

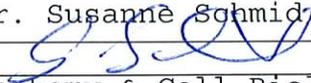
**THE UNIVERSITY OF WESTERN ONTARIO
BIOHAZARDOUS AGENTS REGISTRY FORM**
Approved Biohazards Subcommittee: March 27, 2009
Biosafety Website: www.uwo.ca/humanresources/biosafety/

This form must be completed by each Principal Investigator holding a grant administered by the University of Western Ontario or in charge of a laboratory/facility where the use of Level 1, 2 or 3 biohazardous agents is described in the laboratory or animal work proposed. The form must also be completed if any work is proposed involving animals carrying zoonotic agents infectious to humans or involving plants, fungi, or insects that require Public Health Agency of Canada (PHAC) or Canadian Food Inspection Agency (CFIA) permits.

This form must also be updated at least every 3 years or when there are changes to the biohazards being used.

Containment Levels will be established in accordance with Laboratory Biosafety Guidelines, 3rd edition, 2004, Public Health Agency of Canada (PHAC) or Containment Standards for Veterinary Facilities, 1st edition 1996, Canadian Food Inspection Agency (CFIA).

Completed forms are to be returned to Occupational Health and Safety, (OHS), (Support Services Building, Room 4190) for distribution to the Biohazard Subcommittee. For questions regarding this form, please contact the Biosafety Officer at extension 81135 or biosafety@uwo.ca. If there are changes to the information on this form (excluding grant title and funding agencies), contact Occupational Health and Safety for a modification form. See website: www.uwo.ca/humanresources/biosafety/

| | |
|---------------------------|--|
| PRINCIPAL INVESTIGATOR | Dr. Susanne Schmid |
| SIGNATURE |  |
| DEPARTMENT | Anatomy & Cell Biology |
| ADDRESS | Medical Science Building 470 |
| PHONE NUMBER | 519-661 2111 ext. 82668 |
| EMERGENCY PHONE NUMBER(S) | |
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Location of experimental work to be carried out: Building(s) MSB Room(s) 473

*For work being performed at Institutions affiliated with the University of Western Ontario, the Safety Officer for the Institution where experiments will take place must sign the form prior to its being sent to the University of Western Ontario Biosafety Officer (See Section 12.0, Approvals).

FUNDING AGENCY/AGENCIES: NSERC, OMHF, UWO
GRANT TITLE(S): Mechanisms underlying habituation of startle
Mechanisms underlying disruption of prepulse inhibition

PLEASE ATTACH A BRIEF DESCRIPTION OF YOUR WORK THAT EXPLAINS THE BIOHAZARDS USED AND HOW THEY WILL BE USED. PROJECTS SUBMITTED WITHOUT A SUMMARY WILL NOT BE REVIEWED.

Names of all personnel working under Principal Investigators supervision in this location:

| | |
|------------------------|-------|
| <u>Susanne Schmid</u> | _____ |
| <u>Tyler Brown</u> | _____ |
| <u>Farinna Pinnock</u> | _____ |
| <u>Laura Walker</u> | _____ |
| <u>Megan Clark</u> | _____ |

1.0 Microorganisms

1.1 Does your work involve the use of microorganisms or biological agents of plant or animal origin (including but not limited to viruses, prions, parasites, bacteria)? YES NO

If no, please proceed to Section 2.0

Do you use microorganisms that require a permit from the CFIA? YES NO

If YES, please give the name of the species. _____

What is the origin of the microorganism(s)? _____

Please describe the risk (if any) of escape and how this will be mitigated:

Please attach the CFIA permit.

Please describe any CFIA permit conditions:

1.2 Please complete the table below:

| Name of Biological agent(s)* | Is it known to be a human pathogen? YES/NO | Is it known to be an animal pathogen? YES/NO | Is it known to be a zoonotic agent? YES/NO | Maximum quantity to be cultured at one time? (in Litres) | Source/Supplier | PHAC or CFIA Containment Level |
|------------------------------|---|---|---|---|-----------------|---|
| | <input type="radio"/> Yes <input type="radio"/> No | <input type="radio"/> Yes <input type="radio"/> No | <input type="radio"/> Yes <input type="radio"/> No | | | <input type="radio"/> 1 <input type="radio"/> 2 <input type="radio"/> 3 |
| | <input type="radio"/> Yes <input type="radio"/> No | <input type="radio"/> Yes <input type="radio"/> No | <input type="radio"/> Yes <input type="radio"/> No | | | <input type="radio"/> 1 <input type="radio"/> 2 <input type="radio"/> 3 |
| | <input type="radio"/> Yes <input type="radio"/> No | <input type="radio"/> Yes <input type="radio"/> No | <input type="radio"/> Yes <input type="radio"/> No | | | <input type="radio"/> 1 <input type="radio"/> 2 <input type="radio"/> 3 |
| | <input type="radio"/> Yes <input type="radio"/> No | <input type="radio"/> Yes <input type="radio"/> No | <input type="radio"/> Yes <input type="radio"/> No | | | <input type="radio"/> 1 <input type="radio"/> 2 <input type="radio"/> 3 |

*Please attach a Material Safety Data Sheet or equivalent from the supplier.

2.0 Cell Culture

2.1 Does your work involve the use of cell cultures? YES NO

If no, please proceed to Section 3.0

2.2 Please indicate the type of primary cells (i.e. derived from fresh tissue) that will be grown in culture in the table below

| Cell Type | Is this cell type used in your work? | Source of Primary Cell Culture Tissue | AUS Protocol Number |
|-------------------|--|---------------------------------------|---------------------|
| Human | <input type="radio"/> Yes <input type="radio"/> No | | Not applicable |
| Rodent | <input type="radio"/> Yes <input type="radio"/> No | | |
| Non-human primate | <input type="radio"/> Yes <input type="radio"/> No | | |
| Other (specify) | <input type="radio"/> Yes <input type="radio"/> No | | |

2.3 Please indicate the type of established cells that will be grown in culture in the table below.

| Cell Type | Is this cell type used in your work? | Specific cell line(s)* | Supplier / Source |
|-------------------|--|------------------------|-------------------|
| Human | <input type="radio"/> Yes <input type="radio"/> No | | |
| Rodent | <input type="radio"/> Yes <input type="radio"/> No | | |
| Non-human primate | <input type="radio"/> Yes <input type="radio"/> No | | |
| Other (specify) | <input type="radio"/> Yes <input type="radio"/> No | | |

*Please attach a Material Safety Data Sheet or equivalent from the supplier. (For more information, see www.atcc.org)

2.4 For above named cell types(s) indicate PHAC or CFIA containment level required 1 2 3

3.0 Use of Human Source Materials

3.1 Does your work involve the use of human source materials? YES NO

If no, please proceed to Section 4.0

3.2 Indicate in the table below the Human Source Material to be used.

| Human Source Material | Source/Supplier /Company Name | Is Human Source Material Known to Be Infected With An Infectious Agent? YES/NO | Name of Infectious Agent (If applicable) | PHAC or CFIA Containment Level (Select one) |
|--|-------------------------------|---|--|---|
| Human Blood (whole) or other Body Fluid | | <input type="radio"/> Yes <input type="radio"/> No | | <input type="radio"/> 1 <input type="radio"/> 2 <input type="radio"/> 3 |
| Human Blood (fraction) or other Body Fluid | | <input type="radio"/> Yes <input type="radio"/> No | | <input type="radio"/> 1 <input type="radio"/> 2 <input type="radio"/> 3 |
| Human Organs or Tissues (unpreserved) | | <input type="radio"/> Yes <input type="radio"/> No | | <input type="radio"/> 1 <input type="radio"/> 2 <input type="radio"/> 3 |
| Human Organs or Tissues (preserved) | | <input type="radio"/> Yes <input type="radio"/> No | | <input type="radio"/> 1 <input type="radio"/> 2 <input type="radio"/> 3 |

4.0 Genetically Modified Organisms and Cell lines

4.1 Will genetic modifications be made to the microorganisms, biological agents, or cells described in Sections 1.0 and 2.0? YES NO If no, please proceed to Section 5.0

4.2 Will genetic modification(s) involving plasmids be done? YES, complete table below NO

| Bacteria Used for Cloning * | Plasmid(s) * | Source of Plasmid | Gene Transfected | Describe the change that results |
|-----------------------------|--------------|-------------------|------------------|----------------------------------|
| | | | | |

* Please attach a Material Data Sheet or equivalent if available.

4.3 Will genetic modification(s) involving viral vectors be done? YES, complete table below NO

| Virus Used for Transduction * | Vector(s) * | Source of Vector | Gene Transfected | Describe the change that results |
|-------------------------------|-------------|------------------|------------------|----------------------------------|
| | | | | |

* Please attach a Material Safety Data Sheet or equivalent.

4.4 Will genetic sequences from the following be involved?

- ◆ HIV YES, please specify _____ NO
- ◆ HTLV 1 or 2 or genes from any Level 1 or Level 2 pathogens YES, specify _____ NO
- ◆ SV 40 Large T antigen YES NO
- ◆ E1A oncogene YES NO
- ◆ Known oncogenes YES, please specify _____ NO
- ◆ Other human or animal pathogen and or their toxins YES, please specify _____ NO

4.5 Will virus be replication defective? YES NO

4.6 Will virus be infectious to humans or animals? YES NO

4.7 Will this be expected to increase the containment level required? YES NO

5.0 Human Gene Therapy Trials

5.1 Will human clinical trials be conducted using the viral vector in 4.0? YES NO
 If no, please proceed to Section 6.0 If YES attach a full description of the make-up of the virus.

5.2 Will virus be able to replicate in the host? YES NO

5.3 How will the virus be administered? _____

5.4 Please give the Health Care Facility where the clinical trial will be conducted: _____

5.5 Has human ethics approval been obtained? YES, number: _____ NO PENDING

6.0 Animal Experiments

6.1 Will live animals be used? YES NO If no, please proceed to section 7.0

6.2 Name of animal species to be used rats

6.3 AUS protocol # 2008-010 and 2008-006

6.4 Will any of the agents listed be used in live animals YES, specify: _____ NO

10.0 Plants Requiring CFIA Permits

10.1 Do you use plants that require a permit from the CFIA? YES NO
If no, please proceed to Section 11.0

10.2 If YES, please give the name of the species. _____

10.3 What is the origin of the plant? _____

10.4 What is the form of the plant (seed, seedling, plant, tree...)? _____

10.5 What is your intention? Grow and maintain a crop "One-time" use

10.6 Do you do any modifications to the plant? YES NO
If yes, please describe: _____

10.7 Please describe the risk (if any) of loss of the material from the lab and how this will be mitigated:

10.8 Is the CFIA permit attached? YES NO

10.9 Please describe any CFIA permit conditions:

11.0 Import Requirements

11.1 Will any of the above agents be imported? YES, please give country of origin _____
If no, please proceed to Section 10.0 NO

11.2 Has an Import Permit been obtained from HC for human pathogens? YES NO

11.3 Has an import permit been obtained from CFIA for animal or plant pathogens? YES NO

11.4 Has the import permit been sent to OHS? YES, please provide permit # _____ NO

12.0 Training Requirements for Personnel Named on Form

All personnel named on the above form who will be using any of the above named agents are required to attend the following training courses given by OHS:

- ◆ Biosafety
- ◆ Laboratory and Environmental/Waste Management Safety
- ◆ WHMIS (Western or equivalent)
- ◆ Employee Health and Safety Orientation

As the Principal Investigator, I have ensured that all of the personnel named on the form who will be using any of the biohazardous agents in Sections 1.0 to 9.0 have been trained.

SIGNATURE _____

Description of procedures

We perform behavioural experiments using adult Sprague Dawley rats. Rats are handled and placed into startle boxes in order to measure startle responses. Some animals will be injected systemically with specific neurotransmitter agonists and antagonists. Most of them are toxic in higher doses. We use and store them in microgram quantities.

Other animals undergo stereotaxic surgery in order to have chronic cannulae implanted into their skull. They also are injected with tiny amounts (nanomol quantities) of specific neurotransmitter agonists and antagonists.

We transcardially perfuse these animals after experimentation and remove brains from the skull. We perform tissue slicing from frozen brains and stain them using classical histological procedures.

We also perform electrophysiological experiments in acute rat brain slices. Animals are anaesthetized and decapitated for this procedure. Brains are removed and sliced, using a standard vibrating microtome. Slices are kept in a beaker for a day and transferred to the microscope for experiments. They might get perfused with neurotransmitter agonists and antagonists during these experiments.