[redacted]

Approved Personnel
(Please stroke out any personnel to be removed)

Additional Personnel
(Please list additional personnel here)

	Please stroke out any approved Biohazards to be removed below	Write additional Biohazards for approval below. *
Approved Microorganisms		ATCC#
Approved Cells	Human (established) LoVo, C2C12 myoblast cell ~~PC-3~~	PC-3 Human prostate tumor line CRL-5803 NCI-H1299 Human lung carcinoma CRL-1435
Approved Use of Human Source Material		
Approved GMO	myc+, myb+, ras+, fos+, p53, sis-, abl-, ros-, src-	
Approved use of Animals	Nude rats / ~~Nude mice~~	Nude mice
Approved Toxin(s)		

* *PLEASE ATTACH A MATERIAL SAFETY DATA SHEET OR EQUIVALENT FOR NEW BIOHAZARDS.*
** *PLEASE ATTACH A BRIEF DESCRIPTION OF THE WORK THAT EXPLAINS THE BIOHAZARDS USED AND HOW THEY WILL BE USED.*

Classification: 1

Date of last Biohazardous Agents Registry Form: Dec 7, 2007

Signature of Permit Holder: Ting-Yim Lee

BioSafety Officer(s):

Chair,Biohazards Subcommittee:

Thursday, January 29, 2009

Product Description

Before submitting an order you will be asked to read and accept the terms and conditions of ATCC's Material Transfer Agreement or, certain cases, an MTA specified by the depositing institution.

Customers in Europe, Australia, Canada, China, Hong Kong, India, Japan, Korea, Macau, Mexico, New Zealand, Singapore, and Taiwa R.O.C. must contact a local distributor for pricing information and to place an order for ATCC cultures and products.

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Cell Biology

ATCC® Number:	**CRL-5803™**	**Price:**	**$264.00**
Designations:	NCI-H1299	**Depositors:**	AF Gazdar, JD Minna
Biosafety Level:	1	**Shipped:**	frozen
Medium & Serum:	See Propagation	**Growth Properties:**	adherent
Organism:	*Homo sapiens* (human)	**Morphology:**	epithelial

Source: **Organ:** lung
Disease: carcinoma; non-small cell lung cancer
Derived from metastatic site: lymph node

Cellular Products: neuromedin B

Permits/Forms: In addition to the MTA mentioned above, other ATCC and/or regulatory permits may be required for the tra of this ATCC material. Anyone purchasing ATCC material is ultimately responsible for obtaining the per Please click here for information regarding the specific requirements for shipment to your location.

Related Cell Culture Prod

Restrictions: The line is available with the following restrictions: 1. This cell line was deposited at the ATCC by Dr. A. Ga and Dr. J. Minna and is provided for research purposes only. Neither the cell line nor products derived fr may be sold or used for commercial purposes. Nor can the cells be distributed to third parties for purpos sale, or producing for sale, cells or their products. The cells are provided as service to the research commu They are provided without warranty of merchantability or fitness for a particular purpose or any other warr expressed or implied. 2. Any proposed commercial use of the these cells, or their products must firs negotiated with the University of Texas Southwestern Medical Center at Dallas, 5323 Harry Hines Blvd., D. Texas 75235. Telephone (214) 699-8056, FAX (214) 688-7233.

Applications: transfection host (technology from amaxa
Roche FuGENE® Transfection Reagents)

Age: 43 years adult

Gender: male

Ethnicity: Caucasian

Comments: The cells have a homozygous partial deletion of the p53 protein, and lack expression of p53 protein. reported to be able to synthesize the peptide neuromedin B (NMB) at 0.1 pmol/mg protein, but not the ga releasing peptide (GRP).

Propagation: **ATCC complete growth medium:** The base medium for this cell line is ATCC-formulated RPMI-1640 Mec Catalog No. 30-2001. To make the complete growth medium, add the following components to the medium: fetal bovine serum to a final concentration of 10%.
Temperature: 37.0°C

Subculturing: **Protocol:**

1. Remove and discard culture medium.
2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin - 0.53 mM EDTA solution to remove all trac serum that contains trypsin inhibitor.
3. Add 2.0 to 3.0 ml of Trypsin-EDTA solution to flask and observe cells under an inverted microscope cell layer is dispersed (usually within 5 to 15 minutes).
   Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the to detach. Cells that are difficult to detach may be placed at 37C to facilitate dispersal.
4. Add 6.0 to 8.0 ml of complete growth medium and aspirate cells by gently pipetting.
5. Add appropriate aliquots of the cell suspension to new culture vessels.
6. Incubate cultures at 37C.

Subcultivation Ratio: *A subcultivation ratio of 1:3 to 1:6 is recommended*
Medium Renewal: *Every 2 to 3 days*

Preservation: **Freeze medium:** Complete growth medium supplemented with 5% (v/v) DMSO
Storage temperature: liquid nitrogen vapor phase

Related Products: recommended serum:ATCC 30-2020
purified DNA:ATCC CRL-5803D
Recommended medium (without the additional supplements or serum described under ATCC Medium):ATC 2001

References:
23517: Glaccone G, et al. Neuromedin B is present in lung cancer cell lines. Cancer Res. 52: 2732s-2 1992. PubMed: 1563005
23570: . NCI-Navy Medical Oncology Branch Cell Line Supplement. J. Cell. Biochem. suppl. 24: 1996..
33177: Lin DL, Chang C. p53 is a mediator for radiation-repressed human TR2 orphan receptor express MCF-7 cells, a new pathway from tumor suppressor to member of the steroid receptor superfamily. J. Chem. 271: 14649-14652, 1996. PubMed: 8663350

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These cells will be mixed into matrigel & injected into the left lung. The injections will be done on anesthetized (nude) rats in a BSC in F5-104. all involved will be wearing lab coats, gloves and safety glasses.

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Product Description

Before submitting an order you will be asked to read and accept the terms and conditions of ATCC's Material Transfer Agreement or, certain cases, an MTA specified by the depositing institution.

Customers in Europe, Australia, Canada, China, Hong Kong, India, Japan, Korea, Macau, Mexico, New Zealand, Singapore, and Taiwa R.O.C. must contact a local distributor for pricing information and to place an order for ATCC cultures and products.

See use on page 3

Print this

Cell Biology

ATCC® Number:	**CRL-1435™** [redacted]	**Price:**	**$256.00**
Designations:	PC-3	**Depositors:**	ME Kaighn
Biosafety Level:	1	**Shipped:**	frozen
Medium & Serum:	See Propagation	**Growth Properties:**	adherent (The cells form clusters in soft agar and be adapted to suspensior growth)
Organism:	*Homo sapiens* (human)	**Morphology:**	epithelial
Source:	**Organ:** prostate **Tumor Stage:** grade IV **Disease:** adenocarcinoma **Derived from metastatic site:** bone		
Permits/Forms:	In addition to the MTA mentioned above, other ATCC and/or regulatory permits may be required for the tra of this ATCC material. Anyone purchasing ATCC material is ultimately responsible for obtaining the per Please click here for information regarding the specific requirements for shipment to your location.		
			Related Cell Culture Prod
Applications:	transfection host (technology from amaxa Roche FuGENE® Transfection Reagents)		
Tumorigenic:	YES		
Antigen Expression:	HLA A1, A9		
DNA Profile (STR):	Amelogenin: X CSF1PO: 11 D13S317: 11 D16S539: 11 D5S818: 13 D7S820: 8,11 THO1: 6,7 TPOX: 8,9 vWA: 17		
Cytogenetic Analysis:	The line is near-triploid with a modal number of 62 chromosomes. There are nearly 20 marker chromos commonly found in each cell; and normal N2, N3, N4, N5, N12, and N15 are not found. No norn chromosomes could be detected by Q-band analysis.		
Age:	62 years adult		
Gender:	male		

Ethnicity: Caucasian

Comments: The PC-3 was initiated from a bone metastasis of a grade IV prostatic adenocarcinoma from a 62-year-old Caucasian. [22363]
The cells exhibit low acid phosphatase and testosterone-5-alpha reductase activities.

Propagation: **ATCC complete growth medium:** The base medium for this cell line is ATCC-formulated F-12K Me Catalog No. 30-2004. To make the complete growth medium, add the following components to the medium: fetal bovine serum to a final concentration of 10%.
Atmosphere: air, 95%; carbon dioxide (CO_2), 5%
Temperature: 37.0°C

Subculturing: **Protocol:**

1. Remove and discard culture medium.
2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin- 0.53 mM EDTA solution to remove all trac serum that contains trypsin inhibitor.
3. Add 2.0 to 3.0 ml of Trypsin-EDTA solution to flask and observe cells under an inverted microscope cell layer is dispersed (usually within 5 to 15 minutes).
   Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
4. Add 6.0 to 8.0 ml of complete growth medium and aspirate cells by gently pipetting.
5. Add appropriate aliquots of the cell suspension to new culture vessels.
6. Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:3 to 1:6 is recommended
Medium Renewal: 2 to 3 times per week

Preservation: **Freeze medium:** Complete growth medium supplemented with 5% (v/v) DMSO
Storage temperature: liquid nitrogen vapor phase

Related Products: Recommended medium (without the additional supplements or serum described under ATCC Medium):ATC 2004
recommended serum:ATCC 30-2020

References:
22363: Kaighn ME, et al. Establishment and characterization of a human prostatic carcinoma cell line (F Invest. Urol. 17: 16-23, 1979. PubMed: 447482
22470: Chen TR. Chromosome identity of human prostate cancer cell lines, PC-3 and PPC-1. Cytogenet Genet. 62: 183-184, 1993. PubMed: 8428522
26302: Ohnuki Y, et al. Chromosomal analysis of human prostatic adenocarcinoma cell lines. Cancer Re: 524-534, 1980. PubMed: 7471073
32341: Sheng S, et al. Maspin acts at the cell membrane to inhibit invasion and motility of mammar prostatic cancer cells. Proc. Natl. Acad. Sci. USA 93: 11669-11674, 1996. PubMed: 8876194
32344: Umekita Y, et al. Human prostate tumor growth in athymic mice: inhibition by androgens stimulation by finasteride. Proc. Natl. Acad. Sci. USA 93: 11802-11807, 1996. PubMed: 8876218
32460: Carter RE, et al. Prostate-specific membrane antigen is a hydrolase with substrate and pharmac characteristics of a neuropeptidase. Proc. Natl. Acad. Sci. USA 93: 749-753, 1996. PubMed: 8570628
32486: Nupponen NN, et al. Genetic alterations in prostate cancer cell lines detected by comparative ge hybridization. Cancer Genet. Cytogenet. 101: 53-57, 1998. PubMed: 9460501
32488: Geiger T, et al. Antitumor activity of a PKC-alpha antisense oligonucleotide in combination with sta chemotherapeutic agents against various human tumors transplanted into nude mice. Anticancer Drug De: 35-45, 1998. PubMed: 9474241
32916: Su ZZ, et al. Surface-epitope masking and expression cloning identifies the human prostate carci tumor antigen gene PCTA-1 a member of the galectin gene family. Proc. Natl. Acad. Sci. USA 93: 7252- 1996. PubMed: 8692978

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These cells will be injected subcutaneously in the flank of nude mice. Jennifer Hadway, Lise Desjardins and Lisa Hoffmann will deal with the mice for the injecting. Lisa grows and handles the cells. All involved will be wearing lab coats, gloves, masks and safety goggles. Also the mice will be injected in the BSC in F5-104.

BIO-LHRI-0064

THE UNIVERSITY OF WESTERN ONTARIO
BIOHAZARDOUS AGENTS REGISTRY FORM
Revised Biohazards Subcommittee: January, 2007

This form must be completed by each Principal Investigator holding a grant administered by the University of Western Ontario where the use of biohazardous infectious agents are described in the experimental work proposed. The form must also be completed if animal work is proposed involving the use of biohazardous agents or animal carrying zoonotic agents infectious to humans. Containment Levels will be required in accordance with Laboratory Biosafety Guidelines, 3rd edition, 2004, Health Canada (HC) or Containment Standards for Veterinary Facilities, 1st edition 1996, Canadian Food Inspection Agency (CFIA).

Completed forms are to be returned to Occupational Health and Safety (Stevenson-Lawson Building, Room 60) for forward to the Biohazard Subcommittee. For questions regarding this form, please contact the Biosafety Coordinator at extension 81135. If there are changes to the information on this form (excluding grant title and funding agencies) modifications must be completed and sent to Occupational Health and Safety. See website: www.uwo.ca/humanresources

PRINCIPAL INVESTIGATOR Dr. Ting Yim Lee
SIGNATURE Ting Yim Lee
DEPARTMENT Imaging
ADDRESS 268 Grosvenor St.
PHONE NUMBER 6550 7 (lab) 34131 (office)
EMAIL tlee@lawsonimaging.ca

Location of experimental work to be carried out: Building(s) LHRI / AHS(UWO) UWO Room(s) FS-104 / 6024
*For work being performed at Institutions affiliated with the University of Western Ontario, the Safety Officer for the Institution where experiments will take place must sign the form prior to it being sent to Occupational Health and Safety (See Section 12.0, Approvals). For research being done at Lawson Health Research Institute, London Regional Cancer Centre, Child and Parent Research Institute or Robarts Research Institute, University Biosafety Committee members can also sign as the Safety Officer.

TITLE OF GRANT(S):
Proposed Dynamic Contrast Enhanced Computed Tomography Investigation Using Xenograft Subcutaneous Tumour Models in Rodents

PLEASE ATTACH A BRIEF DESCRIPTION OF YOUR WORK, SUCH A THE RESEARCH GRANT SUMMARY(S) THAT EXPLAINS THE BIOHAZARDS USED. PROJECTS SUBMITTED WITHOUT A SUMMARY WILL NOT BE REVIEWED.

FUNDING AGENCY/AGENCIES Astra Zeneca

Names of all personnel working under Principal Investigators supervision in this location:
i) Jennifer Hadway
ii) Claire Beppe
iii) Dominique Ouimet
iv) Lisa Hoffman
v)

DESCRIPTION MUST BE ATTACHED TO THIS FORM OR PROJECT WILL NOT BE REVIEWED*

1.0 Microorganisms

1.1 Does your work involve the use of microorganisms or biological agents of plant or animal origin (including but not limited to viruses, prions, parasites, bacteria)? ☐ YES ☑ NO
If no, please proceed to Section 2.0

1.2 Please complete the table below:

Name of Biological agent(s)	Is it known to be a human pathogen? YES/NO	Is it known to be an animal pathogen? YES/NO	Is it known to be a zoonotic agent? YES/NO	Maximum quantity to be cultured at one time?
	☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No	
	☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No	
	☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No	
	☐ Yes ☐ No	☐ Yes ☐ No	☐ Yes ☐ No	

1.3 For above named organism(s) or biological agent(s) circle HC or CFIA Containment Level required. 1 2 3

1.4 Source of microrganism(s) or biological agent(s)? _____

2.0 Cell Culture

2.1 Does your work involve the use of cell cultures? ☑ YES ☐ NO
If no, please proceed to Section 3.0

2.2 Please indicate the type of primary cells (ie. derived from fresh tissue) that will be grown in culture in the table below – NOT PRIMARY CELLS

Cell Type	Is this cell type used in your work?	Source of Primary Cell Culture Tissue
Human	☐ Yes ☐ No	
Rodent	☐ Yes ☐ No	
Non-human primate	☐ Yes ☐ No	
Other (specify)		

2.3 Please indicate the type of established cells that will be grown in culture in the table below.

Cell Type	Is this cell type used in your work?	Specific cell line(s)	Supplier / Source
Human	☑ Yes ☐ No	LoVo	ATCC
Rodent	☐ Yes ☐ No		
Non-human primate	☐ Yes ☐ No		
Other (specify)	☐ Yes ☐ No		

2.4 For above named cell types(s) circle HC or CFIA containment level required (1) 2 3 See attached sheet

* DESCRIPTION MUST BE ATTACHED TO THIS FORM OR PROJECT WILL NOT BE REVIEWED*

3.0 Use of Human Source Materials

3.1 Does your work involve the use of human source materials? ☐YES ☑NO
If no, please proceed to Section 4.0

3.2 Indicate if the following will be used in the laboratory

- Human blood (whole) or other bodily fluids ☐ YES ☑ NO If YES, Specify _____
- Human blood (fraction) or other bodily fluids ☐ YES ☑ NO If YES, Specify _____
- Human organs (unpreserved) ☐ YES ☑ NO If YES, Specify_____
- Human tissues (unpreserved) ☐ YES ☑ NO If YES, Specify_____

3.3 Is human source known to be infected with and infectious agent ☐ YES ☑ NO
If YES , please name infectious agent _____

3.4 For above named materials circle HC or CFIA containment level required. (1) 2 3

4.0 Genetically Modified Organisms and Cell lines

4.1 Will genetic modifications be made to the microorganisms, biological agents or cells described in Sections 1.0 and 2.0 ? ☐ YES ☑ NO
If no, please proceed to Section 5.0

4.2 Will genetic sequences from the following be involved:

- HIV ☐ YES ☑ NO
if YES specify_____
- HTLV 1 or 2 or genes from any CDC class 1 pathogens ☐ YES ☐ NO
if YES specify_____
- Other human or animal pathogen and or their toxins ☐ YES ☐ NO
if YES specify_____

4.3 Will intact genetic sequences be used from

- SV 40 Large T antigen ☐ YES ☑ NO If YES specify_____
- Known oncogenes ☑ YES ☐ NO If YES specify See attached ATCC. sheet

4.4 Will a live vector(s) (viral or bacterial) be used for gene transduction ☐ YES ☑ NO
If YES name virus_____.

4.5 List specific vector(s) to be used:_____

4.6 Will virus be replication defective ☐ YES ☐ NO

4.7 Will virus be infectious to humans or animals ☐ YES ☐ NO

4.8 Will this be expected to increase the Containment Level required ☐ YES ☐ NO

* DESCRIPTION MUST BE ATTACHED TO THIS FORM OR PROJECT WILL NOT BE REVIEWED*

5.0 Human Gene Therapy Trials

5.1 Will human clinical trials using the viral vector in 4.0 be conducted? ☐ YES ☑ NO
If no, please proceed to Section 6.0
If YES attach a full description of the make-up of the virus.

5.2 Will virus be able to replicate in the host? ☐ YES ☐ NO

5.3 How will the virus be administered? _____

5.4 Please give the Health Care Facility where the clinical trial will be conducted:_____

5.5 Has human ethics approval been obtained? ☐ YES ☐ NO

6.0 Animal Experiments

6.1 Will any of the agents listed be used in live animals? ☑ YES ☐ NO
If no, please proceed to section 7.0

6.2 Name of animal species to be used Nude rats

6.3 AUS protocol # 2006-044-08

6.4 If using murine cell lines, have they been tested for murine pathogens? ☐ YES ☐ NO N/A

7.0 Use of Animal species with Zoonotic Hazards

But tested anyways + Neg for murine pathogens
NO

7.1 Will any of the following animals or their organs, tissues, lavages or other bodily fluids including blood be used:

- Pound source dogs ☐ YES ☑ NO
- Pound source cats ☐ YES ☑ NO
- Sheep or goats ☐ YES ☑ NO
- Non- Human Primates ☐ YES ☑ NO If YES specify species _____
- Wild caught animals ☐ YES ☑ NO If YES specify species _____
  colony # _____

8.0 Biological Toxins

8.1 Will toxins of biological origin be used? ☐ YES ☑ NO
If no, please proceed to Section 9.0

8.2 If YES, please name the toxin_____

8.3 What is the LD_{50} (specify species) of the toxin_____

9.0 Import Requirements

9.1 Will the agent be imported? ☑ YES ☐ NO
If no, please proceed to Section 10.0
If yes, country of origin USA

9.2 Has an Import Permit been obtained from HC for human pathogens? ☐ YES ☑ NO N/A : cells Lab1 IBiohazard

9.3 Has an import permit been obtained from CFIA for animal pathogens? ☐ YES ☑ NO

9.4 Has the import permit been sent to OHS? ☐ YES ☑ NO
If yes, Permit # _____

10.0 Training Requirements for Personnel named on Form

All personnel named on the above form who will be using any of the above named agents are required to attend the following training courses given by OHS

- Biosafety
- Laboratory and Environmental/Waste Management Safety
- WHMIS

As the Principal Investigator, I have ensured that all of the personnel named on the form who will be using any of the biohazardous agents in Sections 1.0 to 9.0 have been trained.

SIGNATURE Tony Yu-Hsi Lee

11.0 Containment Levels

11.1 For the work described in sections 1.0 to 9.0, please circle the highest HC or CFIA Containment Level required. (1) 2 3

11.2 Has the facility been certified by OHS for this level of containment? ☑ YES ☐ NO

11.3 If yes, please give the date and permit number: _____

12.0 Approvals

UWO Biohazard Subcommittee

Signature G.M. Kidder Date 7 Dec. '07

Safety Officer for Institution where experiments will take place

Signature _____ Date Oct 12/2007

Safety Officer for University of Western Ontario (if different than above)

Signature J Stanley Date Dec 6/07

* DESCRIPTION MUST BE ATTACHED TO THIS FORM OR PROJECT WILL NOT BE REVIEWED*