Modification Form for Permit BIO-LHRI-0064

Permit Holder: Ting Yim Lee

Approved Personnel
(Please stroke out any personnel to be removed)

Adam Blais
Timothy Yeung
Laura Morrison
Lise Desjardins
Mark Dekaban
Errol Stewart
Jennifer Hadway
Joo Ho Tai

Additional Personnel
(Please list additional personnel here)

Becky McGirr
Carlie Charlton
Choi-Fong Cho

	Please stroke out any approved Biological Agent(s) to be removed	Write additional Biological Agent(s) for approval below. Give the full name
Approved Microorganisms		LNCap clone FGC ATCC CRL-1740
Approved Primary and Established Cells	Other [primary] VX2 papilloma virus grown in rabbit thigh. Human [established] LoVo, PC3, NCI-H1299, HT-29. Rodent [established] C6-glioma, N1-S1 Fudr	
Approved Use of Human Source Material		
Approved Genetic Modifications (Plasmids/Vectors)		
Approved Use of Animals	Mice, rats, rabbits	Nude mice
Approved Biological Toxin(s)		

Approved Gene Therapy		
Approved Plants and Insects		

** PLEASE ATTACH A MATERIAL SAFETY DATA SHEET OR EQUIVALENT FOR NEW BIOLOGICAL AGENTS.*
*** PLEASE ATTACH A BRIEF DESCRIPTION OF THE WORK THAT EXPLAINS THE BIOLOGICAL AGENTS USED AND HOW THEY WILL BE STORED, USED AND DISPOSED OF..*

As the Principal Investigator, I have ensured that this project will follow the Western Biosafety Guidelines and Procedures Manual for Containment Level 1 2 Laboratories (and the Level 3 Facilities Manual for Level 3 projects). I will ensure that UWO faculty, staff and students working in my laboratory have an up-to-date Hazard Communication Form, found at http://www.shs.uwo.ca/workplace/newposition.htm

Signature of Permit Holder: Tony Yan Lee

Current Classification: 1 **Containment Level for Added Biohazards:** Level 1

Date of Last Biohazardous Agents Registry Form: May 30, 2011

Date of Last Modification (if applicable):

BioSafety Officer(s): Dec 21/2011

Chair, Biohazards Subcommittee: **Date:**

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Product Description

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Cell Biology

ATCC® Number: CRL-1740™ Order this Item **Price:** $279.00

Will use this blank section for use of cells

These LNCaP cells are injected in the nude mice the same way the PC3 cells are.

It is a ~~Subq~~ subcutaneous injection in the neck or flank. Once the tumor is ~ 0.5cm the mouse will be imaged and then euthanized without recovery at the end of the imaging procedure.

These cells are handled by John Lewis's lab at LRCP - also under his BHARF

Designations:	LNCaP clone FGC
Depositors:	JS Horoszewicz
Biosafety Level:	1
Shipped:	frozen
Medium & Serum:	See Propagation
Growth Properties:	adherent, single cells and loosely attached clusters
Organism:	*Homo sapiens* (human)
Morphology:	epithelial
	
Source:	**Organ:** prostate **Disease:** carcinoma **Derived from metastatic site:** left supraclavicular lymph node
Cellular Products:	human prostatic acid phosphatase; prostate specific antigen
Permits/Forms:	In addition to the MTA mentioned above, other ATCC and/or regulatory permits may be required for the transfer of this ATCC material. Anyone purchasing ATCC material is ultimately responsible for obtaining the permits. Please click here for information regarding the specific requirements for shipment to your location.
Restrictions:	Distribution of this material for commercial purposes will require execution of a Non-exclusive License Agreement. At the time of placing an order, customers must send a request to licensing@ATCC .org. Orders will be shipped when Customer Service receives confirmation from our Licensing officer.
Isolation:	**Isolation date:** 1977
Applications:	transfection host
Receptors:	androgen receptor, positive; estrogen receptor, positive [23045]
Tumorigenic:	Yes
DNA Profile (STR):	Amelogenin: X,Y CSF1PO: 10,11 D13S317: 10,12 D16S539: 11 D5S818: 11,12 D7S820: 9.1,10.3 THO1: 9 TPOX: 8,9 vWA: 16,18
Cytogenetic Analysis:	This is a hypotetraploid human cell line. The modal chromosome number was 84, occurring in 22% of cells. However, cells with chromosome counts of 86 (20%) and 87 (18%) also occurred at high frequencies. The rate of cells with higher ploidies was 6.0%.
Age:	50 years adult
Gender:	male
Ethnicity:	Caucasian
Comments:	LNCaP clone FGC was isolated in 1977 by J.S. Horoszewicz, et al., from a needle aspiration biopsy of the left supraclavicular lymph node of a 50-year-old Caucasian male (blood type B+) with confirmed diagnosis of metastatic prostate carcinoma. These cells are responsive to 5-alpha-dihydrotestosterone (growth modulation and acid phosphatase production). The cells do not produce a uniform monolayer, but grow in clusters which should be broken apart by repeated pipetting when subcultures are prepared. They attach only lightly to the substrate, do not become confluent and rapidly acidify the medium. Growth is very slow. The cells should be allowed to incubate undisturbed for the first 48 hours after subculture. When flask cultures are shipped, the majority of the cells become detached from the flask and float in the medium. Upon receipt, incubate the flask (in the usual position for monolayer cultures) for 24 to 48 hours to allow the cells to re-attach. The medium can then be removed and replaced with fresh medium.

Related Links

NCBI Entrez Search

Cell Micrograph

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Frequently Asked Questions

Material Transfer Agreement

Technical Support

Related Cell Culture Products

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Product Information Sheet

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All prices are listed in U.S. dollars and are subject to change without notice. A discount off the current list price will be applied to most cultures for nonprofit institutions in the United States. Cultures that are ordered as test tubes or flasks will carry an additional laboratory fee. Fees for permits, shipping, and handling may apply.

Back to my Search

THE UNIVERSITY OF WESTERN ONTARIO
BIOLOGICAL AGENTS REGISTRY FORM
Approved Biohazards Subcommittee: October 14, 2010
Biosafety Website: www.uwo.ca/humanresources/biosafety/

This form must be completed by each Principal Investigator holding a grant administered by the University of Western Ontario (UWO) or in charge of a laboratory/facility where the use of Level 1, 2 or 3 biological agents is described in the laboratory or animal work proposed. The form must also be completed if any work is proposed involving animals carrying zoonotic agents infectious to humans or involving plants, fungi, or insects that require Public Health Agency of Canada (PHAC) or Canadian Food Inspection Agency (CFIA) permits.

This form must be updated at least every 3 years or when there are changes to the biological agents being used.

Containment Levels will be established in accordance with Laboratory Biosafety Guidelines, 3rd edition, 2004, Public Health Agency of Canada (PHAC) or Containment Standards for Veterinary Facilities, 1st edition 1996, Canadian Food Inspection Agency (CFIA).

Completed forms are to be returned to Occupational Health and Safety, (OHS), (Support Services Building, Room 4190) for distribution to the Biohazards Subcommittee. For questions regarding this form, please contact the Biosafety Officer at extension 81135 or biosafety@uwo.ca. If there are changes to the information on this form (excluding grant title and funding agencies), contact Occupational Health and Safety for a modification form. See website: www.uwo.ca/humanresources/biosafety

PRINCIPAL INVESTIGATOR	Ting Yim Lee
DEPARTMENT	UWO/Schulich/Robarts Research Institute
ADDRESS	
PHONE NUMBER	Office 24131. Lab 65507 (Jennifer Hadway)
EMERGENCY PHONE NUMBER(S)	
EMAIL	tlee@imaging.robarts.ca

Location of experimental work to be carried out: Building(s) LHRI/St Joes___ Room(s)__F5-115, F5-104, F6-127a, B5-251, C0-232, all cells kept and cell work done by Dr Lisa Hoffman in F4-127a

*For work being performed at Institutions affiliated with the University of Western Ontario, the Safety Officer for the Institution where experiments will take place must sign the form prior to its being sent to the University of Western Ontario Biosafety Officer (See Section 15.0, Approvals).

FUNDING AGENCY/AGENCIES:_____
GRANT TITLE(S): _____
_____
_____

List all personnel working under Principal Investigators supervision in this location:

Name	UWO E-mail Address	Date of Biosafety Training
Jennifer Hadway	jhadway@uwo.ca or lawsonimaging.ca	* June 13, 2005 **On file**
Lisa Hoffman	lhoffmann@lawsonimaging.ca	
Lise Desjardins	ldesjar5@uwo.ca	
Laura Morrison	lmorri67@uwo.ca	
Timothy Yeung	Timothy.Yeung@lhsc.on.ca	
Adam Blais	ablais@imaging.robarts.ca	
Joo Ho Tai	jtai@imaging.robarts.ca	* May 28, 2002
Errol Stewart	estewart@imaging.robarts.ca	* June 15, 2005
Mark Dekaban	mdekaban@imaging.robarts.ca	* June 26, 2006

* added by OH+S

Please explain the biological agents and/or biohazardous substances used and how they will be stored, used and disposed of. Projects without this description will not be reviewed.

Other (Rabbit): VX2 rabbit papilloma virus – is a rabbit tumor that we maintain in the rabbit thigh muscle, then every 3 – 6 weeks we euthanize the rabbit take out small pieces of tumor and mix it with HBSS and inject into a new rabbit thigh and (usually) into the liver (separate protocol) of 2 rabbits on the same day for a liver tumor study. We do also maintain small pieces in -80C freezer but it does not grow as fast or as reliably from the freezer stock. The rabbits are incinerated and the remaining sample that is not used goes into biohazardous waste for autoclaving before disposal. For use in pending protocol 2006-078 and is carried on 2009-087

2.3 Established cells:

Human:

(1) LoVo – ATCC # CCL-229 derived from human colorectal adenocarcinoma metastatic site, ATCC biosafety level 1. This line is injected into the neck/shoulder region of nude rats and then imaged on the CT scanner for several weeks after. The rat is sent for incineration at the end of the study and the tumor is usually removed for histology and the remainder not used is disposed of in biohazardous waste for incineration. Protocol 2010-288

(2) PC3 cells - ATCC CRL-1435 human prostate tumor, ATCC biosafety level 1. This line is injected into the neck/shoulder region of nude mice for imaging purposes (uPET/CT). Again at the end of the study the tumor is removed and the mouse sent for incineration and the tumor kept for histology purposes. Protocol 2009-013

(3) NCI – H1299 – ATCC # CRL-5803 Human non-small cell lung carcinoma, ATCC biosafety level 1. This is injected into the lungs mixed with matrigel, in nude rats, is followed with uPET and CT imaging. Tumor removed at end of study for histology and rat incinerated, histology done on tumor. Protocol 2009-027

(4) HT-29 – ATCC # HTB-38 derived from human colorectal adenocarcinoma, ATCC level 1, this line is injected into the neck/shoulder region of nude ratsand then imaged (CT) for several weeks after. The rat is sent for incineration at the endof the study and the tumor is removed for histology and the remainder not used is disposed of in biohazardous waste.
Protocol mod going in for 2010-288 once protocol released from eSirius, is similar to Lovo cells but added this one as we had a lot of problems with the lovos growing last year

Rodent:

(1) C6-glioma - ATCC # CCL-107 rat brain tumor, ATCC biosafety level 1. These tumor cells are stereotactically injected into the brain of rats and CT and uPET imaging some radiotherapy done. Brain is removed at end of study for histology and rat is incinerated. Protocol 2010 -009

(2)N1–S1 Fudr – ATCC # CRL 1603 derived from the liver of Sprague Dawley rats, rat hepatoma, ATCC biolevel 1, this cell line is injected directly into the liver of SD rats to induce livers tumors which will then be imaged on a CT scanner. Following final day of scanning the liver is removed and the rat is disposed of in biohazard animal area for incineration. Is active mod in eSirius, Protocol 2007-028-01

Please include a one page research summary or teaching protocol.

See Next Page

Human:

(1) LoVo – ATCC # CCL-229 derived from human colorectal adenocarcinoma metastatic site. ATCC biosafety level 1: The purpose of this study is to use a non-invasive imaging technology to examine blood vessel development during growth of tumors in animal models. This study is designed to assess the changes in blood vessels that occur with cancer progression, and cannot be conducted ex vivo or in phantoms; the use of animals will provide us with a means to refine and modify the imaging process prior to potential use in humans.
With NIRS we are trying to see if we can measure tumor physiology using 2 different optical dyes.

(2) PC3 cells - ATCC CRL-1435 human prostate tumor. ATCC biosafety level 1: The mice will have PC3 cells injected and then be checked every 4 - 5 days for tumor growth. Once the tumors are palpable we will start the imaging portion of the study. They will have CT imaging done then 2 - 3 days later microPET imaging done. Both are done with isoflurane anesthetic, and a tail vein catheter. The purpose of this study is to use imaging techniques (PET and CT) to characterize vascular changes and tumor physiology during tumor growth. Imaging can be used to non-invasively monitor many features of tumor physiology. The characterization of specific changes in tumor physiology during growth have important implications for treatment planning.

(3) NCI – H1299 – ATCC # CRL-5803 Human non-small cell lung carcinoma, ATCC biosafety level 1: This study is a precursor to a treatment model, this tumor has been grown in nude mice, we are transferring the model to nude rats to more easily enable us to (see via) CT image the tumor as it grows and the related blood flow using CT perfusion techniques. Human tumor cells from ATCC will be mixed with matrigel then injected intrathoracicly in anesthetized rats. CT (lung) imaging with contrast will be done to follow the tumor growth both visually and also with CT perfusion. This will allow us to learn when the best time line for treatment will be in a follow up protocol.

(4) HT-29 – ATCC # HTB-38 derived from human colorectal adenocarcinoma, ATCC level 1: This cell line will be added protocol 2010-288 as a modification once it is released. The research objectives will be exactly the same as for the Lovo cells in the same protocol and will be used as a substitute not an addition if we are still having difficulties with the Lovo cell line.

Rodent:

(1) C6-glioma - ATCC # CCL-107 rat brain tumor, ATCC biosafety level 1: This study is to see the results of radiotherapy on a C6 glioma tumor in the rat brain and also to assess the ability of the high resolution CT imaging to see changes in perfusion, blood flow etc before and after helical tomotherapy treatment. We will use a control group to compare the changes in the tumor after treatment also and for histology comparison.
We are added 2 new methods for monitoring the growth of tumors in our animals. These are carbogen-challenged perfusion imaging(DCE-CT) and 18-Flurodeoxyglucose (FDG) positron emission tomography (FDG-PET). FDG is also a radioactive material. We are adding a new drug to treat some of the animals. This drug is called Bevacizumab (trade name Avastin), it is an antibody that inhibits the growth of blood vessels

(2)N1–S1 Fudr – ATCC # CRL 1603 derived from the liver of Sprague Dawley rats, rat hepatoma, ATCC biolevel 1: The purpose of this protocol is to cause fibrosis, cirrhosis and tumors in the rat liver and then follow the changes in the liver with contrast enhanced Computed Tomography. This will be done either with the IP injection of CCl4 or DEN given in the water that will first cause cirrhosis and eventually cause tumors to develop. Some of these rats will be followed with CT imaging and a few will be left to develop tumors that will be used to continue the tumor line in other rats. The rats will be imaged with pentobarbital anesthetic weekly with CT perfusion imaging to follow blood flow changes in the liver

1.0 Microorganisms

1.1 Does your work involve the use of biological agents? O YES **X** NO
(non-pathogenic and pathogenic biological agents including but not limited to bacteria and other microorganisms, viruses, prions, parasites or pathogens of plant or animal origin)? If no, please proceed to Section 2.0

Do you use microorganisms that require a permit from the CFIA? O YES **X** NO
If YES, please give the name of the species. _____
What is the origin of the microorganism(s)? _____
Please describe the risk (if any) of escape and how this will be mitigated:

_____
_____
_____

Please attach the CFIA permit.
Please describe any CFIA permit conditions:

_____
_____
_____

1.2 Please complete the table below:

Name of Biological Agent(s)* (Be specific)	Is it known to be a human pathogen? YES/NO	Is it known to be an animal pathogen? YES/NO	Is it known to be a zoonotic agent? YES/NO	Maximum quantity to be cultured at one time? (in Litres)	Source/ Supplier	PHAC or CFIA Containment Level
	O Yes O No	O Yes O No	O Yes O No			O 1 O 2 O 2+ O 3
	O Yes O No	O Yes O No	O Yes O No			O 1 O 2 O 2+ O 3
	O Yes O No	O Yes O No	O Yes O No			O 1 O 2 O 2+ O 3
	O Yes O No	O Yes O No	O Yes O No			O 1 O 2 O 2+ O 3

*Please attach a Material Safety Data Sheet or equivalent from the supplier.

2.0 Cell Culture

2.1 Does your work involve the use of cell cultures? **X** YES O NO
If no, please proceed to Section 3.0

2.2 Please indicate the type of primary cells (i.e. derived from fresh tissue) that will be grown in culture:

Cell Type	Is this cell type used in your work?	Source of Primary Cell Culture Tissue	AUS Protocol Number
Human	O Yes O No		Not applicable
Rodent	O Yes O No		
Non-human primate	O Yes O No		
Other (specify)	**X** Yes O No	VX2 papilloma virus, grown in rabbit thigh	2009-087 & 2006-078

2.3 Please indicate the type of established cells that will be grown in culture in:

Cell Type	Is this cell type used in your work?	Specific cell line(s)*	Containment Level of each cell line	Supplier / Source of cell line(s)
Human	X Yes O No	Lovo PC3 NCI-H1299 HT-29	1 1 1 1	All ATCC
Rodent	X Yes O No	C6 – glioma N1-S1 Fudr	1 1	ATCC ATCC
Non-human primate	O Yes O No			
Other (specify)	O Yes O No			

No NHP ?

*Please attach a Material Safety Data Sheet or equivalen from the supplier. (For more information, see www.atcc.org)

2.4 For above named cell types(s) indicate PHAC or CFIA containment level required X 1 O 2 O 2+ O 3

3.0 Use of Human Source Materials

3.1 Does your work involve the use of human source materials? O YES X NO
If no, please proceed to Section 4.0

3.2 Indicate in the table below the Human Source Material to be used.

Human Source Material	**Source/Supplier /Company Name**	**Is Human Source Material Infected With An Infectious Agent? YES/UNKNOWN**	**Name of Infectious Agent (If applicable)**	**PHAC or CFIA Containment Level (Select one)**
Human Blood (whole) or other Body Fluid		O Yes O Unknown		O 1 O 2 O 2+ O 3
Human Blood (fraction) or other Body Fluid		O Yes O Unknown		O 1 O 2 O 2+ O 3
Human Organs or Tissues (unpreserved)		O Yes O Unknown		O 1 O 2 O 2+ O 3
Human Organs or Tissues (preserved)		Not Applicable		Not Applicable

4.0 Genetically Modified Organisms and Cell lines

4.1 Will genetic modifications be made to the microorganisms, biological agents, or cells described in Sections 1.0 and 2.0? O YES X NO If no, please proceed to Section 5.0

4.2 Will genetic modification(s) involving plasmids be done? O YES, complete table below O NO

Bacteria Used for Cloning *	Plasmid(s) **	Source of Plasmid	Gene Transfected	Describe the change that results from transformation or tranfection

* Please attach a Material Data Sheet or equivalent if available.
** Please attach a plasmid map.

4.3 Will genetic modification(s) of bacteria and/or cells involving viral vectors be made?
O YES, complete table below O NO

Virus Used for Vector Construction	Vector(s) *	Source of Vector	Gene(s) Transduced	Describe the change that results from transduction

* Please attach a Material Safety Data Sheet or equivalent.

4.4 Will genetic sequences from the following be involved?

- HIV O YES, please specify _____ O NO
- HTLV 1 or 2 or genes from any Level 1 or Level 2 pathogens O YES, specify _____ O NO
- SV 40 Large T antigen O YES O NO
- E1A oncogene O YES O NO
- Known oncogenes O YES, please specify _____ O NO
- Other human or animal pathogen and or their toxins O YES, please specify _____ O NO

4.5 Will virus be replication defective? O YES O NO

4.6 Will virus be infectious to humans or animals? O YES O NO

4.7 Will this be expected to increase the containment level required? O YES O NO

5.0 Human Gene Therapy Trials

5.1 Will human clinical trials be conducted involving a biological agent? O YES X NO
(including but not limited to microorganisms, viruses, prions, parasites or pathogens of plant or animal origin)
If no, please proceed to Section 6.0

5.2 If YES, please specify which biological agent will be used: _____
Please attach a full description of the biological agent.

5.2 Will the biological agent be able to replicate in the host? O YES O NO

5.3 How will the biological agent be administered? _____

5.4 Please give the Health Care Facility where the clinical trial will be conducted:_____

5.5 Has human ethics approval been obtained? O YES, number: _____ O NO O PENDING

6.0 Animal Experiments

6.1 Will live animals be used? X YES O NO If no, please proceed to section 7.0

6.2 Name of animal species to be used__mice, rats and rabbits_____

6.3 AUS protocol # _See protocol numbers on description pages 2 and 3_____

6.4 Will any of the agents listed in section 4.0 be used in live animals O YES, specify: _n/a___ O NO

6.5 Will the agent(s) be shed by the animal: O YES X NO, please justify:
_____All cells are injected into the animal and are not shed by the animal once injected_____

7.0 Use of Animal species with Zoonotic Hazards

7.1 Will any animals with zoonotic hazards or their organs, tissues, lavages or other body fluids including blood be used (see list below)? O YES X No If no, please proceed to section 8.0

7.2 Will live animals be used? O YES O No

7.3 If yes, please specify the animal(s) used:

- Pound source dogs O YES O NO
- Pound source cats O YES O NO
- Cattle, sheep or goats O YES, please specify species _____ O NO
- Non-human primates O YES, please specify species _____ O NO
- Wild caught animals O YES, please specify species & colony # _____ O NO
- Birds O YES, please specify species _____ O NO
- Others (wild or domestic) O YES, please specify _____ O NO

7.4 If no live animals are used, please specify the source of the specimens:

_____

8.0 Biological Toxins

8.1 Will toxins of biological origin be used? O YES X NO If no, please proceed to Section 9.0

8.2 If YES, please name the toxin(s)_____
Please attach information, such as a Material Safety Data Sheet, for the toxin(s) used.

8.3 What is the LD_{50} (specify species) of the toxin_____

8.4 How much of the toxin is handled at one time*? _____

8.5 How much of the toxin is stored*? _____

8.6 Will any biological toxins be used in live animals? O YES, Please provide details:_____ O NO

*For information on biosecurity requirements, please see:
http://www.uwo.ca/humanresources/docandform/docs/healthandsafety/biosafety/Biosecurity_Requirements.pdf

9.0 Insects

9.1 Do you use insects? O YES X NO If no, please proceed to Section 10.0

9.2 If YES, please give the name of the species. _____

9.3 What is the origin of the insect? _____

9.4 What is the life stage of the insect? _____

9.5 What is your intention? O Initiate and maintain colony, give location: _____
O "One-time" use, give location: _____

9.6 Please describe the risk (if any) of escape and how this will be mitigated:

_____
_____

9.7 Do you use insects that require a permit from the CFIA permit? O YES O NO
If YES, Please attach the CFIA permit & describe any CFIA permit conditions:

_____

_____

10.0 Plants

10.1 Do you use plants? O YES X NO If no, please proceed to Section 11.0

10.2 If YES, please give the name of the species. _____

10.3 What is the origin of the plant? _____

10.4 What is the form of the plant (seed, seedling, plant, tree...)? _____

10.5 What is your intention? O Grow and maintain a crop O "One-time" use

10.6 Do you do any modifications to the plant? O YES O NO
If yes, please describe: _____

_____

10.7 Please describe the risk (if any) of loss of the material from the lab and how this will be mitigated:

_____

_____

_____

10.8 Is the CFIA permit attached? O YES O NO
If YES, Please attach the CFIA permit & describe any CFIA permit conditions:

_____

_____

11.0 Import Requirements

11.1 Will any of the above agents be imported? O YES, please give country of origin_____ X NO
If no, please proceed to Section 12.0. ATCC cells now come from Cedarlane in Canada

11.2 Has an Import Permit been obtained from HC for human pathogens? O YES O NO

11.3 Has an import permit been obtained from CFIA for animal or plant pathogens? O YES O NO

11.4 Has the import permit been sent to OHS? O YES, please provide permit # _____ O NO

12.0 Training Requirements for Personnel Named on Form

All personnel named on the above form who will be using any of the above named agents are required to attend the following training courses given by OHS:

- Biosafety
- Laboratory and Environmental/Waste Management Safety
- WHMIS (Western or equivalent)
- Employee Health and Safety Orientation

As the Principal Investigator, I have ensured that all of the personnel named on the form who will be using any of the biological agents in Sections 1.0 to 9.0 have been trained.

SIGNATURE Tony Yan Lee

13.0 Containment Levels

13.1 For the work described in sections 1.0 to 9.0, please indicate the highest HC or CFIA Containment Level required. X 1 O 2 O 2+ O 3

13.2 Has the facility been certified by OHS for this level of containment?
O YES, date of most recent biosafety inspection: _____
O NO, please certify
X NOT REQUIRED for Level 1 containment

13.3 Please indicate permit number (not applicable for first time applicants): _____

14.0 Procedures to be Followed

14.1 Please describe additional risk reduction measures will be taken beyond containment level 1, 2, 2+ or 3 measures, that are unique to this agent.

___When all agents are handled those handling and assisting with holding animals with be wearing lab coats and gloves and with the nude animals the injections are done within a BSC so the sash protects our faces_____

_____

14.2 Please outline what will be done if there is an exposure to the biological agents listed, such as a needlestick injury or an accidental splash:

___If there is an accidental needlestick or splash, if required eyewash station will be used to clear eyes/face, if needle stick then area will be run under water and made to bleed then person will go to OH&S_____

14.3 As the Principal Investigator, I will ensure that this project will follow the Western Biosafety Guidelines and Procedures Manual for Containment Level 1 & 2 Laboratories (and the Level 3 Facilities Manual for Level 3 projects). I will ensure that UWO faculty, staff and students working in my laboratory have an up-to-date Hazard Communication Form, found at http://www.wph.uwo.ca/

SIGNATURE Terry M Lee Date: May 12 2011.

15.0 Approvals

1) UWO Biohazards Subcommittee: SIGNATURE: _____
Date: 10 May 2011

2) Safety Officer for the University of Western Ontario
SIGNATURE: J Stanley
Date: May 27, 2011

3) Safety Officer for Institution where experiments will take place (if not UWO):
SIGNATURE: _____
Date: May 12, 2011

Approval Number: _____ Expiry Date (3 years from Approval): _____

Special Conditions of Approval:

- People on page 1 must have Biosafety training.

Designations:	LoVo
Depositors:	M Romsdahl
Biosafety Level:	1
Shipped:	frozen
Medium & Serum:	See Propagation
Growth Properties:	adherent
Organism:	*Homo sapiens* (human)
Morphology:	epithelial
Source:	**Organ:** colon **Tumor Stage:** Dukes' type C, grade IV **Disease:** colorectal adenocarcinoma **Derived from metastatic site:** left supraclavicular region
Cellular Products:	carcinoembryonic antigen (CEA) 908 ng/10 exp6 cells/10 days
Permits/Forms:	In addition to the MTA mentioned above, other ATCC and/or regulatory permits may be required for the transfer of this ATCC material. Anyone purchasing ATCC material is ultimately responsible for obtaining the permits. Please click here for information regarding the specific requirements for shipment to your location.
Applications:	transfection host (Roche FuGENE® Transfection Reagents)
Tumorigenic:	Yes
Oncogene:	myc +; myb + ; ras +; fos +; p53 +; sis -; abl -; ros -; src -
Antigen Expression:	HLA A11, B15, B17, Cw1, Cw3; blood type B
DNA Profile (STR):	Amelogenin: XY CSF1PO: 10,11,13,14 D13S317: 8, 11 D16S539: 9, 12 D5S818: 11, 12, 13 D7S820: 9.3,10, 11 THO1: 9.3 TPOX: 8,9 vWA: 17,18
Cytogenetic Analysis:	The stemline chromosome number is hyperdiploid with the 2S component occurring at about 2.7% and 3 marker chromosomes were common to all S metaphases. Karyotypes were generally homogeneous and stable.
Isoenzymes:	ES-D, 1 G6PD, B PGD, A PGM1, 2 PGM3, 1-2
Age:	56 years
Gender:	male
Comments:	LoVo was initiated in 1971 from a fragment of a metastatic tumor nodule in the left supraclavicular region of a 56-year-old Caucasian male patient with a histologically proven diagnosis of adenocarcinoma of the colon. [1049] The cells are negative for expression of CSAp (CSAp-) and colon antigen 3. The line is positive for expression of c-myc, K-ras, H-ras, N-ras, Myb, sis and fos oncogenes. [22861] Myb, and fos oncogenes. [22861] N-myc and sis oncogene expression were not detected. [22861] Tumor specific nuclear matrix proteins CC-3 and CC-4 are expressed. [23341]
Propagation:	**ATCC complete growth medium:** The base medium for this cell line is ATCC-formulated F-12K Medium, Catalog No. 30-2004. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 10%. **Temperature:** 37.0°C
Subculturing:	**Subcultivation Ratio:** A subcultivation ratio of 1:3 to 1:10 is recommended **Medium Renewal:** 2 to 3 times per week Remove medium, and rinse with 0.25% trypsin, 0.03% EDTA solution. Remove the solution and add an additional 1 to 2 ml of trypsin-EDTA solution. Allow the flask to sit at room temperature (or at 37C) until

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Designations:	PC-3
Depositors:	ME Kaighn
Biosafety Level:	1
Shipped:	frozen
Medium & Serum:	See Propagation
Growth Properties:	adherent (The cells form clusters in soft agar and can be adapted to suspension growth)
Organism:	*Homo sapiens* (human)
Morphology:	epithelial
	
Source:	**Organ:** prostate **Tumor Stage:** grade IV **Disease:** adenocarcinoma **Derived from metastatic site:** bone
Permits/Forms:	In addition to the MTA mentioned above, other ATCC and/or regulatory permits may be required for the transfer of this ATCC material. Anyone purchasing ATCC material is ultimately responsible for obtaining the permits. Please click here for information regarding the specific requirements for shipment to your location.
Applications:	transfection host (Nucleofection technology from Lonza Roche FuGENE® Transfection Reagents)
Tumorigenic:	Yes
Antigen Expression:	HLA A1, A9
DNA Profile (STR):	Amelogenin: X CSF1PO: 11 D13S317: 11 D16S539: 11 D5S818: 13 D7S820: 8,11 THO1: 6,7 TPOX: 8,9 vWA: 17
Cytogenetic Analysis:	The line is near-triploid with a modal number of 62 chromosomes. There are nearly 20 marker chromosomes commonly found in each cell; and normal N2, N3, N4, N5, N12, and N15 are not found. No normal Y chromosomes could be detected by Q-band analysis.
Age:	62 years adult
Gender:	male
Ethnicity:	Caucasian
Comments:	The PC-3 was initiated from a bone metastasis of a grade IV prostatic adenocarcinoma from a 62-year-old male Caucasian. [22363] The cells exhibit low acid phosphatase and testosterone-5-alpha reductase activities.
Propagation:	**ATCC complete growth medium:** The base medium for this cell line is ATCC-formulated F-12K Medium, Catalog No. 30-2004. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 10%. **Atmosphere:** air, 95%; carbon dioxide (CO_2), 5% **Temperature:** 37.0°C
Subculturing:	**Protocol:**

1. Remove and discard culture medium.
2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin- 0.53 mM EDTA solution to remove all traces of serum that contains trypsin inhibitor.
3. Add 2.0 to 3.0 ml of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes).
   Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
4. Add 6.0 to 8.0 ml of complete growth medium and aspirate cells by gently pipetting

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Designations:	NCI-H1299
Depositors:	AF Gazdar, JD Minna
Biosafety Level:	1
Shipped:	frozen
Medium & Serum:	See Propagation
Growth Properties:	adherent
Organism:	*Homo sapiens* (human)
Morphology:	epithelial
Source:	**Organ:** lung **Disease:** carcinoma; non-small cell lung cancer **Derived from metastatic site:** lymph node
Cellular Products:	neuromedin B
Permits/Forms:	In addition to the MTA mentioned above, other ATCC and/or regulatory permits may be required for the transfer of this ATCC material. Anyone purchasing ATCC material is ultimately responsible for obtaining the permits. Please click here for information regarding the specific requirements for shipment to your location.
Restrictions:	The line is available with the following restrictions: 1. This cell line was deposited at the ATCC by Dr. A. Gazdar and Dr. J. Minna and is provided for research purposes only. Neither the cell line nor products derived from it may be sold or used for commercial purposes. Nor can the cells be distributed to third parties for purposes of sale, or producing for sale, cells or their products. The cells are provided as service to the research community. They are provided without warranty of merchantability or fitness for a particular purpose or any other warranty, expressed or implied. 2. Any proposed commercial use of the these cells, or their products must first be negotiated with the University of Texas Southwestern Medical Center at Dallas, 5323 Harry Hines Blvd., Dallas, Texas 75235. Telephone (214) 699-8056, FAX (214) 688-7233.
Applications:	transfection host (Nucleofection technology from Lonza Roche FuGENE® Transfection Reagents)
DNA Profile (STR):	Amelogenin:X CSF1PO:12 D13S317:12 D16S539:12,13 D5S818:11 D7S820:10 THO1:6,9.3 TPOX:8 vWA: 16,17,18
Age:	43 years adult
Gender:	male
Ethnicity:	Caucasian
Comments:	The cells have a homozygous partial deletion of the p53 protein, and lack expression of p53 protein. They reported to be able to synthesize the peptide neuromedin B (NMB) at 0.1 pmol/mg protein, but not the gastrin releasing peptide (GRP).
Propagation:	**ATCC complete growth medium:** The base medium for this cell line is ATCC-formulated RPMI-1640 Medium, Catalog No. 30-2001. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 10%. **Temperature:** 37.0°C **Atmosphere:** air, 95%; carbon dioxide (CO2), 5%
Subculturing:	**Protocol:** 1. Remove and discard culture medium. 2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin - 0.53 mM EDTA solution to remove all traces of serum that contains trypsin inhibitor. 3. Add 2.0 to 3.0 ml of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes). Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37C to facilitate dispersal. 4. Add 6.0 to 8.0 ml of complete growth medium and aspirate

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Designations:	HT-29
Depositors:	J Fogh
Biosafety Level:	1
Shipped:	frozen
Medium & Serum:	See Propagation
Growth Properties:	adherent
Organism:	*Homo sapiens* (human)
Morphology:	epithelial

Source:	**Organ:** colon **Disease:** colorectal adenocarcinoma
Cellular Products:	secretory component of IgA; carcinoembryonic antigen (CEA); transforming growth factor beta binding protein; mucin
Permits/Forms:	In addition to the MTA mentioned above, other ATCC and/or regulatory permits may be required for the transfer of this ATCC material. Anyone purchasing ATCC material is ultimately responsible for obtaining the permits. Please click here for information regarding the specific requirements for shipment to your location.
Restrictions:	The cells are distributed for research purposes only. The Memorial Sloan-Kettering Cancer Center releases the line subject to the following: 1.) The cells or their products must not be distributed to third parties. Commercial interests are the exclusive property of Memorial Sloan-Kettering Cancer Center. 2.) Any proposed commercial use of these cells must first be negotiated with The Director, Office of Industrial Affairs, Memorial Sloan-Kettering Cancer Center, 1275 York Avenue, New York, NY 10021; phone (212) 639-6181; FAX (212) 717-3439.
Isolation:	**Isolation date:** 1964
Applications:	transfection host (Nucleofection technology from Lonza Roche FuGENE® Transfection Reagents)
Receptors:	human adrenergic alpha2A [23560] urokinase receptor (u-PAR) vitamin D (moderate expression) urokinase receptor (u-PAR); vitamin D (moderate expression)
Tumorigenic:	Yes
Oncogene:	myc +; ras +; myb +; fos +; sis +; p53 +; abl -; ros -; src -
Antigen Expression:	Blood Type A; Rh+; HLA A1, A3, B12, B17, Cw5
DNA Profile (STR):	Amelogenin: X CSF1PO: 11,12 D13S317: 11,12 D16S539: 11,12 D5S818: 11,12 D7S820: 10 THO1: 6,9 TPOX: 8,9 vWA: 17,19
Cytogenetic Analysis:	modal number = 71; range = 68 to 72. The stemline chromosome number is hypertriploid with the 2S component occurring at 2.4%. Seventeen marker chromosomes are found in most metaphases, generally in single copy per chromosome. The marker designations are: M1p-(=t(3p-;?) with a deleted short arm), t(7q;?), t(10q;?), i(13q), 19q+a; M6, ?t(8q;9q-), ?Xp, M9, 6q+, t(13;?)a, t(13;?)b, 19q+b, M14, M15, 15p+, and Xq-. Chromosome 13 is nullisomic and chromosomes 8 and 14 are generally monosomic. No Y chromosome was detected by QM band analysis.
Isoenzymes:	AK-1, 1 ES-D, 1 G6PD, B GLO-I, 1-2 Me-2, 1 PGM1, 1-2 PGM3, 1-2

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Designations:	C6
Depositors:	G Sato
Biosafety Level:	1
Shipped:	frozen
Medium & Serum:	See Propagation
Growth Properties:	adherent
Organism:	Rattus norvegicus (rat)
Morphology:	fibroblast
Source:	**Organ:** brain **Disease:** glioma **Cell Type:** glial cell;
Cellular Products:	S-100 protein; produce glyceryl phosphate dehydrogenase in response to glucocorticoids; somatotrophin
Permits/Forms:	In addition to the MTA mentioned above, other ATCC and/or regulatory permits may be required for the transfer of this ATCC material. Anyone purchasing ATCC material is ultimately responsible for obtaining the permits. Please click here for information regarding the specific requirements for shipment to your location.
Applications:	transfection host (Nucleofection technology from Lonza Roche FuGENE® Transfection Reagents)
Receptors:	glucocorticoid
Virus Resistance:	poliovirus 3
Cytogenetic Analysis:	Stemline number is diploid. Karyotype is stable within the stemline number and is that of a normal male. Three cells with breaks; one with a secondary constriction, one with a dicentric, one with a rearrangement and four with terminal or centromere associations.
Comments:	The glial cell strain, C6, was cloned from a rat glial tumor induced by N-nitrosomethylurea by Benda et al. after a series of alternate culture and animal passages [PubMed: 4873531]. S-100 production increases ten fold as cells grow from low density to confluency.
Propagation:	**ATCC complete growth medium:** The base medium for this cell line is ATCC-formulated F-12K Medium, Catalog No. 30-2004. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 2.5%; horse serum to a final concentration of 15%. **Atmosphere:** air, 95%; carbon dioxide (CO2), 5% **Temperature:** 37.0°C
Subculturing:	**Protocol:** 1. Remove and discard culture medium. 2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin - 0.53 mM EDTA solution to remove all traces of serum which contains trypsin inhibitor. 3. Add 2.0 to 3.0 ml of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes). Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37C to facilitate dispersal. 4. Add 6.0 to 8.0 ml of complete growth medium and aspirate cells by gently pipetting. 5. Add appropriate aliquots of the cell suspension to new culture vessels. 6. Incubate cultures at 37C. **Subcultivation Ratio:** A subcultivation ratio of 1:2 to 1:3 is recommended **Medium Renewal:** 2 to 3 times per week
Preservation:	**Freeze medium:** culture medium, 95%; DMSO, 5% **Storage temperature:** liquid nitrogen vapor phase
Related Products:	Recommended medium (without the additional supplements or serum described under ATCC Medium):ATCC 30-2004 recommended serum:ATCC 30-2020 recommended serum:ATCC 30-2040 0.25% (w/v) Trypsin - 0.53 mM EDTA in Hank' BSS (w/o Ca++,

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Cell Biology

ATCC® Number:	CRL-1603™ Order this Item
Designations:	N1-S1 Fudr
Depositors:	JE Becker
Biosafety Level:	1
Shipped:	frozen
Medium & Serum:	See Propagation
Growth Properties:	suspension
Organism:	Rattus norvegicus (rat)
Morphology:	
Source:	**Organ:** liver **Strain:** Sprague-Dawley **Disease:** hepatoma; Novikoff hepatoma
Permits/Forms:	In addition to the MTA mentioned above, other ATCC and/or regulatory permits may be required for the transfer of this ATCC material. Anyone purchasing ATCC material is ultimately responsible for obtaining the permits. Please click here for information regarding the specific requirements for shipment to your location.
Gender:	male
Comments:	The cells are resistant to 5-fluoro-2'-deoxyuridine (FUdR) at 0.05 mM and to 5-fluorouracil. This line is derivative of N1-S1 (ATCC CRL-1604).
Propagation:	**ATCC complete growth medium:** The base medium for this cell line is ATCC-formulated Dulbecco's Modified Eagle's Medium, Catalog No. 30-2002. To make the complete growth medium, add the following components to the base medium: • 10% fetal bovine serum • 0.05 mM FUdR • 0.1% pluronic F68
Subculturing:	**Medium Renewal:** Twice per week Cultures can be maintained by addition or replacement of fresh medium. Start cultures at 4 X 10 exp4 viable cells/ml and subculture at 1 X 10 exp6 cells/ml (about 3 days). Cells can be grown in Florence flasks on a rotary shaker at 180 to 190 RPM.
References:	1084: Potter VR, Morse PA Jr.. Pyrimidine metabolism in tissue culture cells derived from rat hepatomas. I. Suspension cell cultures derived from the Novikoff hepatoma. Cancer Res. 25: 499-508, 1965. PubMed: 14297488 26107: Cory JG, et al. Evidence for role of purine nucleoside phosphorylase in sensitivity of Novikoff hepatoma cells to 5-fluorouracil. Adv. Enzyme Regul. 15: 153-166, 1976. PubMed: 197803

Price: $429.00

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VX2 info

-------- Original Message --------
Subject: Re: Containment level request - Cotton rabit papilloma virus
Date: Mon, 11 Jan 2010 13:48:14 -0500
From: Permit-Permis <permitpermis@phac-aspc.gc.ca>
To: Jennifer Stanley <jstanle2@uwo.ca>

Dear Jennifer Stanley

This is a RG1 human pathogen you can work in a CL1. CFIA may have other
requirements.

Regards

Josee DaviesA/Regulatory Technologist/ technologiste en réglementation
Pathogen Regulation Directorate (formerly Office of Laboratory Security) /
Direction de la réglementation des agents pathogènes (anciennement le Bureau de la sécurité des laboratoires)
Public Health Agency of Canada/ Agence de santé publique du Canada
100 ch. Colonnade Rd. AL: 6201A Ottawa, Ontario, Canada K1A 0K9
Tel: (613) 957-1779
Fax: (613)941-0596

------- Original Message --------
Subject: Re: Containment level request - Cotton rabit papilloma virus
Date: Tue, 12 Jan 2010 09:58:52 -0500
From: ImportZoopath <ImportZoopath@inspection.gc.ca>
To: Jennifer Stanley <jstanle2@uwo.ca>

Level 2.

Cinthia Labrie

Office of Biohazard Containment & Safety, CFIA | Bureau du confinement des biorisques et de la sécurité, ACIA
Government of Canada | Gouvernement du Canada
59 Camelot, Ottawa ON K1A0Y9
Phone/Tél.: (613) 221-7068
Fax/ Téléc.: (613) 228-6129
ImportZoopath@inspection.gc.ca

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http://www.inspection.gc.ca/english/sci/bio/bioe.shtml /
Vous trouverez une nouvelle version de la liste de vérification NC2 dans la section Listes de vérification et formulaires de notre site internet; http://www.inspection.gc.ca/francais/sci/bio/biof.shtml

>>> Jennifer Stanley <jstanle2@uwo.ca> 2010-01-12 09:58 >>>
Sorry...which containment level is it?

On 1/12/2010 9:57 AM, ImportZoopath wrote:
> Hi Ms. Stanley,
>
> My database list the Shope / Rabbit papilloma virus as a containment
> level organism affecting rabbit skin.
>
> Have a nice day,
>
> Cinthia Labrie
>
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> confinement des biorisques et de la sécurité, ACIA
> Government of Canada | Gouvernement du Canada
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