----------	Sure	Kan	Stratagene
901	pGem-SNT1 3' UTR (mouse)	XL10-gold	amp	CJK
902	pAS211-PITS (5'Bam)	XL10-gold	amp	CJK
903= S31	pBPX-HA-TrkAS8 Y^{679}: $Y^{683}D:Y^{684}E$	XL10-Gold	amp	JM
904= S29	pBPX-HA-TrkAS9 Y^{679}: $Y^{683}D:Y^{684}E$	XL10-Gold	amp	JM
905	----------	XL1-Blue	Tet	Stratagene
906	pCR2.1-p16	XL10-gold	kan/amp	CJK
907	GST ATF-2		amp	J. Woodgett
908	GST c-Jun	BL21	amp	J. Woodgett
909	pBIG 2i HA TrkA clone #2	DH5-α	amp	E. Gryz
910	pBIG 2i HA TrkS13a clone #12	DH5-α	amp	E. Gryz
911	pBIG 2i HA TrkA13b clone #1	DH5-α	amp	E. Gryz
912	pBABE-IRES-AP-PURO p16	XL10-gold	amp	CJK

	NAME	HOST Str	ANTIBIOTIC	SOURCE
913	pCMX-HA Trk ($Y^{683}Y^{684}$→$F^{683}F^{684}$)	XL10-gold	amp	CJK
914	pCMX-HA Trk ($Y^{683}Y^{684}$→$N^{683}Q^{684}$)	XL10-gold	amp	CJK
915	pCMX-HA Trk ($Y^{683}Y^{684}$→$Q^{683}N^{684}$)	XL10-gold	amp	CJK
916	pEBG MAM 86	XL10-gold	amp	JM
917	pBSTα1-nestin express	XL10-gold	amp	CJK
918	pEGFP-N1-HA-Trk (full)	XL10-gold	amp	CJK
919	pEBG MAM 65	XL10-gold	amp	JM
920	pEGB MAM 249	XL10-gold	amp	JM
921	pEBG MAM 122	XL10-gold	amp	JM
922	- - -	XL1-BLUE MRA	- - -	Stratagene
923	- - -	XL1-BLUE MRA (P2)	- - -	Stratagene
924	pEFP-rasGRF1	XL10-gold	kan	Pieter/Ferguson
925	pMT3-ΔN158rasGRF1	XL10-gold	amp	Kato Kaelin/L Feig
926	pEGFP-N3-EG1 (human-full)	XL10-gold	kan	CJK
927	pMT3-1Q(-) - rasGRF1	XL10-gold	amp	Kato Kaelin/L Feig
928	pMT3-n-PH(-) - rasGRF1	XL10-gold	amp	Kato Kaelin/L Feig
929	pMT3-cc(-) - rasGRF1	XL10-gold	amp	Kato Kaelin/L Feig
930	pMT3-ΔN59 - rasGRF1	XL10-gold	amp	Kato Kaelin/L Feig
931	pBSKS-FRS2-E1<lox> (Xba/Spe frag.from pFLOX2)	XL10-GOLD	amp	MJ
932	pBSKS-floxNEO-L (RI fragment)	XL10-gold	amp	MJ
933	pBSKS-floxNEO-R (RI fragment)	XL10-gold	amp	MJ
934	pBSKS-FRS2-E1<lox>E2-4	XL10-gold	amp	MJ
935	pGEMT-EZ-mouseSNT3'UTR(polyA)	XL10-gold	amp	CJK
936	Tα1-nestin~overexpress-Numb1(++)	XL10-gold	amp	CJK
937	Tα1-nestin~overexpress-Numb3(+-)	XL10-gold	amp	CJK
938	pAS2.1-rat TrkC (ICD) SalI	XL10-gold	amp	CJK
939	pcDNA3.1 myc HIS A	XL10-gold	amp	Invitrogen
940	pcDNA3.1 myc HIS B	XL10-gold	amp	Invitrogen
941	pcDNA3.1 myc HIS C	XL10-gold	amp	Invitrogen
942	pBPXY-5-4	XL10-gold	amp	JM
943	pBPXY-30-5	XL10-gold	amp	JM
944	pBS(KS) MIG A (RI fragment 1900bp)	XL10-gold	amp	Mina/Ferguson
945	pSE280-rat TrkC HindIII/Xba	XL10-gold	amp	CJK
946	pEGFP-C2	XL10-gold	kan	Clontech
947	pAS2.1-EG1 (PRR)	XL10-gold	amp	CJK
948	pGBKT7-trkB (kinase dead)	XL10-gold	kan	CJK
949	pLNCXZ-HAWT trkA	XL10-gold	amp	CJK
950	pADTRACK-cmv-SNT1-myc HIS	XL10-gold	kan	CJK
951	pADTRACK-cmv-SNT2-myc HIS	XL10-gold	kan	CJK
952	**pCDNA3.1-mycHISB-som1 Y436A**	**XL10-gold**	**amp**	**MJ Wrong**
953	**pCDNA3.1-mycHIS-som1 y471a**	**XL10-gold**	**amp**	**MJ Wrong**
954	pCDNA-mouse1 (HA tagged)	HB101	amp	JM
955				

	NAME	HOST Str	ANTIBIOTIC	SOURCE
956	pCMX-HA-trkB (kinase dead)	XL-GOLD	amp	CJK
957	pEGFP-N1 DEL (no kozak or atg)	XL-GOLD	kan	CJK
958				
959	pEGFP-N3 DEL (no kozak or atg)	XL-GOLD	kan	CJK
960	PEGFP-N3	XL-Gold	Kan	ferguson
961	pEGFP-C1-EG1	XL-GOLD	kan	CJK
962	pEGFP-N(DEL Kozak)-FRS-2	XL-GOLD	kan	CJK
963				
964				
965				
966				
967				
968	pTag FRS2-Flag (clone 1)	DH5α	kan	KV
969	pIRES	DH5α	amp	clontech
970	pTag-FRS3-FLAG	DH5α	kan	KV
971	pGEMT-EZ-FRS3- 3' UTR (mouse)	XL-GOLD	amp	CJK **in vitro/insitu probes**
972	pEBG-MAM171 (full length)	XL-GOLD	amp	JM
973				
974	pCMX-HA-trkC (kinase dead)	XL-GOLD	amp	CJK
975				
976	pEGFP-C1-MAM171 (full length)	XL-GOLD	kan	CJK
977				
978	pCDNA-mouse1-mycHIS (NESCA)	XL-GOLD	amp	JM
979	pEGFP-N1 DEL (no kozak or ATG)	XL-GOLD	kan	CJK
980				
981				
982				
983				
984				
985				
986				
987				
988	pECFP-N1DEL-HA-trkA	XL-GOLD	kan	CJK
989				
990				
991				
992				
993	pcDNA3.1-mycHISA-human FRS3	XL-GOLD	amp	CJK
994				
995				
996				
997				
998				
999				
1000				
1001	pCDNA3.1-mycHIS-h**FRS2** (ptb)	XL10-gold	amp	KM
1002				

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1003				
1004				
1005				
1006				
1007				
1008				
1009				
1010				
1011				
1012	pEGFP-N3DEL-mouse1 (Nesca)	XL10-gold	kan	CJK
1013				
1014				
1015				
1016				
1017				
1018				
1019				
1020				
1021				
1022				
1023				
1024				
1025				
1026	pGAD10-trkB-ca24 (Casp/RIPK1-adapter)	HB101	amp	WL
1027	pGAD10-trkB-ca69 (Not 56-like)	HB101	amp	WL
1028				
1029	pGAD10-trkB-ca109 (PI-BIP Phosphatase)	HB101	amp	WL
1030	pACT2-trkB-br45 (Mouse/Human EsST 130882)	HB101	amp	WL
1031				
1032				
1033				
1034				
1035	pACT2-trkB-M2 (p66 Shc)	HB101	amp	WL
1036				
1037				
1038	pACT2-trkB-M38/Tid1	HB101	amp	WL
1039				
1040				
1041				
1042	pACT2-trkC-M38 (sck)	HB101	amp	WL
1043				
1044				
1045				
1046				
1047				

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1048	pACT2-trkC-M138 (Human BR2 cDNA)	XL10-gold	amp	WL
1049				
1050				
1051	pBPX-HA-trkA S21	XL10-gold	amp	CJK
1052				
1053	pCMX-HA-trkA ADE = S28	XL10-gold	amp	JM
1054	pCMX-HA-trkA S9 ADE = S29	XL10-Gold	amp	JM
1055	pCMX-HA-trkA S8/S9 ADE = S30	XL10-gold	amp	JM
1056	pAS1-mouse1 (Nesca)	XL10-gold	amp	JM
1057				
1058	pCR2.1-mouse1/Nesca (SH3minus)	XL10-gold	amp/kan	JM
1059				
1060				
1061				
1062				
1063	pEGFP-C1-Nesca (SH3 minus)	XL10-Gold	kan	JM
1064				
1065				
1066				
1067	pEGFP-N3DEL-nesca (2Y-A mutant)	XL10-gold	kan	CJK
1068	pGEX4T2-hSNT2 (PTB domain)	XL10-gold	amp	CJK
1069	pEBG-trkC-M38 (ShcB fragment)	XL10-gold	amp	WL
1070	pCDNA-HA-EG1 (deleted PEST)	XL10-gold	amp	JM
1071	pGEX4T2-nesca (SH3-)	XL10-gold	amp	JM
1072	pTADV-EG1-deleted PEST	XL10-gold	amp/kan	JM
1073	pTADV-MAM171-EE (Tao1, EE mutant)	XL10-gold	amp/kan	JM
1074	pEGFP-N3DEL-human FRS3	XL10-gold	kan	CJK
1075				
1076	FRS-2 KO1 1-0	XL10-gold	amp	JM
1077				
1078				
1079				
1080				
1081	nestin Xpress-LACZ	XL10-gold	amp	CJK
1082				
1083				
1084	pEBG3- trkB-M34 (human EST)	XL10-gold	amp	WL
1085	pEBG3-trkB-M38 ($Tid1_L$-L^{215}~S^{480})	XL10-gold	amp	WL
1086	pEBG3- trkB-M53 (K1AA1091)	XL10-gold	amp	WL
1087	pEBG3- trkB-br45 (Mouse/Human EsST 130882)	XL10-gold	amp	WL
1088	pEBG3- trkC-M94 (Kruppel, HKR3)	XL10-gold	amp	WL
1089	pEBG3-trkC-M109	XL10-gold	amp	WL
1090	pEBG3- M138 (Human BR2 cDNA)	XL10-gold	amp	WL
1091	pEBG3-trkC-M29 (TSC-22)	XL10-gold	amp	WL

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1092	pCDNA3.1 (+)-HA-sck (ShcB)	XL 0-gold	amp	L
1094	pRFP-N1DEL (no kozak or ATG)	XL 0-gold	kan	JK
1095	pRFP-N2DEL (no kozak or ATG)	XL 0-gold	kan	JK
1096	pRFP-N3DEL (no kozak or ATG)	XL 0-gold	kan	JK
1097	pCDNA-HA-MAM171 (Tao1, EE mutant)	XL 0-Gold	amp	I
1107	pEGFP-N1del-Nesca (SH3minus)	0-gold	kan	
1108	pIRES2-EGFP-**FRS2**-mycHIS	0-gold	kan	JK—7th last aa has a silent mutation
1109	pIRES2-EGFP-**FRS3**-mycHIS	XL 0-gold	kan	JK
1110	pRFP-N3DEL-**FRS2**	XL 0-gold	kan	JK
1111	pCDNA-IRS1 (PH-PTB)		amp	XB
1113	FGFR1.1	XL 0-gold	amp	
1114	trkC-ca17 (Unknown interactor)	XL 0-gold	amp	L
1115	pEGB3-Mam212 (Synapse Associated Protein)	-Gold	amp	L
1116	KO1.1	XL 0-gold	amp	JK
1117	KO2	XL -gold	amp	JK
1118	pGEX4T2-nesca(SH3 minus)		amp	
1119				
1120	pcDNA3.1-mouse FGFR-1	XL 0-gold	amp	JK
1121	pGEM-T-Easy-FRS2 (mychis) NheI/SelI		amp	XB – *Duplicated T7 Site*
1122	pGEM-T-Easy- FRS2 (mychis) NheI/SelI		amp	XB -- *Duplicated T7 Site*
1123	pGEM-T-Easy-FRS2 (flag) NheI/SelI		amp	B -- *Duplicated T7 Site*
1124				
1125	pEGFP-N2DEL-**FRS2 (PTB domain)**	-gold	kan	M
1126		0-gold		
1127	pNESTIN-**hFRS2**-mycHIS-IRES-LACZ	-g	amp(lac Z not in frame)	K stops at 5'mychis
1128	pNESTIN-**hFRS3**-mycHIS-IRES-LACZ	-g	amp (lac Z not in frame)	K stops at 5'mychis
1129				
1130	DS Red pRFP-N1DEL-HA-trkA	-gold	kan	K
1131	pBIG2i	0-g	amp	K/Strathdee

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1132	pBIG2i-**hFRS2**-mycHIS	XL10-gold	amp	CJK
1133	pBIG2i-**hFRS2**-RFP	XL10-gold	amp	CJK
1134	pBIG2i-**hFRS3**-mycHIS	XL10-gold	amp	CJK
1135	pBIG2i-**hFRS3**-EGFP	XL10-gold	amp	CJK
1136				
1137				
1138	pBIG2i-**Nesca**-EGFP	XL10-gold	amp	CJK
1139	pAS1-trkA (S8/Y679A/S9)=S27	XL10-gold	amp	JM
1140	pAS1-trkA (S8/Y679A/Y683D/Y684E)	XL10-gold	amp	JM
1141	pAS1-trkA (S9/Y679A/ Y683D/Y684E)=S29	XL10-Gold	amp	JM
1142	pGEMT-MAM171 **(human Tao1)**	XL10-Gold	amp	JM
1143	pBIG2i-Nesca(SH3-)-EGFP	XL10-gold	amp	CJK
1144	pBPX-GST-Nesca	XL10-gold	amp	
1145	pBPX-GST-Nesca (SH3minus)	XL10	amp	JM
1146	pBlueBac HISA	XL10	amp	Invitrogen
1147	pBlueBac HISB	XL10	amp	Invitrogen
1148	pBlueBac HISC	XL10	amp	Invitrogen
1149	pGAD-Nesca	XL10	amp	JM
1150	pGAD-Nesca (SH3-)	XL10	amp	JM
1151	pAS-2-trkA-S1	XL10	amp	JM
1152	pAS-2-trkA-S3	XL10	amp	JM
1153	pAS-2-trkA-S8	XL10	amp	JM
1154	pAS-2-trkA-S9	XL10	amp	JM
1155	pAS-2-trkA-S11	XL10	amp	JM
1156	pAS-2-trkA-S19	XL10	amp	JM
1157	pAS-2-trkA-Y679A/Y683D/Y684E (S28)	XL10	amp	JM
1158	pBackpack8 FRS3-mycHIS	XL10	amp	JM
1159	pGADT7	XL10	amp	Clontech
1160	pGADT7-T	XL10	amp	Clontech
1161	pBP-8-nesca-mycHIS	XL10	amp	JM
1162	pACTII-nesca screen 1	XL10	amp	JM
1163	pACTII-nesca screen 11 (DRAL)	XL10	amp	JM
1164	pACTII-nesca screen 15 (GRAP)	XL10	amp	JM
1165	pACTII-nesca screen 26 (STAT3)	XL10	amp	JM
1166	pACTII-nesca screen 31 (PP6)	XL10	amp	JM
1167	pACTII-nesca screen 38 (RIL)	XL10	amp	JM
1168	pACTII-nesca screen 43 (ACK)	XL10	amp	JM
1169	pACTII-nesca screen 46 (NPCIP)	XL10	amp	JM
1170		XL10	amp	
1171	pGEX-SHC (full length)	Bl21 RIL	amp	Chris/JM
1172	pGEX-SHC (full length)	BL21 RP	amp	Chris/JM
1173	pGEX-SHC (PTB)	BL21 RP	amp	JM
1174	pGEX-SHC (PTB)	BL21 RIL	amp	JM
1175	pCDNA-M171 (RI/SacII)		amp	JM
1176	pEBG P68		amp	
1177		XL10gold	chloramphenicol	Stratagene
1178	pCDNA-HA-N31 (PP6)	XL10Gold	amp	JM

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1179	pCDNA-HA-N38 (RIL)	XL10Gold	amp	JM
1180	pCDNA-HA-N43 (Ack1)	XL10Gold	amp	JM
1181	pACTII-NUP214 (nesca1)	XL10Gold	amp	JM
1182	pGEX4T2 PP6 (nesca31)	BL21 RP	amp	JM
1183	pGEX4T2 RIL (nesca 38)	BL21 RP	amp	JM
1184	pGEX4T2 ACK (nesca43)	BL21 RP	amp	JM
1185	pGEX4T2 PP6 (nesca31)	XL10Gold	amp	JM
1186	pGEX4T2 RIL (nesca 38)	XL10Gold	amp	JM
1187	pGEX4T2 ACK (nesca 43)	XL10Gold	amp	JM
1188	pAS2.1-trkA S21	XL10Gold	amp	CJK/SJD
1189	pAS2.1-FRS3 (PTB)	XL10Gold	amp	CJK/SJD
1190	pAS2.1-FRS3 (full length)	XL10Gold	amp	SJD remade 06/02 good!
1191	pAD Track	XL10Gold	kan	L. Dagnino
1192	pAD Track-CMV	XL10Gold	kan	L. Dagnino
1193	pAD Easy-1	XL10Gold	amp	L. Dagnino
1194	pIND TID1/Long	DH5α	amp	K. Munger
1195	pIND TID1/Short	DH5α	amp	K. Munger
1196	pBP EG-1	XL10Gold	amp	JM
1197	pBP GST Peripherin	XL10Gold	amp	JM
1198	pGEX4T2 NUP214 (nesca1)	XL10Gold	amp	JM
1199	pLNCX2	XL10Gold	amp	Clontech
1200	pEBG3-trkB-ca43 (unknown)	XL10Gold	amp	HYL
1201	pEBG3-trkB-ca69 (Not 56-like)	XL10Gold	amp	HYL
1202	pEBG3-trkB-ca73 (Unknown)	XL10Gold	amp	HYL
1203	pEBG3-trkB-ca81 (Unknown)	XL10Gold	amp	HYL
1204	pEBG3-trkB-ca109 (PI-BIP-phosphatase)	XL10Gold	amp	HYL
1205	pBridge #2	DH5 α	amp	KV (clontech)
1206	pIND/Pme1 (-)	XL10Gold	amp	HYL
1207	pGEXK6-Pin1	DH5α	amp	SJD
1208	pEBG3-trkB-ca24 (Casp/RIPK1 adapter)	DH5α	amp	HYL
1209	pEBG3-trkB-ca123 (Unknown)	DH5α	amp	HYL
1210				
1211	PGADT7 – TRK – KD	XL10Gold		
1212	pEBG DRAL (nesca 11)	XL10Gold	amp	JM
1213	pEBG PP6 (nesca 31)	XL10Gold	amp	JM
1214	pEBG RIL (nesca 38)	XL10Gold	amp	JM
1215	pEBG ACK (nesca 43)	XL10Gold	amp	JM
1216	pADEasy HA trkA #4	XL10Gold	kan	EG
1217	pADEasy HA S11b #6	XL10Gold	kan	EG
1218	pADEasy HA S13a #4	XL10Gold	kan	EG
1219	pADEasy HA S13b #1	XL10Gold	kan	EG
1220	pGEX2T-GST **Suc1 (p13)**	DE3-RP	amp	SJD
1221	pLNCX2-mycHIS-nesca	XL10Gold	amp	JM
1222	pLNCX2-nesca-R	XL10Gold	amp	JM
1223	pEGB3-trkB-ca91 (937223 Unknown cDNA)	DH5α	amp	HYL
1224	pLNCX2-EGFP-nesca	XL10Gold	amp	JM

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1225	pAS2.1 **CKS1**	XL10Gold	amp	JM
1226	pAS2.1 **CKS2**	XL10Gold	amp	JM
1227	pRK5-myc-hTID1$_L$	DH5α	amp	Hui-Yu Liu
1228	pRK5-myc-hTID1$_S$	DH5α	amp	Hui-Yu Liu
1229	pAD Easy TrkA clone 5 (doesn't work)	XL10Gold	kan	EG
1230	pAD Easy S11b clone 6	XL10Gold	kan	EG
1231	pAD Easy S13a clone 4	XL10Gold	kan	EG
1232	pAD Easy S13b clone1	XL10Gold	kan	EG
1233	pCDNA HA DRAL (nesca 11)	XL10Gold	amp	JM
1234				
1235	pGADT7 **CKS1**	XL10Gold	amp	JM
1236	pGADT7 **CKS2**	XL10Gold	amp	JM
1237	pACT II nesca 46 (NPIP)	XL10Gold	amp	JM
1238	pGEMT trkA CAV(-) frag	XL10Gold	amp	JM
1239	pcDNA HA NUP214 (nesca1)	XL10Gold	amp	JM
1240	pcDNA HA NPIP (nesca 46)	XL10Gold	amp	JM
1241	pcDNA HA GRAP1 (nesca 15)	XL10Gold	amp	JM
1242	pcDNA mycHIS **CKS1**	XL10Gold	amp	JM
1243	pGEMT M171 EE Frag Xba-SalI			James
1244	pGEX-**ENIGMA** (LIM1,2,3)	XL10Gold	amp	Gordon Gill
1245	pGEX-**ENIGMA** (LIM1,2,3)	BL21-RP	amp	Gordon Gill
1246	pGEMT **Tao1** Xba/Sal EE frag	XL10Gold	amp	JM
1247	pcDNA mycHIS **CKS1**	XL10Gold	amp	JM
1248	pcDNA mycHIS **CKS2**	XL10Gold	amp	JM
1249	pcDNA HA PP6	XL10Gold	amp	JM
1250	pEBG nesca 1 (NUP214)	XL10Gold	amp	JM
1251	pEBG nesca 46 (NPIP)	XL10Gold	amp	JM
1252	pEYFP-N1	DH5α	amp	CLONTECH
1253	pRFP-N1DEL HA trkA S8	XL10Gold	kan	CJK
1254	pRFP- N1DEL HA trkA S17	XL10Gold	kan	CJK
1255	pGEMT-EZ-200bpfrag nestin coding	XL10Gold	amp	KMcD
1256	pRFP-N1DEL-HA trkA S9	XL10Gold	kan	CJK
1257	pGEMT PP6	XL10Gold	amp	JM
1258	pGEMT trkA CAV(-) Neo/Kpn frag	XL10Gold	amp	JM
1259	pBKS PP6	XL10Gold	amp	JM
1260	pGEX4T2 GRAP (nesca 15)	BL21 RP	amp	JM
1261	pGEX4T2 NPIP (nesca 46)	BL21 RP	amp	JM
1262	pEGFP SH3 (nesca)	XL10Gold	kan	JM
1263	pCMX HA trkA CAV(-)	XL10 Gold	amp	JM
1264	pcDNA3.1-HA-TrkC-m138 (Br2 cDNA Human EST)	DH5α	amp/neo	HYL
1265	pEYFP-C1	DH5α	amp/neo	Clontech
1266				
1267				
1268	pRK5-myc-ShcB[SH2(-)]	DH5α	amp	HY Liu
1269	pGEX4T2 nesca 15 (GRAP)	XL10Gold	amp	JM
1270	pGEX 4T2 nesca 46 (NPIP)	XL10Gold	amp	JM

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1271	pBKS M171 HA	XL10Gold	amp	JM
1272	pBKS HA M171 EE	XL10Gold	amp	JM
1273				
1274	pEGFP N1 nesca DEL1	XL10Gold	kan	JM
1275	pEGFP N1 nesca DEL2	XL10 Gold	kan	JM
1276	pAS2.1 trkA T1	XL10Gold	amp	JM
1277	pACTII nesca screen 50 (ART-4)	XL10Gold	amp	JM
1278	pcDNA HA nesca screen 50 (ART-4)	XL10Gold	amp	JM
1279	pcDNA HA nesca screen 26 (STAT3)	XL10Gold	amp	JM
1280	pAD Track-human FRS3-$Y^{417}A$ (no tags)	XL10 Gold	kan	SJD
1281	pAD Track-human FRS3-$Y^{485}A$ (no tags)	XL10 Gold	kan	SJD
1282	PAdTrack FRS3-1A mutant	DH5α	amp	KV
1283				
1284				
1285	pGEX4T2 trkB-m38 (Tid1 part)	DH5α	amp	HYL
1286	pFLAG-CMV2-NPIP	XL10Gold	amp	Graham Goodwin
1287	pcDNA mycHIS A nesca 50 (ART-4)	XL10Gold	amp	JM
1288	pCMX trkA CAV(-) $W^{714}A$ $F^{716}A$	XL10Gold	amp	JM
1289	pRK5 myc SH2-B	XL10 Gold	amp	Ginty
1290	pMT 35 CMV-cjun-HIS_6	?	amp	D. Bowman
1291	pMT 107 CMV-ubiquitin-HIS_6	?	amp	D. Bowman
1292	pMT 123 CMV-ubiquitin-HA	?	amp	D. Bowman
1293	pGEX4T2-Sck (ShcB)	DH5α	amp	Hui-Yu Liu
1294	pCMX trkA $W^{721}A$ $F^{724}A$	XL10Gold	amp	JM
1295	pACT II Nesca 40	XL10Gold	amp	JM
1296	pAd Track Trk A-1	XL10 Gold	kan	EG
1297	pAd Track Trk A-3	XL10 Gold	kan	EG
1298	pAd Track Trk A-6	XL10 Gold	kan	EG
1299	pAd Track Trk A-8	XL10 Gold	kan	EG
1300	pAd Track Trk A-11	XL10 Gold	kan	EG
1301	pGEX 4T2 SH2B-1	BL21 RIL	amp	EG
1302	pGEX4T2 SH2B-7	BL21 PR	amp	EG
1303	pGEX4T2 SH2B-7	BL21 RIL	amp	EG
1304	BJ5183 (cells for AV work)		strp	Dr. Dagnino
1305	pEGFP HA nesca	XL10 gold	kan	JM
1306	pcDNA cdc42 T17N **(DN)**	DH5α	amp	R. Cerione
1307	pcDNA cdc42 Q61L **(CA)**	DH5α	amp	R. Cerione
1308	pcDNA cdc42	DH5α	amp	R. Cerione
1309	pcDNA HA-ACK2	JM109	amp	R. Cevione
1310				
1311	pGEX4T2-hTid-1	BL21 RIL	amp	Hui-Yu Liu
1312	pGBKT7	XL10 gold	kan	Clontech
1313	pLNCX2-**FRS2**-EGFP (DEL kozak)	XL10 gold	amp	SJD sequenced –missing last 5 aa
1314	pLNCX2-**FRS3**-EGFP (DEL kozak)	XL10 gold	amp	SJD
1315	pEGFP nesca DELNLS	XL10 gold	kan	JM

	NAME	I	)ST Str	ANTIBIOTIC	SOURCE
1316					
1317					
1318	pAS2.1 nesca DEL1	X	10 gold	amp	JM
1319	pAS2.1 Nesca DEL2	X	0 gold	amp	JM
1320	pGADT7 trkA T1	X	10 gold	amp	SJD
1321	pXJ4-HA- ACK1	X	10 Gold	amp	Ed Manser
1322	pGEXIN SH3 (N)	P	21	amp	JM
1323	pGEX FRS3-PTB	J	21RIL	amp	JM
1324	pGEX FRS3-PTB	L	21RP	amp	JM
1325	pRK5 EGFR	H	3101		Mike Moran
1326	pECL-v-src	X	10Gold		Mike Moran
1327	pLNCX2-EGFP	X	0gold	amp	SJD
1328	pMT3-Cat (**RasGRF**)	X	0gold	amp	colin (L.Feig)
1329	pMT3-PH1-Cat (**RasGRF**)	X	10gold	amp	colin (L.Feig)
1330	pMT3-PH1-CC-Cat (**RasGRF**)	X	10gold	amp	colin (L.Feig)
1331	pMT3-PH1-CC-IQ-Cat (**RasGRF**)	X	10gold	amp	colin (L.Feig)
1332	pGEX4T1 Filamin	J	.1 RP	amp	JM
1333	pGEX4T1 Filamin	J	101	amp	JM
1334	pGEX4T3 NPIP	J	21 RP	amp	JM
1335	pGEX4T3 NPIP	J	01	amp	JM
1336	pLNSX-EGFP	X	0gold	amp	SJD
1337	pSV42	X	0gold	amp	SJD
1338	pHM840	J	01	amp	T. Stamminger
1339	pHM829	J	01	amp	Jane Rylett
1340	pAdTrack TrkA-1		01	kan	EG
1341	pAdTrack TrkA-3	J	01	kan	EG
1342	pAdTrack TrkA-5	J	01	kan	EG
1343	pAdTrack TrkA-7		01	kan	EG
1344	pAdTrack TrkA-11	J	1	kan	EG
1345	Ap2 Retrovector		old	amp	Jacques Galipeau
1346	pcDNA FGFR1(Xho frag)			amp	JM
1347	pcDNA myc FGFR		01	amp	JM
1348	pAdTrack trkC kinase dead		01	kan	CR
1349	pAdTrack trkC-3		01	kan	CR
1350	pAdTrack trkC-4		01	kan	CR
1351	pAdEasy Trk A5-4		3	kan	EG
1352	pAdEasy Trk A5-5		3	kan	EG
1353	pAdEasy Trk A5-6			kan	EG
1354	pAdEasy Trk A7-3		3	kan	EG
1355	pAdEasy Trk 7-4		3	kan	EG
1356	pAdEasy Trk A-18		3	kan	EG
1357	pcDNA3.1-mycHIS-'A'-human FRS3 $Y^{192}A/Y^{322}A$ Grb2 sites mutant			amp	SJD
1358	pEBG R Ras GRF2			amp	CJR (P.Anborg)
1359	pEFP R RasGRF2			kan	CJR (P.Anborg)
1360	pMT3 RasGRF-GAP-PH			amp	CJR (L.Feig)
1361	pGAD424-human FRS3			amp	SJD
1362	pAdEasy TrkC 20			kan	CR

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1363	pAdEasy TrkB (kinase dead) clone 11	HB101	kan	CR
1364	pAdEasy TrkB (kinase dead) clone 18	HB101	kan	CR
1365	pAdEasy TrkB clone 20	HB101	kan	CR
1366	pAdEasy TrkB (kinase dead) clone 12	HB101	kan	CR
1367	pAdEasy TrkB clone 17	HB101	kan	CR
1368	pHM829 nesca NLS short	HB101	amp	JM
1369				
1370				
1371	pcDNA3.1-mycHIS-human **FRS3** -$Y^{322}A$ (Grb2'C')	HB101	amp	SJD
1372	pcDNA3.1-mycHIS-human **FRS3** - $Y^{192}A/Y^{287}A/Y^{322}A$ "3A"	HB101	amp	SJD
1373	pHM829 nesca NLS long	HB101	amp	JM
1374	Ngn1 (neurogenin 1) new plasmid	HB101	amp	D. Anderson
1376	Ngn2 (neurogenin 2)	HB101	amp	D. Anderson
1377	pGEMT Easy myc-FGFR 5'frag PCR	HB101	amp	JM
1378	pGEMT Easy HA-EGFR 5'frag PCR	HB101	amp	JM
1379	pGEMT Easy **hFRS3**-$Y^{417}A/Y^{455}A$ "2A"	HB101	amp	SJD
1380	pcDNA3.1 mycHISA-**hFRS3**-$Y^{417}A/Y^{455}A$ "2A"	HB101	amp	SJD
1381	pGAD424-human **FRS3**-PTB	HB101	amp	SJD
1382	pGEX4T2-Sck-CH1	BL21 RIL	amp	Hui-Yu Liu
1383	pGAD424-human **FRS2**-PTB	HB101	amp	SJD
1384	pEGFP N1 nesca DEL A	HB101	kan	JM
1385	pEGFP N1 nesca DEL B	HB101	kan	JM
1386	pEGFP N1 nesca DEL C	HB101	kan	JM
1387	pEGFP N1 nesca DEL D	HB101	kan	JM
1388	Mo/FGFR 3-1 cQ	HB101	amp	D. Ornitz *-need his*
1389	Mo/FGFR 3 iii c myc	HB101	amp	D. Ornitz *permission*
1390	pCS FGFR 3 iii c myc	HB101	amp	D. Ornitz *to use*
1391	pCS FGFR 2 iii b myc	HB101	amp	D. Ornitz *these*
1392	pcDNA mychisC-human **FRS3**-PTB	HB101	amp	SJD
1393				
1394	pcDNA 3.1 mychis FGFR1	HB101	amp	JM
1395	LTR2HX FGFR4*contains a 3kb Bgal	HB101	amp	Alitalo*recloned pcDNA glyc 1457
1396				
1397	pGEMT-EZ-human **FRS3**- $Y^{192}A/Y^{287}A/Y^{322}A/Y^{455}A$ "4A"	HB101	amp	SJD
1398	pcDNA3.1-FGFR4 *not in frame*	HB101	amp	subcloned from Dr. Alitalo's FGFR4 *new gyc 1457
1399	pcDNA3.1-myc HISA- human **FRS3** $Y^{192}A/Y^{287}A/Y^{322}A/Y^{455}A$ ('4A')		amp	SJD
1400	pcDNA3.1 FGFR 2 * fusion to FRS3*	HB101	amp	subcloned from Ornitz 1391 - need permission
1401	pGBKT7-p53	?	kan	Clontech
1402	pGADT7-SV40 large T antigen	?	amp	Clontech
1403	pGEMT-FRS3 mouse	?	amp	JV
1404				

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1405	pGEX-Crk SH2	BL21	amp	L.Z.
1406	pGEX-Crk full length	BL21	amp	LZ
1407	pEGFP N1 nesca DEL 3-4	DH5a	kan	JM
1408	Ap2-h**FRS3**-myc	HB101	amp	SJD
1409				
1410	pRK5-myc-**Tid1** DNA J Domain	XL10 Gold	amp	Hui-Yu Liu
1411	pcDNA3.1-myc-HIS-B_**p66ShcA** (human)	XL10 Gold	amp	HYL
1412	Ap2-h**FRS3**-PTB (myc?)	HB101	amp	SJD
1413	Mo/FR1/IRES	HB101	amp	D. Ornitz
1414	pEGFPN1 Nesca Del 1-2	HB101	kan	JM
1415	pEGFPN3 Nesca NES1	HB101	kan	JM
1416	pEGFPN3 Nesca NES2	HB101	kan	JM
1418	pEGFPN3 Nesca NES3	HB101	kan	JM
1419	pGEX2T-C-**Raf RBD**	HB101	amp	Hermann
1420	pGEX2T-**RalGEF RBD**	HB101	amp	"
1421	pRK5-myc-mShcC p55 (Sal-Not)	DH5a	amp	Hui-Yu Liu
1422	pRK5-myc-mShcC p55 (Sal-Bam)	DH5a	amp	HYL
1423	pGEX4T2-Sck full	BL21 RP	amp	HYL
1424	pGEX4T2-Sck full	BL21 RIL	amp	HYL
1425	pGEX4T2-Sck PTB	BL21 RP	amp	HYL
1426	pGEX4T2-Sck PTB	BL21 RIL	amp	HYL
1427	pGEX4T1-Sck SH2	BL21 RP	amp	Hui-Yu Liu
1428	pGEX4T1-Sck SH2	BL21 RIL	amp	HYL
1429	pGEX4T2-Sck CH1	BL21 RP	amp	HYL
1430	pGEMTeasy hSNT2 '5A' Y192A, Y287A, Y322A, Y417A, Y455A ('5A' mutant)	HB101	amp	SJD
1431	pEGFP N1del hTid1L (new 4/23/03)	XL10 gold	kan	Hui-Yu Liu
1432	pEGFP N1 Nesca A118	HB101	kan	JM
1433	pEGFP N3 Nesca NES 205	HB101	kan	JM
1434	pGEMT-EZ FRS2-2MG hSNT1 transgene	XL10 gold	amp	KMcD
1435	pGEMT-EZ FRS2-1X1 hSNT1 transgene	XL10 gold	amp	KMcD
1436	pGEMT-EZ FRS2-21Y1 hSNT1 transgene	XL10 gold	amp	KMcD
1437	pGEMT murine Nesca Carb. term.	XL10 gold	amp	JM
1438	pGEMT murine Nesca NH4 term.	XL10 gold	amp	JM
1439	pcDNAmychis hSNT1 transgene FRS2-21Y1	XL10 gold	amp	KMcD
1440	pcDNAmychis hSNT1 transgene FRS2-1X1	XL10 gold	amp	KMcD
1441	pAS2.1 p13 HA	HB101	amp	SJD
1442	pGEX PAK CRIB	DH5?	amp	Bosco
1443	pGEX PBD	DH5?	amp	Bosco
1445	pcDNAmychis hSNT1 transgene FRS2-2MG	XL10 gold	amp	KMcD
1446	pGEMT murine Nesca full length	HB101	amp	JM
1447	pEGFP N3 KSRR	HB101	amp	JM
1448	pET-3d	HB101	amp	JM
1449	pGEMT easy RasGRF1	XL10 gold	amp	K Manto
1450	pDs-RED N1 NPIP	HB101	kan	JM
1451	p5X-NF κB-Luc	DH5α	amp	Dr. Greene, W.C., UCSF

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1452	pGAD T7 P75 ICD	XL10 gold	amp	CJK
1453	pET-3d Nesca	BL21 De3 RP	amp	JM
1454	pET-3d-FRS2-PTB	BL21 De3 RP	amp	SJD
1455	pET-3d-FRS3-PTB	BL21 De3 RP	amp	SJD
1456	pLXSH c-Cbl	XL10 gold	amp/hygro	Dr. Bisson, U. Calgary
1457	pcDNA3.1(c) - FGFR4 (no myc)	XL10gold	amp	KMcD
1458	pEGFP-calmodulin	XL10gold	kan	(Andy) S. Ferguson
1459	pMT-HA-ubiquitin	XL10 gold	amp	J. Wrana (S. Bonni)
1460	pCMV5-BIF1	XL10 gold	amp	J. Wrana
1461	pSUPER	DH5α	amp	Dr. R. Agami (see note)
1462	pVgRXR	DH5α	Zeo	Invitrogen
1463	pGEMT mouse NESCA 2	XL10 gold	amp	JM
1464	pGEMT mouse NESCA 3	XL10 gold	amp	JM
1465	pGEMT mouse NESCA 4	XL10 gold	amp	JM
1466	~~pcDNA myc his B Rasgrf1~~	~~XL10 gold~~	~~amp~~	~~K Manto~~ *No Myc KR*
1467	pIND-HA-trkA WT (Not1 deleted)	DH5α	amp	Hui-Yu Liu
1468	pSuper-anti-Tid1 control	XL10 gold	amp	Hui-Yu Liu
1469	pY16	DH5 α	amp	C. Kappen
1470	pY16	XL10 gold	amp	C. Kappen
1471				
1472	pcDNA- Gab-2 (HA tag)x1 fused directly to the start codon	XL10-gold	amp	GenFeng MTA signed do not distribute
1473	pcDNA-HA-Gab-1 (3x HA tag)	XL10-gold	amp	GenFeng MTA signed-do not distribute
1474	PGEMTEZ-Rasgrf2 3' end BstX1-Xhol PCR fragment	XL10-gold	amp	K. Manto
1475	PLHCX	DH5 α	amp/Hygro	Hui-Yu Liu
1476	pDS-Red ERK2	HB101	Kan	Ferguson
1477	pEGFP-Erk2	HB101	Amp	Ferguson
1478	p20 – Nestin (human) intronic region	XL10 Gold	Amp	C. Kappen (do not distribute)
1479	p20 – Nestin (human) intronic region (back-up)	XL10 Gold	Amp	C. Kappen (do not distribute)
1480	pCMV-Tag4A (FLAG cloning vector)	XL10 Gold	Kan	Stratagene
1481	pCMV-Tag4A (back-up)	XL10 Gold	Kan	Stratagene
1482	pCMV-Tag4B (FLAG cloning vector)	XL10 Gold	Kan	Stratagene
1483	pCMV-Tag4B (back-up)	XL10 Gold	Kan	Stratagene
1484	pCMV-Tag4C (FLAG cloning vector)	XL10 Gold	Kan	Stratagene
1485	pCMV-Tag4C (back-up)	XL10 Gold	Kan	Stratagene
1486	pCMV5L-His6-HA-Erk1	DH5-alpha	Amp	Dr. M. Cobb (do not distribute)
1487	pMCL-MKK1 (R4F) Ha-tagged	DH5-alpha	Amp	Dr. N. Ahn (do not distribute)
1488	pGADT7-trk ICD	XL10 Gold	Amp	A. Vilos
1489	pET28a	HB101	Kan	Novagen

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1490	pCAGGS	DH5-α	Amp	Jun-Ichi Miyayaki (do not distribute)
1491	phrGFP	HB101	Amp	Stratagene
1492	AP2-hTid1s-myc	DH5-alpha	Amp	Hui-Yu Liu
1493	pGemT-Run I (Nesca)	XL10 Gold	Amp	James
1494	pGemT-Run II (Nesca)	XL10 Gold	Amp	James
1495	pET28a-Run I (Nesca)	BL21 DE3 RIL	Kan	James
1496	pET28a-Run II (Nesca)	BL21 DE3 RIL	Kan	James
1497	pcDNA-MycHis-FGFR4 (full)	XL10 Gold	Amp	Kathy McD.
1498	pGex4T2-ShcC-PTB (mouse)	BL21 DE3 RIL	Amp	Hui-Yu Liu
1499	pGemT-mouse Nesca- 3' UTR	XL10 Gold	Amp	Fahreen/James
1500	pZeo-myc-Tid1S	XL10 Gold	Zeo	Chunhui Li/HYL
1501	pGex4T2-Mek	BL21 DE3 RIL	Amp	James
1502	pGex4T2-Erk2	BL21 DE3 RIL	Amp	James
1503	pMT-HA-ubiquitin	XL10 Gold	Amp	A. Vilos (back-up)
1504	pcDNA-hrGFP (ATG Deleted)	HB101	Amp	James
1505	pcDNA-HA-EGFR	XL10 Gold	XL10 Gold	James
1506	BL21 DE3 RIL Codon Plus bacteria	BL21 DE3 RIL	Chloramph.	Stratagene
1507	pCEP4L Control vector for MKK1 R4F	DH5-α	Amp	Dr. N. Ahn (do not distribute)
1508	pEGFP-C2-Rab5	DH5-α	Kan	Steve Fergusson
1509	pProExHTa	DH5-α	Amp	Invitrogen
1510	pCMX-TrkA-S11b (No tag)	XL10 Gold	Amp	L. Zhou
1511	pCMV5L-His6-HA-Erk2 ($L^{73}P,S^{151}D$)	XL10 Gold	Amp	Dr. N. Ahn (do not distribute)
1512	pEYFP-Golgi	XL10 Gold	Kan	Clontech
1513	pEYFP-Endoplasmic reticulum	XL10 Gold	Kan	Clontech
1514	pEYFP-mitochondria	XL10 Gold	Kan	Clontech
1515	pGex-Pak		Amp	N. Lamarche-Vane
1516	pGex-WASP		Amp	N. Lamarche-Vane
1517	pZeo-myc-Tid1L	XL10 Gold	Zeo	Hui-Yu Liu
1518	pZeo-anti-Tid1 (control)	XL10 Gold	Zeo	Hui-Yu Liu
1519	pEGFP-Golgi	XL10 Gold	Kan	Hui-Yu Liu
1520	pGex4T2-N part of Tid1L	BL21 DE3 RIL	Amp	Hui-Yu Liu
1521	pGex4T2-C part of Tid1L	BL21 DE3 RIL	Amp	Hui-Yu Liu
1522	pGex4T2-5Tyrs of C part of Tid1L	BL21 DE3 RIL	Amp	Hui-Yu Liu
1523	pGex4T2-DNA J Domain of Tid1L	BL21 DE3 RIL	Amp	Hui-Yu Liu
1524	pCMV5-His6-Erk2 (L73P/S151D, Constitutively Active)	XL10 Gold	Amp	Natalie Ahn (do not distribute without permission)

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1525	pcDNAhrGFPΔATG	XL10 Gold	Amp	J. MacDonald
1526	pGemT-Easy-Rat Nesca-PC12	XL10 Gold	Amp	J. MacDonald
1527	pGemT-Easy- BMP2-2 (mouse) 200bp	DH5-α	Amp	
1528	pGemT-Easy- TrkC (200bp ECD fragment)	DH5-α	Amp	K. Volkening
1529	pGemT-Easy- FGFR1 (200bp ECD fragment)	DH5-α	Amp	K. Volkening
1530	pGemT-Easy TGFB1 (200bp fragment)	XL10 Gold	Amp	K. Volkening
1531	pGemT-Easy Snail (200bp fragment)	XL10 Gold	Amp	K. Volkening
1532	pGemT-Easy Pax3 (200bp fragment)	XL10 Gold	Amp	K. Volkening
1533	pGemT-Easy BMP4 (200bp fragment)	XL10 Gold	Amp	K. Volkening
1534	pAS2.1-TrkAS13 ($YY^{683/684}$ EE)	XL10 Gold	Amp	Hui-Yu Liu
1535	pAS2.1-TrkAS13e (Y^{684}E)	XL10 Gold	Amp	Hui-Yu Liu
1536	pAS2.1-TrkAS13g (Y^{683}E)	XL10 Gold	Amp	Hui-Yu Liu
1537	pSilencer RNESCA 112	XL10 Gold	Amp	J. M.
1538	pSilencer RNESCA 412	XL10 Gold	Amp	J. M.
1539	pSilencer RNESCA 1027	XL10 Gold	Amp	J. M.
1540	β1- Arrestin	XL10 Gold	Amp	S. Ferguson
1541	β2- Arrestin	XL10 Gold	Amp	S. Ferguson
1542	pRcCMV-nSrc	DH5α	Amp	L. Kalia
1543	pc- SRC-iGFP	DH5α	Kan	L. Kalia
1544				
1545	pCS-CDF-CG-PRE (GFP only), Lentivirus vector	XL10 Gold	Amp/Zeo	Hui-Yu Liu
1546	pMDLg/p.RRE, lentivirus helper 1	XL10 Gold	Amp	Hui-Yu Liu
1547	pRSV-Rev, lentivirus helper 2	XL10 Gold	Amp	Hui-Yu Liu
1548	pMD.G, lentivirus helper 3	XL10 Gold	Amp	Hui-Yu Liu
1549	pGemT-Easy-βgal for riboprobes	XL10 Gold	Amp	K. Volkening
1550	pGemT-Easy TrkA 200bp fragment	XL10 Gold	Amp	K. Volkening
1551	pGemT-Easy TrkB 200bp fragment	XL10 Gold	Amp	K. Volkening
1552	pGemT-Easy FRS3 int frag	XL10 Gold	Amp	K. Volkening
1553	pGemT-Easy-MASH1 for riboprobes	XL10 Gold	Amp	K. Volkening
1554	pCMV-HA-hFRS2-flag	XL10 Gold	Neo/Kan	K. Manto –sequenced – 7th last aa has a silent mutation – otherwise, full length (except stop codon, obviously)
1555	pXJ40-HA-ACK1-K158R (Kinase Dead)	XL10 Gold	Amp	Dr. Ed Manser
1556	pIRES2-EGFP	XL10 Gold	Kan	Invitrogen
1557	pSilencer3-1-H1-hygro	XL10 Gold	Amp	Ambion
1558	pSilencer3-1-H1-hygro-αTid1 new	XL10 Gold	Amp	Hui-Yu Liu
1559	pLentiLox 3.7 (Lenti-SiRNA-EGFP)	XL10 Gold	Amp	Luk Van Parijs (MIT) *MTA signed*
1560	pcDNA 3.1A FRS2 [cc-term) / (a.a.410->)	XL10 Gold	Amp	Angelo
1561	pcDNA3.1A FRS2M123 [T452,455,458-->A]	XL10 Gold	Amp	Angelo *Not sure if correct*
1562	pGemT-Easy FRS2M123 [T452,455,458-->A]	XL10 Gold	Amp	Angelo *Not sure if correct*

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1563	pLenti-hTid1L (no tag)	XL10 Gold	Amp/Zeo	Hui-Yu Liu
1564	pLenti-anti-Tid1-EGFP	XL10 Gold	Amp/Zeo	Hui-Yu Liu
1565	Ap2 FRS2-myc	XL10 Gold	Amp	Kara Manto—sequenced – missing last 5 aa
1566				
1567	pLenti-Tid1S (no tag)	XL10 Gold	Amp/Zeo	Hui-Yu Liu
1568	pcDNA3-HA-RasGrf1-DN (W1056E) for Ras	XL10 Gold	Amp	L. Alberghina JBC 274: 36656 (1999)
1569	pcDNA3-HA-RasGrf1	XL10 Gold	Amp	L. Alberghine
1570	pEGFP-Erk2	XL10 Gold	Kan	P. Stork
1571	pCDNA3-CBR (C3G)	DH5α	Amp	P. Stork
1572	pGEMT-Easy-RasGrf1-PH1	DH5α	Amp	K. Robinson
1573	pGEMT-Easy-RasGrf1-PH1-1Q	DH5α	Amp	K. Robinson
1574	pGST-Ran (G19V); Mimics GTP-bound form	XL10 Gold	Amp	B. Paschal
1575	pFLAG-CRT (Calreticulin)	XL10 Gold	Amp	B. Paschal JCB 152: 127-149 (2001)
1576	pcDNA-CRM1 (WT) Myc-His	XL10 Gold	Amp	C. Dargemont Exp. Cell Res. 252: 236-241 (1999)
1577	pGEMT-Easy-Shh (200bp)	DH5α	Amp	K. V.
1578	pGEMT-Easy-NheI-RasGrf1	DH5α	Amp	K. R.
1579	pRC-TAP	DH5α	Amp	D. Litchfield
1580	pRC-cKIP TAP	DH5α	Amp	D. Litchfield
1581	pRC- TAP cKIP	DH5α	Amp	D. Litchfield
1582	pcDNA-mychisB-RasGrf1-PH1	DH5α	Amp	K. R.
1583	pcDNA-mychisB-RasGrf1-PH1, CC, 1Q	DH5α	Amp	K. R.
1584	pADEasy cells (DH10B)	DH10B	Amp	
1585	pGEM-T EasySNT2Exon2,3 (Clone 3)	XL10 Gold	Amp	Li Zhou
1586	pGEM-T EasySNT2Exon2,3 (Clone 9)	XL10 Gold	Amp	Li Zhou
1587	pGEM-T EasySNT2 5'UTR (Clone 1)	XL10 Gold	Amp	Li Zhou
1588	pGEM-T EasySNT2 5'UTR (Clone 5)	XL10 Gold	Amp	Li Zhou
1589	pGEM-T EasySNT2Exon2,4 (Clone 2)	XL10 Gold	Amp	Li Zhou
1590	pGEM-T EasySNT2Exon2,4 (Clone 3)	XL10 Gold	Amp	Li Zhou
1591	pGEM-T EasySNT2Exon4,5-1	XL10 Gold	Amp	Li Zhou
1592	pGEM-T EasySNT2Exon4,5-2	XL10 Gold	Amp	Li Zhou
1593	pGEM-T EasySNT2Exon5	XL10 Gold	Amp	Li Zhou
1594	pAdEasySNT1 (Clone 6)	BJ5183	Amp	Li Zhou missing last 5 aa
1595	pAdEasySNT1 (Clone 9)	BJ5183	Amp	Li Zhou missing last 5 aa
1596	pAdEasySNT1 (Clone 1)	XL10 Gold	Amp	Li Zhou missing last 5 aa
1597	pAdEasySNT1 (Clone 2)	XL10 Gold	Amp	Li Zhou missing last 5 aa

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1598	pAdEasySNT2	BJ5183	Amp	Li Zhou
1599	pAdEasySNT2	XL10 Gold	Amp	Li Zhou
1600	pcDNA3.1MycHisBSNT1Y436A (Clone 2)	XL10 Gold	Amp	Li Zhou—sequenced—missing last 5 aa
1601	pcDNA3.1MycHisBSNT1Y436A (Clone 3)	XL10 Gold	Amp	Li Zhou
1602	pcDNA3.1MycHisASNT1 4A (Clone 2)	XL10 Gold	Amp	Li Zhou
1603	pcDNA3.1MycHisASNT1 4A (Clone 6)	XL10 Gold	Amp	Li Zhou
1604	pcDNA3.1MycHisBSNT1Y471A	XL10 Gold	Amp	Li Zhou
1606	pcDNA3.1MycHisBSNT1Y436, 471A (Clone 2)	XL10 Gold	Amp	Li Zhou
1607	pcDNA3.1MycHisBSNT1 6A	XL10 Gold	Amp	Li Zhou
1608	pAdTrack-CMV SNT1Y436A	XL10 Gold	Kan	Li Zhou
1609	pAdTrack-CMV SNT1Y471A	XL10 Gold	Kan	Li Zhou
1610	pAdTrack-CMV SNT1Y436, 471A	XL10 Gold	Kan	Li Zhou
1611	pAdTrack-CMV SNT1 4A	XL10 Gold	Kan	Li Zhou
1612	pAdTrack-CMV SNT1 6A	XL10 Gold	Kan	Li Zhou
1613	pAdEasySNT1Y436A	BJ5183	Amp	Li Zhou
1614	pAdEasySNT1Y471A	BJ5183	Amp	Li Zhou
1615	pAdEasySNT1Y436, 472A (Clone 6)	BJ5183	Amp	Li Zhou
1616	pAdEasySNT1Y436, 472A (Clone 7)	BJ5183	Amp	Li Zhou
1617				
1618	pAdEasySNT1 4A (Clone 3)	BJ5183	Amp	Li Zhou
1619	pAdEasySNT1 4A (Clone 4)	BJ5183	Amp	Li Zhou
1620	pAdEasySNT1 6A	BJ5183	Amp	Li Zhou
1621	pAdEasySNT1Y436A	XL10 Gold	Amp	Li Zhou
1622	pAdEasySNT1Y471A	XL10 Gold	Amp	Li Zhou
1623	pAdEasySNT1Y436, 471A	XL10 Gold	Amp	Li Zhou
1624	pAdEasySNT1 4A	XL10 Gold	Amp	Li Zhou
1625	pAdEasySNT1 6A	XL10 Gold	Amp	Li Zhou
1626	pCMXTrkA S11b (no HA tag)	XL10 Gold	Amp	Li Zhou
1627	pcDNA3.1mychisA SNT1T5A (Clone 1) (T376A)	XL10 Gold	Amp	Li Zhou
1628	pcDNA3.1mychisA SNT1T5A (Clone 3)	XL10 Gold	Amp	Li Zhou
1629	pcDNA3.1mychisA SNT1T1,2,3A (Clone 1)	XL10 Gold	Amp	Li Zhou
1630	pcDNA3.1mychisA SNT1T1,2,3A (Clone 2) ($T^{452,455,458}A$)	XL10 Gold	Amp	Li Zhou
1631	pGEM-Teasy-RasGRF1-DH/PH2	DH5a	Amp	K. Robinson
1632	pLenti-HA-TrkA	DH5a	Amp	Hui-Yu Liu
1633	pSCA1, Semliki Forest virus vector	DH5a	Amp	Rod Bremner, (Toronto) *MTA signed*
1634	pSCAhelper, for SFV producing	DH5a	Amp	Rod Bremner, (Toronto) *MTA signed*

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1635	pSHAME2a, SFV vector	DH5a	Amp	Rod Bremner, (Toronto) *MTA signed*
1636	pSCA1-EGFP	DH5a	Amp	Hui-Yu Liu
1637	pSHAME2a-myc-Tid1$_L$	DH5a	Amp	Hui-Yu Liu
1638	pSHAME2a-myc-Tid1$_S$	DH5a	Amp	Hui-Yu Liu
1639	pSHAME2a-HA-TrkA	DH5a	Amp	Hui-Yu Liu
1640	pSHAME2a-HA-TrkAS89	DH5a	Amp	Hui-Yu Liu
1641	pSHAME2a-HA-TrkA13a	DH5a	Amp	Hui-Yu Liu
1642	pRSETB-mRFP1 (good)	DH5a	Amp	Roger Y. Tsien (UCSD)\ *(MTA signed)* PNAS 2002 99:7877-7882
1643	pRK5-myc-Tid1S dam-; dcm- (don't use)	SCS101	Amp	Hui-Yu-Liu
1644	pMT2-SHP2 DM: D^{425}A, C^{459}S: substrate trapping dominant -ve	DH5α	Amp	M. Hayman JBC 278:13952-13958, 2003
1645	pMT2-SHP2 wt	DH5 α	Amp	M. Hayman, Stoney Brook University, NY
1646	pMT2-SHP2 R^{465}E: phosphate minus	DH5 α	Amp	M. Hayman Mol.Cell. Biol. 23: 7875-7886, 2003
1647	pSFVPD	DH5 α	Amp	Alaa El Din El-Husseini
1648	pEBG3-DNAJ (Y^{94}-S^{159})	DH5 α	Amp	Hui-Yu Liu
1649	pEBG3-Tid1N (A^{2}-E^{214})	DH5 α	Amp	Hui-Yu Liu
1650	pEBG3-TidC308 (V^{308}-S^{480})	DH5 α	Amp	Hui-Yu Liu
1651	pEBG3-Tid1L (full length)	DH5 α	Amp	Hui-Yu Liu
1652	pRK5-myc-TidN	DH5 α	Amp	Hui-Yu Liu
1653	pRK5-myc-Tid1L 3' (T215C)	DH5 α	Amp	Hui-Yu Liu
1654	pRK5-myc-TidL 5(Tyr)s (3'end) (R277C)	DH5 α	Amp	Hui-Yu Liu
1655	pRK5-myc-Tid DnaJ (see stock #1410)	DH5 α	Amp	Hui-Yu Liu
1656	pHelper 2 (for pSFV PD)	DH5 α	Amp	Hui-Yu Liu
1657	pRK5-myc-Tid1L 5'	DH5 α	Amp	Hui-Yu Liu
1658	pSFVpD-EGFP	DH5 α	Amp	Hui-Yu Liu
1659	pSFVpD-HA-TrkA	DH5 α	Amp	Hui-Yu Liu
1660	pSCAβ	DH5 α	Amp	Rod Bremner (Toronto)
1661	pSHAME-LacZ	DH5 α	Amp	Rod Bremner (Toronto)
1662	pRK5-myc-Tid J→ C region (Y^{94}~S^{480})	DH5 α	Amp	Hui-Yu Liu
1663	pEBG3-myc-Tid J→C region (Y^{94}~S^{480})	DH5 α	Amp	Hui-Yu Liu
1664	pEFP-RasGrf1-Y130A	XL1Blue	Kan	K.R.
1665	pEFP-RasGrf1-Y66G	XL1Blue	Kan	K.R.
1666	PEFP-RasGrf1-Y95A	XL1Blue	Kan	K.R.
1667	pEFP RasGrf1-Y66A	XL1Blue	Kan	K.R.
1668	pSHAME-EGFP	DH5 α	Amp	Hui-Yu Liu
1669	pRK5-myc-NJ (Tid1A^{2}~S^{169})	DH5 α	Amp	Hui-Yu Liu
1670	pRK5-myc-NV (Tid1A^{2}~V^{304})	DH5 α	Amp	Hui-Yu Liu
1671	pRK5-myc-NE (Tid1A^{2}~E^{429})	DH5 α	Amp	Hui-Yu Liu

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1672	pRK5-myc-NG^{223} (Tid)	DH5 α	Amp	Hui-Yu Liu
1673	pGAD-21Y1-PTB domain (FRS3 trans mice)	DH5 α	Amp	K.V.
1674	pGAD-1X1-PTB domain (FRS3 trans mice)	DH5 α	Amp	K.V.
1675	pCMV-Tid1S H121Q	XL10 Gold	amp	K. Munger (Harvard)
1676	pCMV-Tid1L (H121Q) (DNA J domain mutant)	XL10 Gold	amp	K. Munger (Harvard)
1677	pEFP-RasGrfl-Y95A, Y130A	DH5α	kan	K. Robinson
1678	pEFP-RasGrfl-W1056E	DH5α	kan	K. Robinson
1679	pcDNA3-ShcB-3F	DH5α	amp	Tony Pawson (Toronto)
1680	pCDNAmychisB-RasGrf1 PH-1Q Y130A	DH5α	amp	K. Robinson
1681	pEFP-RasGrf1 S916A	DH5α	kan	K. Robinson
1682	pBridge-v-src	DH5α	amp	T. Hryciw
1683	pSFVPD-Tid1 E GFP fusion	XL10 Gold	amp	Hui-Yu Liu
1684	pSFVPD-Golgi-EYFP	DH5α	amp	Hui-Yu Liu
1685	pSFVPD-mitochondrion-EYFP	DH5α	amp	Hui-Yu Liu
1686	pTag-FRS3-FLAG	DH5α	kan	KV
1687	pBridge-v-src-ShcBSH2	DH5α	amp	T. Hryciw
1688	pBridge-v-src-FRS3-nomyr	DH5α	amp	KV
1689	pBridge-v-src-FRS3-COOH	DH5α	amp	KV
1690	pAS2.1-FGFR1-myc-his	DH5α	amp	KV
1691	HA-H-Ras	?	amp	James Keller
1692	pGEX4T2-hTid1L (full length)	BL21 RIL	amp	Hui-Yu Liu
1693	pGEX4T2-hTid1L (full length)	BL21 RP	amp	Hui-Yu Liu
1694	pSuper-FRS3-siRNA-#1	DH5α	amp	Shaochon Yan
1695	pSuper-FRS3-siRNA-#2	DH5α	amp	Shaochon Yan
1696	pSuper-FRS3-siRNA-#3	DH5α	amp	Shaochon Yan
1697	pSuper-FRS3-siRNA-control	DH5α	amp	Shaochon Yan
1698	pRC656 Bc12	DH5α	amp	Sean Creegan
1699	pcDNA3.1mychis human FRS3 (5A)	DH5α	amp (stop codon prior to mychis)	Shaochon Yan
1700	pIRES	DH5α	amp	Clontech
1701	pBridge	DH5α	amp	Clontech
1702	pTag-FRS2-Flag (clone 2)	DH5α	amp	KV missing last 5 aa
1703	pcDNA3.1 mychis mouse FRS3 (with 3'UTR)	DH5α	amp	Shaochon Yan
1704	pcDNA3.1 mychisB FRS2 ^{Y436,471}A	DH5α	amp	RML (same as 1605, diff. cell line) sequenced—missing last 5 aa (SL)
1705	pGBKT7 Nesca SH3 only	DH5α	kan	RML
1706	pcDNA3.1 mychis FRS2T^{452,455,458,473}A	DH5α	amp	RML (PxTP mutant)
1707	pCDX HA-NuP214	DH5α	amp	RML
1708	pSuper-FRS3-SiRNA- #4	DH5α	amp	Shaochon Yan

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1709	pSuper-FRS3-SiRNA- #5	DH5α	amp	Shaochon Yan
1710	pEGFP-NS-Nesca-W261A	DH5α DH5α	kan	Matt C.
1711	pRK5-mycShc B (mouse, CH2-)	DH5α	amp	RML(same as 1268, diff. cell line)
1712	pcDNAmychisB-RasGrfPHIQ Y66A, Y130A	DH5α	amp	K. Robinson
1713	pGBKT7myc FRS2T1234A (^{452,455,458,473})	DH5α	kan	RML
1714	pEGFPN3 Nesca W284A	DH5α	kan	Matt
1715	pEGFPN3 Nesca LZ mutant	DH5α	kan	J.M.
1716	pcDNAmychisBRasGrf PHIQ Y66A, Y95A, Y130A	DH5α	amp	K. Robinson
1717	pcDNA3-ShcB-3F-Y316A	DH5α	amp	RKB
1718	pcDNAmychisB Y66,95,130,233A PHIQ	DH5α	Amp	K. Robinson
1719	pcDNAmychisB Y66,95,130,146,233A PH1IQ	DH5α	Amp	K. Robinson
1720	pcDNAmychisC-FRS3-5A no stop	DH5α	Amp	Kim Bertens
1721	pAS2.1FRS2T1234A($T^{452,455,458,473}A$)	DH5α	Amp	RML
1722	pSuper Ras Grf si909	DH5α	Amp	KR
1723	pSuper Ras Grf si2090	DH5α	Amp	KR
1724				
1725				
1726				
1727				
1728	pGAD424-Grb2SH2 domain	DH5α	Amp	KV
1729	pcDNA3.1mychis ShcBΔPTB-Flag	DH5α	Amp	Todd
1730	pcDNA3.1mychis FRS2 N-T12A ($T^{145,148}A$)	DH5α	Amp	RML
1731				
1732				
1733	pcDNA3.1mychis FRS2 N-T1234A ($T^{145,148,150,151}A$)	DH5α	Amp	RML
1734	pSCA-HA-TrkA wt	DH5α	Amp	Hui-Yu Lui
1735	pADTrack-si #4	DH5α	Kan	KV
1736	pAS2.1-FGFR2	DH5α	Amp	Kim Bertens
1737	pAS2.1-FGFR3	DH5α	Amp	Kim Bertens
1738	pcDNA3-ShcB-3A-3Flag ($Y^{316,317,395}A$)	DH5α	Amp	Kim Bertens
1739	pEBG CKS1	XL10	Amp	JM
1740	pEBG CKS2	XL10	Amp	JM
1741	pEBG $p13^{sucl}$	XL10	Amp	JM
1742	pBridge-Leu (replaced Trp→ Leu)	DH5α	Amp	KV
1743	pCMV-Tag4c-c-cbl	DH5α	Kan	RML

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1744	pAdEasy si hFRS3 #1-5	DH5α	Kan	KV
1745	pAdEasy si hFRS3 #2-1	DH5α	Kan	KV
1746	pAdEasy si hFRS3 #3-9	DH5α	Kan	KV
1747	p CW7 myc-ubiquitin	DH5α	Amp	RML
1748	pAd Track-CMV-hFRS3 $Y^{192}A$, $Y^{322}A$ (Grb 2)	DH5α	Kan	JG
1749	pAd Track-CMV-FRS3-5A	DH5α	Kan	JG
1750	pBridge(leucine)-FRS3-COOH-vsrc	DH5α	amp	KV
1751	pBridge(leucine)-FRS3-NM-vsrc	DH5α	amp	KV
1752	pEGFP N3 RasGrf1	DH5α	kan	JG
1753	pEGB ShcB CH1	XL10 gold	amp	J.M.
1754	pcDNAmychis-Grf2	DH5α	amp	JG
1755	pcDNA3.1 Nesca-ΔSH3	XL10 gold	amp	JM
1756	pRFPmut		kan	JM
1757	pShuttle	DH10B	kan	Bert Vogelstein
1758	pADTrack CMV-TrkA S8	DH5α	kan	Todd H.
1759	pShuttle CMV	DH5α	kan	KV
1760	GST-PBD (PAK binding domain)	DH5α	amp	David Scott/ Martin Schwartz
1761				
1762				
1763	pGEX4T3-hFynTSH2	DH5α	amp	Shawn Li
1764	pRc/CMV-FynTmyc	DH5α	amp	Shawn Li
1765	pGEM-FRS3-Flag (Mlu primer sites) clone 1	DH5α	amp	KV
1766	pGEM-FRS3-Flag (Mlu primer sites) clone 2	DH5α	amp	KV
1767	pAdTrack CMV-FRS3 1A mutant $Y^{322}A$	DH5α	kan	KV
1768	pcDNA3.1 PSD-95 myc	DH5α	amp	Kevin Holmes
1769	pcDNA3.1 STEP61wt	DH5α	amp	P.J. Lombroso
1770	pcDNA3.1 STEP61 C4725	DH5α	amp	P.J. Lombroso
1771	pcDNA3.1 STEP46 wt	DH5α	amp	P.J. Lombroso
1772	pcDNA3.1 STEP46 C300S	DH5α	amp	P.J. Lombroso
1773	GFP-NR1-1a	DH5α	amp	Stefano Vincni
1774	GFP-NR2A	XL10 gold	amp	Stefano Vincni
1775	GFP-NR2B	XL10 gold	amp	Stefano Vincni
1776	pBridge-ShcB SH2	DH5α	amp	Todd Hryciw
1777	pShuttle-CMV-Nesca-EGFP	DH5α	kan	JG
1778	pcDNAmychis Nesca L174A, L195A	XL10 gold	amp	JM
1779	pNTAPCmychis Nesca L174A, l195A	XL10 gold	kan	JM
~~1780~~	~~pSuperFRS2 si #2~~	~~DH5α~~	~~amp~~	~~KV~~—*Do Not Use*

	NAME	HOST Str	ANTIBIOTIC	SOURCE
~~1781~~	~~pSuperFRS2 si#1~~	~~DH5α~~	~~amp~~	~~KV~~ *Do Not Use*
~~1782~~	~~pSuper RasGrf 909siRNA~~	~~DH5α~~	~~amp~~	~~KV~~ *Do Not Use*
~~1783~~	~~pSuper RasGrf 2009siRNA~~	~~DH5α~~	~~amp~~	~~KV~~ *Do Not Use*
1784	pcDNA3.1ShcB-flag-myc-his	DH5α	amp	Todd Hryciw
1785	pIRES-EGFP-DBH-hFRS3	DH5α	amp	KV
1786	pcDNA-Flag-mGluR5	DH5α	amp	S Ferguson
1787	pAdTrack-CMV-FRS3-Y192A,Y287A,Y322A	DH5α	Kan	KV
1788	pAdTrack-CMV-FRS3-Y192A,Y287A, Y322A, Y455A	DH5α	Kan	KV
1789	pCMX-FRS3 (no tag)	DH5α	Amp	KV
1790	pAdTrack-FRS3 Y417A, Y455A	DH5α	Kan	KV
1791	pAdTrack-NESCA	DH5α	KAN	JG
~~1792~~	~~pSUPER-RasGRF1 shRNA311~~	~~DH5α~~	~~Amp~~	~~KV~~ *Do Not Use*
~~1793~~	~~pSUPER-RasGRF1 shRNA1082~~	~~DH5α~~	~~Amp~~	~~KV~~ *Do Not Use*
~~1794~~	~~RASGRF1-EGFP (clone 1)~~	~~DH5α~~	~~Kan~~	~~KV~~ *Do Not Use*
1795	RasGRF1-EGFP (clone 9) in pEFP	DH5α	Kan	KV
1796	pcDNA beta-catenin (Xenopus)	DH5α	Amp	T Drysdale
1797	pCS2-MT-beta-catenin (Xenopus)	DH5α	Amp	T Drysdale
1798	pCMX-FRS2 (no tag)	DH5α	Amp	KV missing last 5 aa
~~1799~~	~~pSUPER-RasGrf1/2 shRNA627~~	~~DH5α~~	~~Amp~~	~~KV~~ *Do Not Use*
1800	pcDNA-myc-hisB PH1-Y95A	DH5α	Amp	KR
1801	pGEMTEasy-EGFP-BamH1	DH5α	Amp	KR
1802	pGEMTEasy-EGFP-Xho1	DH5α	Amp	KR
1803	beta-arrestin1-GFP	DH5α	Kan	S. Ferguson
1804	beta-arrestin2-GFP	DH5α	Kan	S. Ferguson
1805	beta-arrestin1-Flag	DH5α	Amp	S. Ferguson
1806	beta-arrestin2-Flag	DH5α	Amp	S. Ferguson
1807	pcDNA-HA Rac V12	DH5α	Amp	S. Ferguson
1808	pcDNA-HA Rac N17	DH5α	Amp	S. Ferguson
1809	pcDNA-HA Rac WT	DH5α	Amp	S. Ferguson
1810	pcDNA-myc-hisB-PH1-Y95A,Y130A	DH5α	Amp	KR
1811	pCS2-mapk (xenopus)	DH5α	Amp	(KV) T Drysdale
1812	pCS2MT	DH5α	Amp	(KB) T Drysdale
1813	pcmx-TrkA K485A	DH5α	Amp	KV
1814	HA Ubiquitin	DH5α	Amp	KV Wooten lab? (ask Rosslynn for details & maps
1815	HA Ubiquitin K29R	DH5α	Amp	" " "
1816	HA Ubiquitin K48R	DH5α	Amp	" " "
1817	HA Ubiquitin K63R	DH5α	Amp	" " "
1818	HA Ubiquitin	DH5α	Amp	Geetha et al 2005
1819	HA Ubiquitin K29R	DH5α	Amp	Geetha et al 2005
1820	HA Ubiquitin K48R	DH5α	Amp	Geetha et al 2005
1821	HA Ubiquitin K63R	DH5α	Amp	Geetha et al 2005

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1822	pCMX-TrkB K464R a	DH5α	Amp	KV
1823	pCMX-TrkB K464R b (diff. clone)	DH5α	Amp	KV
1824	pCMX-TrkC K465R a	DH5α	Amp	KV
1825	pCMX-TrkC K465R b (diff. clone)	DH5α	Amp	KV
1826	pCMX-HA-TRKB-S10 (HIKE)	DH5α	Amp	KR
1827	pcDNA-myc-hisB-Ras Grf1	DH5α	Amp	KR
1828				
1829				
1830				
1831				
1832	pBABE mycSin	DH5α	Amp	Jan Sap
1833	pEX Lox Sin	DH5α	Amp	Jan Sap
1834	pBABE IEG SinSD	DH5α	Amp	Jan Sap
1835	pRK5 SinSD	DH5α	Amp	Jan Sap
1836	pBABE IEG	DH5α	Amp	Jan Sap
1837	pBABE IEG Sin	DH5α	Amp	Jan Sap
1838	pRK5 Sin	DH5α	Amp	Jan Sap
1839	Sin DSR	DH5α	Amp	Jan Sap
1840	pCMX-HA-TrkB S21	DH5α	amp	Andrew Lu
1841	pcDNA3 flag-axin	xl10gold	amp	F Costantini Columbia U. Do not distribute
1842	pcs2mt myc-axin	xl10gold	amp	F Costantini Columbia U. Do not distribute
1843	pEGFP(N1)ShcB	DH5α	Kan	Todd Hryciw
1844	pcdna-rasgrf1, y130a, y233a	DH5α	kan	kr
1845	pcdna-rasgrf1, y95a, y130a, y233a	DH5α	kan	kr
1846	pCALNL5	DH5α	amp	Riken BRC (Japan)
1847	pENTR2B-βCatenin	transformation did not work yet - tba	amp	David Mulholland
1848	pDEST53-GFP	XL10gold	amp	David Mulholland
1849	pEGFP-βCatenin wild type	DH5α	kan	Chin-Yin Tai, Caltech
1850	pEGFP-βCatenin Y654F	DH5α	kan	Chin-Yin Tai, Caltech
1851	pEGFP-βCatenin Y654E	DH5α	kan	Chin-Yin Tai, Caltech
1852	pcDNA3.1myc-HISc-p55ShcC	DH5α	amp	Todd Hryciw
1853	pcDNA3.1myc-HISc-p55ShcCΔSH2	DH5α	amp	Todd Hryciw
1854	pEGFP-N1-p55ShcC	DH5α	kan	Todd Hryciw
1855	mRFP-Rab5	DH5α	kan	Ferguson (Pieter)
1856	mRFP-Rab7	DH5α	kan	Ferguson (Pieter)
1857	pCALNL5-Frs2-EGFP	DH5α	Amp	S. LeMay missing last 5aa
1858	pCMX-hFrs3 myr(-)	DH5α	amp	Todd Hryciw
1859	pSuperRasGrf1-si909	DH5α	amp	KR
1860	pSuperRasGrf1-si2009	DH5α	amp	KR
1861	pEGFPN1-Frs2	DH5α	kan	Sara LeMay

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1863	pcDNA3.1mycHIS-ShcC CH1	DH5α	amp	Todd Hryciw
1864	pcDNA3.1mycHIS-Frs3(myr$^-$)	DH5α	amp	Todd Hryciw
1865	pShuttle hrGFP-1	xl10gold	kan	stratagene
1866	pgex4t1 shcC-sh2	DH5α	amp	ian
1867	pcs2mt-frs2	xl10gold	amp	james
1868	pAd Track	dh10b	kan	Bert Vogelstein
1869	pAd Track-CMV	DH10B	kan	Bert Vogelstein
1870	pEGFP-LC-3	DH5α	kan	Yoshimori(NIPPON)
1871	pEGFP-dn-Rab5	DH5α	kan	Ferguson(pieter)
1872	pEGFP-dn-Rab7	DH5α	kan	Ferguson(pieter)
1873	pRS	DH5α	amp	OriGene
1874	pRS-shGFP(29)non-effective	DH5α	amp	OriGene
1875	pRS-sh Beclin1 -29	DH5α	amp	OriGene
1876	pRS-sh Beclin1 -30	DH5α	amp	OriGene
1877	pRS-sh Beclin1 -31	DH5α	amp	OriGene
1878	pRS-sh Beclin1 -32	DH5α	amp	OriGene
1879	pEGFP-N1-Frs3	DH5α	kan	sara lemay
1880	pCAL-Frs2-EGFP (full length Frs2)	DH5α	amp	sara lemay***needs to be re-cloned
1881	pRS-sh Frs2 -69	DH5α	amp	OriGene
1882	pRS-sh Frs2 –70	DH5α	amp	OriGene
1883	pRS-sh Frs2 –71	DH5α	amp	OriGene
1884	pRS-sh Frs2 -72	DH5α	amp	OriGene
1885	pCDNA-TrkC	DH5α	amp	Jen F
1886	pCAL-Frs3-EGFP	DH5α	amp	sara lemay
1887	pAD Cre	DH5α	Kan\	
1888	pBridge-v-src-shcC SH2	DH5α	amp	Ian G
1889	NR2B-CFP	DH5α	amp	
1890	PSD95-YFP	DH5α	kan	
1891	pGex ShcC	DH5α	AMP	Ian G
1892	pBridge ShcC SH2	DH5α	Amp	Ian G
1893	pGex2t Step 46	DH5α	Amp	Ian G
1894	pGex2T Step 46 CS	DH5α	Amp	Ian G
1895	P Gex2T Step 61	DH5α	Amp	Ian G
1896	pcDNA 3.1 HA PKC zeta	DH5α	Amp	Wooten/Moocat
1897	pcDNA 3.1 HA PKC zeta CA	DH5α	Amp	Wooten/Moocat
1898	pcDNA 3.1 HA Myc-PKC zeta	DH5α	Amp	Wooten/Moocat
1899	pcDNA 3.0-MKK7-JNK1	DH5α	Amp	Davis R
1900	pcDNA 3.0-MKK7JNK1 (apf)	DH5α	Amp	Davis R
1901	pcDNA 3.0-Flag-Rap1	DH5α	Amp	Stoke/Oregon
1902	pcDNA 3.0-Myc-Rap1	DH5α	Amp	Stoke/Oregon
1903	pcDNA 3.1 DN-PKC zeta	XL10 Gold	Amp	Jupinder
1904	pcDNA 4-myc HisA	DH5α	Amp	Zhong/U Montreal
1905	pcDNA 4 Foxo3-DN + Myc	DH5α	Amp	Zhong/U Montreal
1906	pcDNA 4 Foxo 3-CA + Myc	DH5α	Amp	Zhong/U Montreal

	NAME	HOST Str	ANTIBIOTIC	SOURCE
1915	pBIG2i	DH5α	Amp	Craig A.Strathdee/ Amgen
1916	pBIG2r	DH5α	Amp	Craig A.Strathdee/ Amgen
1917	pBIG3i	DH5α	Amp	Craig A.Strathdee/ Amgen
1918	pBIG3r	DH5α	Amp	Craig A.Strathdee/ Amgen

2007-06

BIO-RRI-0011

robarts RESEARCH

BIOHAZARDOUS AGENTS REGISTRY FORM

Reviewed by Biosafety Subcommittee: February 2006

This form must be completed by each Principal Investigator when completing a grant application or grant renewal to be administered by the Robarts Research Institute, if the use of biohazardous and/or infectious agents is proposed. For any proposed animal work involving the use of biohazardous agents or animals carrying zoonotic agents infectious to humans, this form must also be completed.

COMPLETED FORMS ARE TO BE RETURNED TO BIOSAFETY SUBCOMMITTEE CHAIR, ROOM 3-34.1.

If there are any changes to the information on these forms (excluding grant title and funding agencies) a new form must be completed and sent to the Biosafety Subcommittee Chair BEFORE implementation of these changes can occur.

If multi-team grants are being applied for, each individual Investigator of the team must submit a Biohazardous Agents Registry Form to the Biosafety Subcommittee Chair.

Containment Levels will be required in accordance with Health Canada (HC), Laboratory Biosafety Guidelines, 3rd edition 2004, or Canadian Food Inspection Agency (CFIA), Containment Standards for Veterinary Facilities, 1st edition 1996.

For questions regarding this form, please contact Biosafety Subcommittee Chair at ext. 34125.

1.0 Contact Information

PRINCIPAL INVESTIGATOR: Dr. Susan Meakin

SIGNATURE: Susan Meakin

DATE: May 3rd, 2007

DEPARTMENT: Cell Biology and Stem Cell Biology, Dept of Biochemistry

ADDRESS: Robarts Research Institute, Office 3-16.1

TELEPHONE: 663-5777 ext. 34304

EMAIL: smeakin@robarts.ca

Location of experimental work to be carried out:

Building(s): Robarts Research Institute

Room(s): 3-20.1, 3-17.1 (Tissue Culture Room)

**For work being performed at Institutions affiliated with the Robarts Research Institute, the Safety Officer for the Institution where experiments will take place must sign the form prior to it being sent to Robarts Research Institute, Biosafety Subcommittee Chair. See Section 13.0, Approvals*

GRANT TITLE(S): 1. TrkA Activation of Autophagy in Human Neural Tumors
2. The Role of the FRS3 Adapter in Regulating b-catenin Signaling and Cellular Proliferation
3. The Roles of the FRS adapters in Stem Cell Survival and Proliferation
4. TrkB, ShcC and RasGrf1 Regulate NMDA Receptor Activity
5. Nesca, a Novel Signaling Adapter that Regulates Neuronal Growth and Function
6. Autophagy Induced Cell Death in Human Brain Tumors

ATTACH A BRIEF DESCRIPTION OF YOUR WORK, SUCH AS THE RESEARCH GRANT SUMMARY(S) EXPLAINING THE BIOHAZARD(S) USED.

FUNDING AGENCY/AGENCIES:_Cancer Research Society: Grants #1 and #2_____

_Krembil/ORDCF: Grant #3, NSERC: Grant #4, CIHR: Grant #5 (Applied), McDonnell Foundation: Grant #6 (Applied)

Anticipated Grant End Date: _Aug 31, 2008 (both CRS grants), _Sept, 2012 (CIHR),___March 2012 (NSERC)_____

Names of all personnel working under Principal Investigator's supervision in this location:

Dr. James MacDonald(Res. Assoc.)_____

Mrs. Sara LeMay(Res. Tech)_____

Dr. Todd Hryciw(PDF)_____

_Dr. Sandy Vascotto__(PDF)_____

Mrs. Kim Brookes(Ph.D. Student)_____

Mr. Chunhui Li(Res. Tech)_____

Ms. Jennifer Gerasimoff(M.Sc. Student)_____

___Ms. Jennifer Forsyth (M.Sc. Student)_____

___Mr. Andrew Lu (Summer Student)_____

___Mr. Jupinder Bains (Summer Student)_____

_____

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Note : A list of human pathogens categorized according to Risk Group can be obtained by calling the Office of Laboratory Security directly at (613) 957-1779 or accessing their Web site : http://www.phac-aspc.gc.ca/ols-bsl/index.html

2.0 Microorganisms

2.1 Does your work involve the use of microorganisms? **YES** X **NO** O
If **NO**, please proceed to Section 3.0

2.2 Please complete the table below:

Name of Microorganism	Is microorganism a known human pathogen? YES/NO	Is microorganism a known animal pathogen? YES/NO	Is microorganism a known zoonotic agent? YES/NO	Maximum quantity to be cultured at one time?	Health Canada or CFIA Containment Level (select one)
Yeast	No	No	No	500 ml	1O 2 X 3O
Bacteria (E.coli) containing plasmids	No	No	No	1000 ml	1O 2 X 3O
					1O 2O 3O

3.0 Cell Culture

3.1 Does your work involve the use of cell cultures? YES X NO O
If **NO**, please proceed to Section **4.0**.

3.2 Please indicate in the table below the type of cells that will be grown in culture.

Cell Type	Is this cell type used in your work? **YES / NO**	Established or Primary *	Supplier of Primary Cell Culture Tissue
Human	Yes	Established (see below)	
Rodent	Yes	Both (See Below)	E17, P1, P3, P7, P30 Mice
Non-human primate	Yes	Cos 7 cells (established)	
Other (specify)	Sf21 Insect Cells, HEK 293T, cortical, neurospheres, hippocampal, motoneurons, nnr5, Medulloblastoma, gliomas, neuroblastomas, nnr5, PC12,		

* i.e. derived from fresh tissue

Complete the following table.

Specific Cell Line	Source / Supplier	HC or CFIA Containment Level (select one)
nnr5, PC12	In house	1 O 2 X 3 O
Medulloblastoma, Gliomas	In house, ATCC	1 O 2 X 3 O
~~HEK, Cos 7~~	~~In house, ATCC~~	1 O 2 X 3 O

4.0 Use of Human Source Materials

4.1 Does your work involve the use of human source materials? YES O NO X
If **NO**, please proceed to Section **5.0**

4.2 Indicate in the table below the Human Source Material to be used.

Human Source Material	Specify Source, or Not Applicable (NA)	Is Human Source Material known to be infected with an infectious agent? YES/NO	Name of Infectious Agent	HC or CFIA Containment Level (select one)
Human Blood (whole) or other Body Fluid				1O 2O 3O
Human Blood (fraction) or other Body Fluid				1O 2O 3O
Human Organs (unpreserved)				1O 2O 3O
Human Tissues (unpreserved)				1O 2O 3O

5.0 Genetically Modified Organisms and Cell lines

5.1 Will genetic modifications be made to the organism, virus or cell line? YES X NO O
If NO, please proceed to Section 6.0

5.2 Will genetic sequences from any of the following be involved?

- HIV YES O NO X

If YES, specify: _____

- HTLV 1 or 2 YES O NO X

If YES, specify: _____

- Other human or animal pathogen and/or their toxins YES X NO O

If YES, specify: Lentiviral vectors_____

5.2 Will intact genetic sequences be used from:

- SV 40 Large T antigen YES X NO O
- Adeno E1A YES O NO X
- Known or suspected oncogenes YES O NO O X

If YES, specify: _In all mammalian expression vectors,_____

5.4 Will a live vector(s) (viral or bacterial) be used for gene transduction? YES X NO O

If YES, name vector: _pCMX, pEGFP, pGex, pTapTag, pCAL, pAdEasy, _____

5.5 List specific vector(s) to be used:_Adenoviral, Lentiviral (Invitrogen), pBPX (insect)_____

5.6 Will vector be replication defective? YES X NO O

5.7 Will vector be infectious to humans or animals? YES X NO O

5.8 Will this be expected to increase the Containment Level required? YES X NO O

6.0 Human Gene Therapy Trials

6.1 Will human clinical trials using the vector(s) in 5.5 be conducted? YES O NO X
If NO, please proceed to Section 7.0
If YES, attach a full description of the make-up of the virus.

6.2 Will vector be able to replicate in the host? YES O NO O

6.3 How will the vector be administered? _____

6.4 Please give the Health Care Facility where the clinical trial will be conducted:

_____

6.5 Has human ethics approval been obtained? YES O NO O

Approval # _____

7.0 Animal Experiments

7.1 Will any of the agents listed be used in live animals? YES O NO X
If **NO**, please proceed to section 8.0

7.2 Name of animal species to be used: _____

7.3 AUS protocol # _____

7.4 If using murine cell lines, have they been tested for murine pathogens? YES O NO O

8.0 Use of Animal species with Zoonotic Hazards

8.1 Will any of the following animals or their organs, tissues, lavages or other bodily fluids including blood be used?

- Pound source dogs YES O NO X
- Pound source cats YES O NO X
- Sheep or goats YES O NO X
- Non- Human Primates YES O NO X

If **YES** specify species _____

- Wild caught animals YES O NO X

If **YES** specify species _____

9.0 Biological Toxins

9.1 Will toxins of biological origin be used? YES O NO X
If **NO**, please proceed to Section **10.0**
If **YES**, please name the toxin _____

9.2 What is the LD_{50} (specify species) of the toxin? _____

10.0 Import Requirements

10.1 Will the agent be imported? YES X NO
If **NO**, please proceed to Section **11.0**
If **YES**, country of origin _____

10.2 Has an Import Permit been obtained from HC for human pathogens? YES X NO O

10.3 Has an import permit been obtained from CFIA for animal pathogens? YES X NO O

10.4 Has the import permit been sent to Biosafety Subcommittee Chair? YES X NO O

If **YES**, Permit # __P-13851 for transformed 293 cells _____

11.0 Training Requirements for Personnel Named on Form

All personnel named in section 1.0 of this form who will be using any of the above named agents are required to attend the following training courses given by OH&S.

- Biosafety
- Laboratory and Environmental/Waste Management Safety
- WHMIS

As the Principal Investigator, I have ensured that all of the personnel named on the form who will be using any of the biohazardous agents in Sections 2.0 to 10.0 have been trained as required.

SIGNATURE Susan Meakin

12.0 Containment Levels

12.1 For the work described in sections **2.0** to **10.0**, select the highest HC or CFIA Containment Level required. 1O 2 X 3O

With Lentiviral work operationally handled with level 3 practices

12.2 Has the facility been certified by Biosafety Subcommittee Chair for this level of containment?

YES X NO O

If **YES**, give date: ~~August 2003~~ February 28, 2007 and permit number: 2007-02(3-17).

13.0 Approvals

Robarts Research Institute

Signature _____ Date May 03, 2007

Biosafety Officer for the Institution where experiments will take place

Signature _____ Date _____

Biosafety Officer of Robarts Research Institute (if different than above)

Signature _____ Date _____

Note: This permit will be in effect from May 04, 2007 to May 2010, subject to annual facility re-certification.