

**THE UNIVERSITY OF WESTERN ONTARIO
BIOLOGICAL AGENTS REGISTRY FORM**
Approved Biohazards Subcommittee: July 9, 2010
Biosafety Website: www.uwo.ca/humanresources/biosafety/

This form must be completed by each Principal Investigator holding a grant administered by the University of Western Ontario (UWO) or in charge of a laboratory/facility where the use of Level 1, 2 or 3 biological agents is described in the laboratory or animal work proposed. The form must also be completed if any work is proposed involving animals carrying zoonotic agents infectious to humans or involving plants, fungi, or insects that require Public Health Agency of Canada (PHAC) or Canadian Food Inspection Agency (CFIA) permits.

This form must be updated at least every 3 years or when there are changes to the biological agents being used.

Containment Levels will be established in accordance with Laboratory Biosafety Guidelines, 3rd edition, 2004, Public Health Agency of Canada (PHAC) or Containment Standards for Veterinary Facilities, 1st edition 1996, Canadian Food Inspection Agency (CFIA).

Completed forms are to be returned to Occupational Health and Safety, (OHS), (Support Services Building, Room 4190) for distribution to the Biohazards Subcommittee. For questions regarding this form, please contact the Biosafety Officer at extension 81135 or biosafety@uwo.ca. If there are changes to the information on this form (excluding grant title and funding agencies), contact Occupational Health and Safety for a modification form. See website: www.uwo.ca/humanresources/biosafety/

PRINCIPAL INVESTIGATOR	<u>Dr. Shawn Li</u>
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EMAIL	<u>sli@uwo.ca</u>

Location of experimental work to be carried out: Building(s) **SDRI** Room(s) **108A**

*For work being performed at Institutions affiliated with the University of Western Ontario, the Safety Officer for the Institution where experiments will take place must sign the form prior to its being sent to the University of Western Ontario Biosafety Officer (See Section 15.0, Approvals).

FUNDING AGENCY/AGENCIES: **Canada Cancer Society**

GRANT TITLE(S): **Numb as a Tumor Suppressor**

List all personnel working under Principal Investigators supervision in this location:

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Huadong Liu	hliu223@uwo.ca	May 17, 2007
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Ran Wei	rwei5@uwo.ca	April 30, 2009
Wendy Zhu	wzhu24@uwo.ca	Sept 10, 2009
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Gurpreet Dhani	gdhani2@uwo.ca	Nov 9, 2009

Please explain the biological agents and/or biohazardous substances used and how they will be stored, used and disposed of. Projects without this description will not be reviewed.

1. E. coli DH5 α was from Invitrogen. E. coli BL21 was from Novagen. E. coli DH5 α and BL21 strains are used as competent cell for transformation. They are stored at -80°C freezer. When using for transformation, they will be thawed to 4°C on ice, then take 100 μ l of E. coli to mix with 10 ng plasmid DNA for 20 min on ice, hot shock at 42°C for 90 seconds, stand on ice for 2 min. Add 1 ml of LB medium culture at 37°C for 1 hr, take 200 μ l of mixture on antibiotic LB culture plate at 37°C overnight. The colonies will be used for protein or DNA purification. Waste E. coli, liquids and containers will be treated by bleach to decontaminate. All containers or tubes will be washed and autoclaved. All autoclave treated stuff will be disposed as regular garbage.
2. Cell lines HEK293, P19 and MDCK2 were from ATCC. HEK293, P19 and MDCK2 are used as experimental models. They are stored in liquid nitrogen. When using for experiments, they will be thawed to 37°C in water bath quickly, then put them into culture plate with 10% FBS completed medium to grow up at 37°C for 2 days or so. All operation will be preceded in class II biological safety cabinet in containment level 2 culture room. Then aspirate culture medium from plates. Briefly rinse the cell layer with PBS to remove all traces of serum. Add 2.0 to 3.0 ml of Trypsin-EDTA solution to plate and observe cells under an inverted microscope until cell layer is dispersed. Add 6.0 to 8.0 ml of complete growth medium and aspirate cells by gently pipetting. Add appropriate aliquots of the cell suspension to new culture vessels. Incubate cultures at 37°C for 1-2 days. Cells will be treated by different reagents and methods depending on each research purpose. Waste culture medium and cells will be collected into bleach containing waste bottle to decontaminate. All used experiment materials (cultural plates, pipets, tips, etc.) will be collected and autoclaved. All autoclave treated stuff will be disposed as regular garbage.

Please include a one page research summary or teaching protocol.

Numb is a protein originally identified for its role in animal development. However, it was recently found to also act as a tumor suppressor which regulates cell growth, cell-cell adhesion, and cell migration. In breast and prostate cancer cells, mutation of the *numb* gene or loss of the protein correlates with poor prognosis.

Our lab has identified a new function for Numb in regulating cell-cell adhesion and epithelial-to-mesenchymal transition (EMT) - a critical step in cancer progression in which cancer cells lose adhesiveness and orientation and migrate away from the original site of lesion to invade surrounding tissues and remote sites.

Work from our lab and others make us believe that Numb is a multi-faceted protein that plays an important role in the onset and progression of cancers by regulating cell growth, survival, orientation and cell-cell adhesion.

As such, it represents a druggable target for cancer treatment. In this current proposal, we wish to characterize these diverse functions of Numb using a combination of biochemical, biophysical and cellular methods.

Specifically, we will examine a dynamic interplay between Numb and tyrosine kinases, enzymes in the body that modify other proteins by adding a phosphate group. Excessive tyrosine kinase activity is often associated with cancer. Moreover, we will determine the mechanism behind a role for Numb in regulating cell-cell adhesion, cell orientation and movement. And finally, we will characterize an interaction between Numb and Alk, a kinase that is aberrantly activated in a wide spectrum of different cancers, and elucidate how this interaction affects cell growth and death.

It is anticipated that these studies will provide novel insights into Numb function in cancer and aid in efforts to develop cancer therapies that specifically target Numb or its binding partners.

Numb interacting protein (NIP) is another protein of interest in our lab. NIP is involved in the production of reactive oxygen species (ROS), and its overexpression in stem cells enhances neuronal and cardiac differentiation. We have linked some of this phenotype to enhanced ROS production, but both our protein's cellular localization and its amino acid sequence suggest other roles.

We believe this protein may work as a co-activator for a variety of hormones and transcription factors. We are interested in determining whether overexpression of our protein of interest will enhance luciferase production in both the pGL3-NFAT and pGL3-RARE reporter vectors. We have selected these particular reporters since NFAT has previously been shown to be stimulated by ROS. We have selected the retinoic acid response element (RARE) because our protein of interest stimulates differentiation in the absence of retinoic acid induction, and we believe it may be acting as a co-activator for the retinoic acid receptor. We are planning to transfect 293 cells with these reporter vectors and our protein of interest in order to start testing our hypotheses. We hope that our work will identify new roles for this protein.

ADAMTS12 is a protease that cleaves proteins in response to cell signaling thereby changing the cells response to specific signals.

We recently see co-purified ADAMTS12 with a transcription factor TLE4. TLE4 can act as an activator of transcription and when cleaved, by an unknown protease; a transcriptional repressor. Since ADAMTS12 is a protease and can bind to and interact with TLE4 we speculate that ADAMTS12 is in fact the protein responsible for cleavage of TLE4.

We will test this hypothesis through standard Li laboratory protocols involving transient transfection of HEK293 cells with ADAMTS12 or a enzymatically inactive form of ADAMTS12. We will then compare the size of TLE4 by standard Li laboratory immunoblot techniques.

Previous studies have shown that TLE4 not only changes size but also has different co-factors in the cleaved form. We will evaluate the effect of ADAMTS12 through both assaying of TLE4 size by immunoblot but also through determining the proteins associated with TLE4 and the ratio of known exclusive cleaved interactors such as KDM5b.

1.0 Microorganisms

1.1 Does your work involve the use of biological agents? YES
 (non-pathogenic and pathogenic biological agents including but not limited to bacteria and other microorganisms, viruses, prions, parasites or pathogens of plant or animal origin)? If no, please proceed to Section 2.0

Do you use microorganisms that require a permit from the CFIA? NO

If YES, please give the name of the species. _____

What is the origin of the microorganism(s)? _____

Please describe the risk (if any) of escape and how this will be mitigated:

Please attach the CFIA permit.

Please describe any CFIA permit conditions:

1.2 Please complete the table below:

Name of Biological agent(s)*	Is it known to be a human pathogen? YES/NO	Is it known to be an animal pathogen? YES/NO	Is it known to be a zoonotic agent? YES/NO	Maximum quantity to be cultured at one time? (in Litres)	Source/Supplier	PHAC or CFIA Containment Level
E. coli DH5α	<input type="radio"/> No	<input type="radio"/> No	<input type="radio"/> No			<input type="radio"/> 1
E. coli BL21	<input type="radio"/> No	<input type="radio"/> No	<input type="radio"/> No			<input type="radio"/> 1
	<input type="radio"/> Yes <input type="radio"/> No	<input type="radio"/> Yes <input type="radio"/> No	<input type="radio"/> Yes <input type="radio"/> No			<input type="radio"/> 1 <input type="radio"/> 2 <input type="radio"/> 2+ <input type="radio"/> 3
	<input type="radio"/> Yes <input type="radio"/> No	<input type="radio"/> Yes <input type="radio"/> No	<input type="radio"/> Yes <input type="radio"/> No			<input type="radio"/> 1 <input type="radio"/> 2 <input type="radio"/> 2+ <input type="radio"/> 3

*Please attach a Material Safety Data Sheet or equivalent from the supplier.

2.0 Cell Culture

2.1 Does your work involve the use of cell cultures? YES
 If no, please proceed to Section 3.0

2.2 Please indicate the type of primary cells (i.e. derived from fresh tissue) that will be grown in culture:

Cell Type	Is this cell type used in your work?	Source of Primary Cell Culture Tissue	AUS Protocol Number
Human	<input type="radio"/> No		Not applicable
Rodent	<input type="radio"/> No		
Non-human primate	<input type="radio"/> No		
Other (specify)	<input type="radio"/> No		

Specific effects

Carcinogenic effects	No information available
Mutagenic effects	No information available
Reproductive toxicity	No information available
Sensitization	No information available

Target Organ Effects Eyes. Skin.

4. FIRST AID MEASURES

Skin contact	Wash off immediately with plenty of water
Eye contact	Rinse thoroughly with plenty of water, also under the eyelids.
Ingestion	Never give anything by mouth to an unconscious person
Inhalation	Move to fresh air
Notes to physician	Treat symptomatically

5. FIRE-FIGHTING MEASURES

Suitable extinguishing media	Dry chemical
Special protective equipment for firefighters	Wear self-contained breathing apparatus and protective suit

6. ACCIDENTAL RELEASE MEASURES

Personal precautions	Use personal protective equipment
Methods for cleaning up	Soak up with inert absorbent material

7. HANDLING AND STORAGE

Handling	Avoid contact with skin and eyes.
Storage	Keep in properly labelled containers

8. EXPOSURE CONTROLS / PERSONAL PROTECTION

Occupational exposure controls

Exposure limits

Chemical Name	OSHA PEL (TWA)	OSHA PEL (Ceiling)	ACGIH OEL (TWA)	ACGIH OEL (STEL)
dimethylsulfoxide	-	-	-	-

Engineering measures Ensure adequate ventilation, especially in confined areas

Personal protective equipment

Respiratory protection	In case of insufficient ventilation wear suitable respiratory equipment
Hand protection	Protective gloves
Eye protection	Safety glasses with side-shields
Skin and body protection	Lightweight protective clothing
Hygiene measures	Handle in accordance with good industrial hygiene and safety practice
Environmental exposure controls	Prevent product from entering drains

9. PHYSICAL AND CHEMICAL PROPERTIES

General Information

Form Liquid

Important Health Safety and Environmental Information

Boiling point/range	°C 189	°F No data available
Melting point/range	°C 18.4	°F No data available
Flash point	°C 94	°F No data available
Autoignition temperature	°C No data available	°F No data available
Oxidizing properties	No information available	
Water solubility	soluble	

10. STABILITY AND REACTIVITY

Stability	Stable under normal conditions.
Materials to avoid	No information available
Hazardous decomposition products	No information available
Polymerization	Hazardous polymerisation does not occur

11. TOXICOLOGICAL INFORMATION

Acute toxicity

Chemical Name	LD50 (oral, rat/mouse)	LD50 (dermal, rat/rabbit)	LC50 (inhalation, rat/mouse)
dimethylsulfoxide	14500 mg/kg (Rat)	No data available	No data available

Principle Routes of Exposure/ Potential Health effects

Eyes	Irritating to eyes.
Skin	Irritating to skin. Components of the product may be absorbed into the body through the skin.
Inhalation	May cause irritation of respiratory tract.
Ingestion	May be harmful if swallowed.

Specific effects

Carcinogenic effects	No information available
Mutagenic effects	No information available
Reproductive toxicity	No information available
Sensitization	No information available

Target Organ Effects Eyes. Skin.

12. ECOLOGICAL INFORMATION

Ecotoxicity effects	No information available.
Mobility	No information available.
Biodegradation	Inherently biodegradable.
Bioaccumulation	Does not bioaccumulate.

13. DISPOSAL CONSIDERATIONS

Dispose of in accordance with local regulations

14. TRANSPORT INFORMATION

IATA

Proper shipping name	Not classified as dangerous in the meaning of transport regulations
Hazard Class	No information available
Subsidiary Class	No information available
Packing group	No information available
UN-No	No information available

Proper shipping name Not classified as dangerous within the meaning of transport regulations

15. REGULATORY INFORMATION

International Inventories

Chemical Name	TSCA	PICCS	ENCS	DSL	NDSL	AICS
dimethylsulfoxide	Listed	Listed	Listed	Listed	-	Listed

U.S. Federal Regulations

SARA 313
Not regulated

Clean Air Act, Section 112 Hazardous Air Pollutants (HAPs) (see 40 CFR 61)
This product contains the following HAPs:

U.S. State Regulations

Chemical Name	Massachusetts - RTK	New Jersey - RTK	Pennsylvania - RTK	Illinois - RTK	Rhode Island - RTK
dimethylsulfoxide	-	-	-	-	-

California Proposition 65

This product contains the following Proposition 65 chemicals:

WHMIS hazard class:
D2B Toxic materials

This product has been classified according to the hazard criteria of the CPR and the MSDS contains all of the information required by the CPR

16. OTHER INFORMATION

This material is sold for research and development purposes only. It is not for any human or animal therapeutic or clinical diagnostic use. It is not intended for food, drug, household, agricultural, or cosmetic use. An individual technically qualified to handle potentially hazardous chemicals must supervise the use of this material.

The above information was acquired by diligent search and/or investigation and the recommendations are based on prudent application of professional judgment. The information shall not be taken as being all inclusive and is to be used only as a guide. All materials and mixtures may be present unknown hazards and should be used with caution. Since Invitrogen Corporation cannot control the actual methods, volumes, or conditions of use, the Company shall not be held liable for any damages or losses resulting from the handling or from contact with the product as described herein. THE INFORMATION IN THIS MSDS DOES NOT CONSTITUTE A WARRANTY, EXPRESS OR IMPLIED, INCLUDING ANY IMPLIED WARRANTY OF MERCHANTABILITY OR FITNESS FOR ANY PARTICULAR PURPOSE.



Order Number

Customer Number

1. Product and Company Identification

Supplier: Manufactured by EMD Biosciences, Inc.
 441 Charming Drive
 Madison, WI 53719
 (608)238-6110
 (800)207-0144
 FAX: (608)238-1388

P.O. Box 12087
 La Jolla, CA 92039-2087
 (858)450-5558
 (800)854-3417
 FAX: (858)453-3552

Catalog #: 69449
 In Case of
 Emergency:

Product name: BL21 Competent Cells

2. Composition and Information on Ingredients

Ingredient Name	CAS No.	Product No.	EC Symbol	R-Phrases
Calcium Chloride	10043-52-4	RC0030	Xi	R36

Note: See section 8 for occupational exposure limits and section 11 for LC50/LD50 information.

3. Hazards Identification

Primary Hazards and Critical Effects	: RC0030	CAUTION! BE HARMFUL IF SWALLOWED. MAY CAUSE EYE IRRITATION. Avoid contact with eyes. Do not ingest. Wash thoroughly after handling.	MAY
Physical/Chemical hazards	:	Not applicable.	
Human Health Hazards	: RC0030	Irritating to eyes.	
Environmental Hazards	:	Not applicable.	

4. First Aid Measures

Inhalation	: RC0030	If inhaled, remove to fresh air. If not breathing, give artificial respiration. If breathing is difficult, give oxygen. Get medical attention.
Ingestion	: RC0030	Do NOT induce vomiting unless directed to do so by medical personnel. Never give anything by mouth to an unconscious person. If large quantities of this material are swallowed, call a physician immediately.
Skin Contact	: RC0030	In case of contact, immediately flush skin with plenty of water. Remove contaminated clothing and shoes. Wash clothing before reuse. Thoroughly clean shoes before reuse. Get medical attention.
Eye Contact	: RC0030	In case of contact, immediately flush eyes with plenty of water for at least 15 minutes. Get medical attention.
Notes to Medical Doctor	:	Not available.

5. Fire-Fighting Measures

Extinguishing Media	:	Use foam or all purpose dry chemicals to extinguish.
Fire-Fighting Procedures	:	Fire fighters should wear positive pressure self-contained breathing apparatus (SCBA) and full turnout gear.
Explosion Hazards	:	Not applicable.
Hazardous Decomposition Products	:	Not available.

6. Accidental Release Measures

Personal Precautions	:	Immediately contact emergency personnel. Keep unnecessary personnel away. Use suitable protective equipment (Section 8). Follow all fire fighting procedures (Section 5).
Environmental Precautions and Clean-up Methods	:	If emergency personnel are unavailable vacuum or carefully scoop up spilled materials and place in an appropriate container for disposal. Avoid creating dusty conditions and prevent wind dispersal. Minimize contact of spilled material with soils to prevent runoff to surface waterways.

Note: See section 1 for emergency contact information and section 13 for waste disposal.

7. Handling and Storage

Handling	:	RC0030	Avoid contact with eyes. Do not ingest. Wash thoroughly after handling.
Storage	:		Keep container tightly closed. Keep container in a cool, well-ventilated area.
Packaging Materials	:		Use original container.

8. Exposure Controls and Personal Protection

Occupational Exposure Limit

Ingredient Name	Occupational Exposure Limits
RC0030	Not available.

Engineering Controls	:	RC0030	No special containment is required. Local exhaust ventilation should be provided.
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Personal Protective Equipment

Respiratory System	:	RC0030	Use an approved, properly fitted, HEPA filter cartridge respirator, or a respirator of greater protection if there is the potential to exceed the exposure limit(s).
Skin and Body	:	RC0030	Work uniform or laboratory coat.
Hands	:	RC0030	Use chemical resistant, impervious gloves. Additional body garments should be used based upon the task being performed (e.g., sleevelets, apron, gauntlets, disposable suits). Appropriate techniques should be used to remove potentially contaminated clothing.
Eye	:	RC0030	Safety glasses. Goggles, face shield, or other full-face protection if potential exists for direct exposure to dust.

9. Physical and Chemical Properties

Key Components

0.4 ml Cells
 69318: 1 x 10 ul Test Plasmid
 69319: 2 x 2 ml SOC Medium
 69449: 2 x 0.2 ml BL21 Competent Cells, containing Calcium Chloride (RC0030)

1.0 ml Cells
 69318: 1 x 10 ul Test Plasmid
 69319: 4 x 2 ml SOC Medium
 69449: 5 x 0.2 ml BL21 Competent Cells, containing Calcium Chloride (RC0030)

Flash Point

Not available.

10. Stability and Reactivity

Stability	: RC0030	The product is stable.
Conditions and Materials to Avoid	: RC0030	Reactive with acids.
Hazardous Decomposition Products	:	Not available.

11. Toxicological Information

Toxicity Data

<u>Ingredient Name</u>	<u>LC50</u>	<u>LD50</u>	<u>Route</u>	<u>Species</u>
RC0030	LD50	1000 mg/kg	Oral	Rat
	LD50	1940 mg/kg	Oral	Mouse

Routes of Entry : Eye contact.

Acute Effects

Inhalation	:	Not available.
Ingestion	: RC0030	Harmful if swallowed.
Skin Contact	:	Not available.
Eye Contact	: RC0030	Moderately irritating to the eyes.

Chronic Effects

Adverse Effects	:	Not available.
Target Organs	:	Not available.
Carcinogenic Effects	:	Not available.
Mutagenic Effects	:	Not available.
Developmental and Teratogenic Effects	:	Not available.
Reproductive Effects	:	Not available.

Other Information : RC0030 Repeated or prolonged exposure is not known to aggravate medical condition.

12. Ecological Information

Toxicity Data

<u>Ingredient Name</u>	<u>Species</u>	<u>Period</u>	<u>Result</u>
RC0030	Not available.	Not available.	Not available.

13. Disposal Consideration

Waste Handling and Disposal : Waste must be disposed of in accordance with federal, state and local environmental control regulations.

14. Transport Information

Air

DOT Hazard Class : Not controlled under IATA.

Packing Group

15. Regulatory Information

EU Regulations

Hazard Symbols	: Xi
Risk Phrases	: R36- Irritating to eyes.
Safety Phrases	: S24- Avoid contact with skin.

US Regulations

Hazard Classification	: Not controlled under the HCS (United States).
LEL	: Not available.
UEL	: Not available.

Canadian Regulations

WHMIS : Not controlled under WHMIS (Canada).
CEPA : No products were found.
Provincial : No products were found.

16. Other Information

Validated by jew on 8/28/2003.

Version : 1.0

Date of Printing : 8/28/2003.

Notice to Reader

To the best of our knowledge, the information contained herein is accurate. However, neither the above named supplier nor any of its subsidiaries assumes any liability whatsoever for the accuracy or completeness of the information contained herein.

*Final determination of suitability of any material is the sole responsibility of the user. All materials may present unknown hazards and should be used with caution. Although certain hazards are described herein, we cannot guarantee that these are the only hazards that exist. **This product is intended for research use only.***

Cell Line			
ATCC® Number	CRL-1573™	Order this Item	Price \$256.00
Designation	223 (HEp-203)		
Depositor	F. C. Graham		
Priority Date	2/20/64 (9/24/64 N. A. PATENT NO. 3,142,805)		
Shipped	in ice		
Medium & Special	see Propagator		
Growth Parameters	adherent		
Organism	Human Epithelial Cells		
Morphology	Epithelial		
Section	Organism: Eukaryote, Animal Cell Type: Transformed, with genome DNA		
Remarks	In addition to the ATCC cell line accession number, ATCC highly recommends that you may also include the number of the ATCC material. Anyone purchasing ATCC material is also asked to subscribe for details of the primary Research Catalog. For information regarding the subscription forms, for example, to your lab, see:		
References	These cells are a derivative of the research preparation, 223 cells, that are similar to the original but are not directly attributable to that particular culture, using 39768.		
Additional	Proteolysin: ATCC 300006 Isotype: ATCC 300006 (Epithelial, Fibroblast) Viral Reactivity: (R576)		
Antibodies	Viral Reactivity: None		
Transcript	None		
Gen. Probe	ATCC Probe: 3776 CSF1PO: 11-12 E: 353, 7, 12-14 P: 45539, 8-17 D5Sb: 3, 4, 5 D7Sb19: 11-11 T: 21, 2, 3, 5 TPOx: 11 VWA: 16-19		
Chromosomal	This is a hypodiploid human cell line. The modal chromosome number was 64, occurring in 35% of cells. The rate of cells with higher ploidy was 4.2%. The marker 18q115 (p62-q15), der(19)t(11;19)(q12;q13), der(12)t(6;12)(p12;p12), and four other marker chromosomes were common to near cells. Five other markers occurred in some cells only. The markers der(1) and 48 for Xq11 were often paired. There were four copies of N17 and N22. Notably, in addition to three copies of X chromosomes, there were paired Xq11 and a single Xq11 in most cells.		
Age	Adult		
Comments	Although an earlier report suggested that the cells contained Adenovirus 3 DNA from both the right and left ends of the viral genome (RF22764), it is now clear that only left end sequences are present. (39768) The line is excellent for studying human adenovirus. The cells express an unusual cell surface receptor for proteinase composed of the protease beta-1 subunit and the vascular cell adhesion molecule-1 (39768). The Ad5 genome was cloned and sequenced, and it was determined that a linear sequence from base 1 to 4264 is integrated into chromosome 17 (193142). (39768)		
Propagator	ATCC complete growth medium: The base medium for this cell line is ATCC-generated Eagle's Minimum Essential Medium, Catalog No. 30-2009. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 10%. Atmosphere: air, 95% carbon dioxide (CO ₂) 5%. Temperature: 37°C The cell line does not adhere to the substrate when left at room temperature for any length of time; therefore, new cultures may be recovered with the cells detached. The cells will reattach to the flask over a period of several days in culture at 37°C.		
Subculture	Protocol: 1. Remove and discard culture medium. 2. Briefly rinse the cell layer with 0.25% (v/v) Trypsin, 0.03 mM EDTA solution to remove all traces of serum that contains trypsin inhibitor. 3. Add 2.0 to 3.0 mL of Trypsin-EDTA solution to flask and observe cell under an inverted microscope until cell layer is dispersed (usually within 2 to 3 minutes). Note: To avoid clumping of cell aggregates, the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal. 4. Add 8.0 mL of ATCC complete growth medium and transfer cells by gentle mixing. 5. Add appropriate aliquots of the cell suspension to new culture vessels. An aliquot of 2 x 10 ⁶ to 3 x 10 ⁶ viable cells/ml is recommended. 6. Inoculate cultures at 37°C. Subculture when cell concentration is between 0.5 x 10 ⁶ and 7 x 10 ⁶ cells/ml.		
Freeze/Thaw	Subcultivation Ratio: 1:10 to 1:20 weekly Medium Renewal: Every 1 to 2 days Freeze medium: Complete growth medium supplemented with 10% (v/v) DMSO Storage temperature: liquid nitrogen - deep phase		

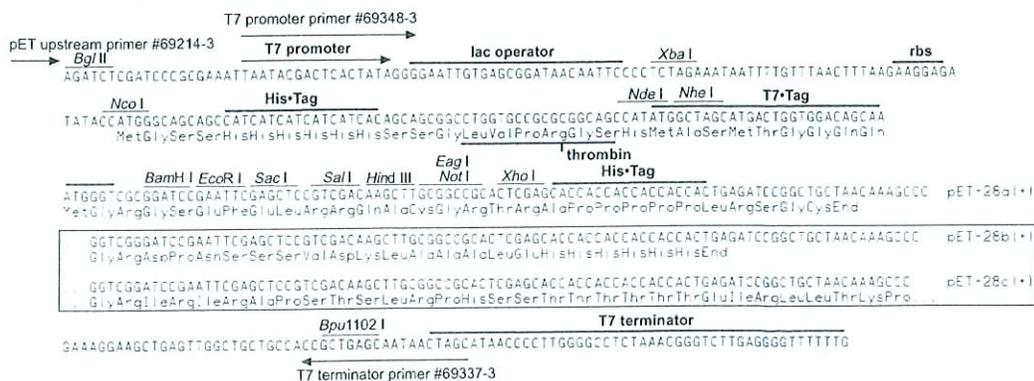
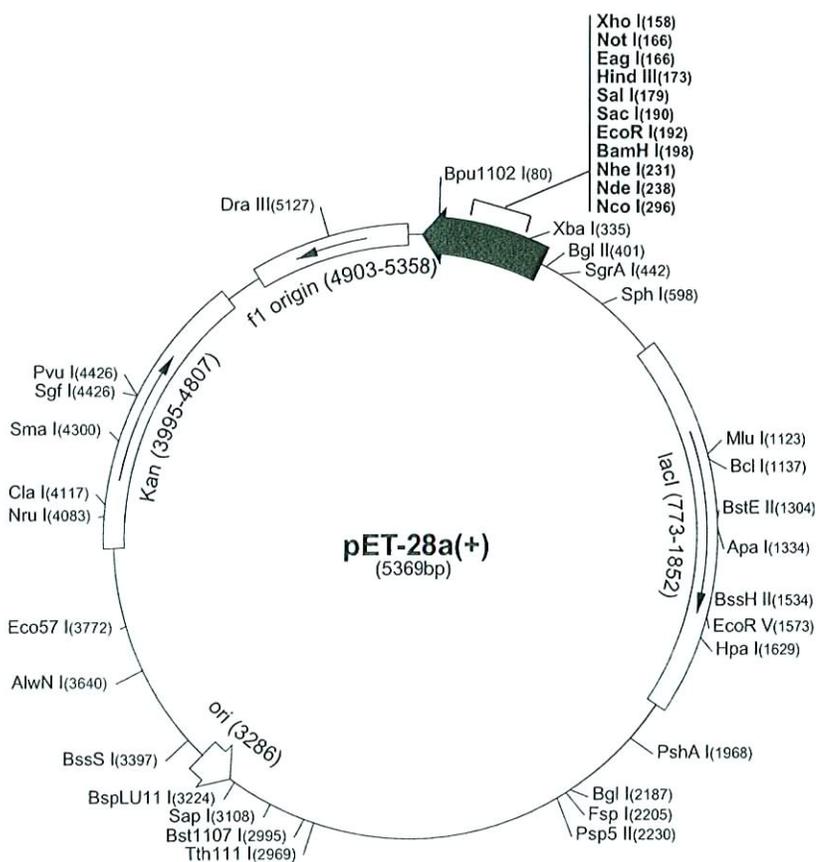
Cell Culture	ATCC® Number	CRL-1825™	Order this Item	Price	\$275.00
Designations	P18				
Depositor	HW McBurney				
Availability Level	1				
Shipped	Frozen				
Medium & Serum	See Propagator				
Growth Parameters	Adherent				
Organism	<i>Melanocytus immutabilis</i>				
Morphology	Epithelial				
Source	<p>Strain Origin: Organ: embryo Disease: melanocytoma, embryonic carcinoma</p>				
Reference(s)	<p>In addition to the ATCC publications above, other ATCC and/or proprietary publications may be required for the transfer of this ATCC material. Anyone purchasing ATCC material is automatically assumed not for obtaining the patent. Please refer to the information regarding the specific requirements for submitting your request.</p>				
Associated Organisms/Associated Cells	<p>Transferrin Receptor (TfR) gene, human, type 1 from cultured cells C-40 (X) - C-40 (X) (ATCC) C-40</p>				
Comments	<p>The P18 line was derived from an embryonic carcinoma induced in a C3H/He mouse (C1779). The number to derive at high efficiency medium sustains 0.1 ml/24 hr suspension (2250). The cells are diploid. The cell line is induced to differentiate into neural and epithelial cells in the presence of 10% FCS in fresh medium (20492). In the presence of 10% to 100% dimethyl sulfoxide (DMSO), the cells differentiate into cardiac and skeletal muscle-like elements, but do not form neuroepithelial cells (2813). In the presence of both DMSO and retinoic acid, the cells differentiate as in the presence of retinoic acid alone (22613).</p>				
Propagator	<p>ATCC complete growth medium: The base medium for this cell line is Alpha Minimum Essential Medium with nicotinamides and deoxyribonucleosides. To make the complete growth medium, add the following components to the base medium: bovine calf serum to a final concentration of 7.5%, fetal bovine serum to a final concentration of 2.5%. Temperature: 37 °C Atmosphere: air, 95% carbon dioxide (CO2), 5%</p>				
Subculturing	<p>Protocol: Do not allow the cells to become confluent.</p> <ol style="list-style-type: none"> 1. Remove and discard culture medium. 2. Enzymatically digest cells with 0.25% (w/v) Trypsin-0.53 mM EDTA solution to remove all traces of serum which contains trypsin inhibitor. 3. Add 2.0 to 3.0 ml of Trypsin-EDTA solution to flask and observe cells until an inverted microcap with a layer of dispersed (suspense) wash to 15 minutes. (Note: To avoid clumping, do not agitate the cells by shaking or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be treated at 37 °C to facilitate dispersal.) 4. Add 0.1 to 2.0 ml of complete growth medium and aspirate cells by gently aspirating. 5. Add appropriate amounts of the cell suspension to new culture vessels. 6. Incubate cultures at 37 °C. 				
Recovery	<p>Subcultivation Ratio: A 1:2 subcultivation rate of 1:10 every 2 to 3 days is recommended. Medium Renewal: Add fresh medium at least every 48 hours. Freeze medium: Complete growth medium (5% DMSO), 1% Storage temperature: liquid nitrogen vapor phase</p>				

Cell Name		Price	\$355.00
ATCC# Number	CRL 2936™ Order this Item		
Designations	MDCK-2		
Depositor	T. Peit, B. Gehrke and B. Edward-Aymar, ATCC		
Registry Code	-		
Strain	triple		
Media & Setup	Sem-Propagated		
Growth Conditions	adherent		
Organism	<i>Canis familiaris</i>		
Reference	C. Mansueti		
Source	Organism <i>Canis familiaris</i> Disease none		
Recovery	In addition to the ATCC standard protocol, other ATCC products may require special handling. For the transfer of this ATCC material, anyone purchasing ATCC material is ultimately responsible for obtaining the proper license agreements. For information regarding the special requirements for shipment to your facility.		
Isolation	Isolated from skin (1977)		
Virus Susceptible	Influenza A virus Influenza B virus		
Antigen Expression	E-cadherin (cell-cell adhesion molecule), expressed Zona Occludens (ZO-1 tight junction protein), expressed The Blast-specific protein (BP), not expressed cadherins (CD) 4, 5, 6, 8, 10, 13, 15, 18; expressed K18 (keratins) alpha 2 & 5 (epithelial keratins) alpha 2 & 6 (epithelial keratins), expressed		
Cytogenetic Analysis	Hybridized canine cell line with a modal chromosome number of 61 with low polyploidy ratio. Several unidentified marker chromosomes were present in most of the cells examined.		
Age	adult		
Comments	Cell line was derived by the long (primary) donor, the parent, canine MDCK (CCL 34). The cell line is susceptible to a wide range of influenza virus and is suitable to culture both of <i>C. pasteurianus</i> .		
Propagator	ATCC complete growth medium: The base medium for this cell line is ATCC-formulated Eagle's Minimum Essential Medium, Catalog No. 30-2005. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 10%.		
Subculture	Protocol: Volumes used in this protocol are for 75 cm ² flask, proportionally reduce or increase amount of dissociation medium for culture vessels of other sizes. <ol style="list-style-type: none"> 1. Remove into clean culture medium. 2. Briefly rinse the cell layer with DMEM (w/o FBS, Dulbecco's phosphate-buffered saline (DPBS) or 0.25% trypsin + 0.53 mM EDTA solution to remove all traces of serum which neutralizes trypsin inhibitors. 3. Add 1.0 ml of 0.25% trypsin-EDTA solution to flask and remove cells under an inverted microscope once cell layer is disrupted (usually within 5 to 15 minutes). Note: To avoid clumping do not agitate the cells by lifting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be treated by TCC to facilitate dispersion. 4. Add 6.0 to 8.0 ml of complete growth medium and aspirate cells by gently pipetting. 5. Add appropriate aliquots of the cell suspension to new culture vessels. An inoculum of 1 X 10⁴ to 2 X 10⁴ viable cells/cm² cells is recommended. 6. Incubate cultures at 37°C. Subculture when the cell concentration is between 1 X 10⁴ and 1 X 10⁶ cells/cm². Subcultivation ratio: A subcultivation ratio of 1:2 to 1:8 is recommended. Medium renewal: Every 3 to 7 days.		
Preservation	Freeze medium: Complete growth medium 80 to 100% v/v liquid nitrogen vapor phase		
Duration Time	approximately 30 hours		

pET-28a(+) sequence landmarks

T7 promoter	370-386
T7 transcription start	369
His•Tag coding sequence	270-287
T7•Tag coding sequence	207-239
Multiple cloning sites (<i>Bam</i> H I - <i>Xho</i> I)	158-203
His•Tag coding sequence	140-157
T7 terminator	26-72
<i>lac</i> I coding sequence	773-1852
pBR322 origin	3286
Kan coding sequence	3995-4807
f1 origin	4903-5358

The maps for pET-28b(+) and pET-28c(+) are the same as pET-28a(+) (shown) with the following exceptions: pET-28b(+) is a 5368bp plasmid; subtract 1bp from each site beyond *Bam*H I at 198. pET-28c(+) is a 5367bp plasmid; subtract 2bp from each site beyond *Bam*H I at 198.



pGEX-2T (27-4801-01)

Thrombin
Leu Val Pro Arg⁺Gly Ser¹Pro Gly Ile His Arg Asp
CTG GTT CCG CGT GGA TCC CCG GGA ATT CAT CCG GAC TGA CTG ACG
BamH I Sma I EcoR I Stop codons

pGEX-2TK (27-4587-01)

Thrombin Kinase
Leu Val Pro Arg⁺Gly Ser¹Arg Arg Ala Ser Val¹
CTG GTT CCG CGT GGA TCT CCG GCA TCT GTT GGA TCC CCG GGA ATT CAT CCG GAC TGA
BamH I Sma I EcoR I Stop codons

pGEX-4T-1 (27-4580-01)

Thrombin
Leu Val Pro Arg⁺Gly Ser¹Pro Glu Phe Pro Gly Arg Leu Glu Arg Pro His Arg Asp
CTG GTT CCG CGT GGA TCC CCG GAA TTC CCG GGT CGA CTC GAG CCG CCG CAT CCG GAC TGA
BamH I EcoR I Sma I Sal I Xho I Not I Stop codons

pGEX-4T-2 (27-4581-01)

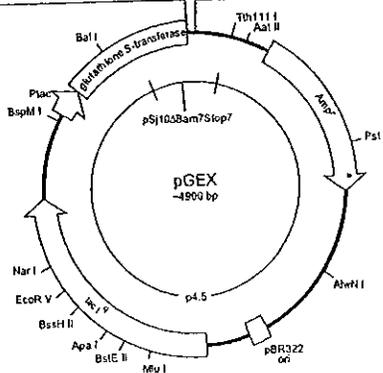
Thrombin
Leu Val Pro Arg⁺Gly Ser¹Pro Gly Ile Pro Gly Ser Thr Arg Ala Ala Ala Ser
CTG GTT CCG CGT GGA TCC CCA GGA ATT CCG GGG TCG ACT CGA GCG GCC GCA TCG TGA
BamH I EcoR I Sma I Sal I Xho I Not I Stop codon

pGEX-4T-3 (27-4583-01)

Thrombin
Leu Val Pro Arg⁺Gly Ser¹Pro Asn Ser Arg Val Asp Ser Ser Gly Arg Ile Val Thr Asp
CTG GTT CCG CGT GGA TCC CCG AAT TCC CCG GTC GAC TCG AGC GGC CGC ATC GTC ACT GAC TGA
BamH I EcoR I Sma I Sal I Xho I Not I Stop codons

pGEX-6P-3 (27-4599-01)

PreScissionTM Protease
Leu Glu Val Leu Phe Gln⁺Gly Pro¹Leu Gly Ser Pro Asn Ser Arg Val Asp Ser Ser Gly Arg
CTG GAA GTT CTG TTC CAG GGG CCC CTG GGA TCC CCG AAT TCC CCG GTC GAC TCG AGC GGC CGC
BamH I EcoR I Sma I Sal I Xho I Not I



2.3 Please indicate the type of established cells that will be grown in culture in:

Cell Type	Is this cell type used in your work?	Specific cell line(s)*	Supplier / Source
Human	<input type="radio"/> Yes	HEK293	ATCC
Rodent	<input type="radio"/> No		
Non-human primate	<input type="radio"/> No		
Other (specify)	<input type="radio"/> Yes	P19, MDCK2	ATCC

*Please attach a Material Safety Data Sheet or equivalent from the supplier. (For more information, see www.atcc.org)

2.4 For above named cell types(s) indicate PHAC or CFIA containment level required 2

3.0 Use of Human Source Materials

3.1 Does your work involve the use of human source materials? NO
If no, please proceed to Section 4.0

3.2 Indicate in the table below the Human Source Material to be used.

Human Source Material	Source/Supplier /Company Name	Is Human Source Material Infected With An Infectious Agent? YES/NO	Name of Infectious Agent (If applicable)	PHAC or CFIA Containment Level (Select one)
Human Blood (whole) or other Body Fluid		<input type="radio"/> Yes <input type="radio"/> Unknown		<input type="radio"/> 1 <input type="radio"/> 2 <input type="radio"/> 2+ <input type="radio"/> 3
Human Blood (fraction) or other Body Fluid		<input type="radio"/> Yes <input type="radio"/> Unknown		<input type="radio"/> 1 <input type="radio"/> 2 <input type="radio"/> 2+ <input type="radio"/> 3
Human Organs or Tissues (unpreserved)		<input type="radio"/> Yes <input type="radio"/> Unknown		<input type="radio"/> 1 <input type="radio"/> 2 <input type="radio"/> 2+ <input type="radio"/> 3
Human Organs or Tissues (preserved)		Not Applicable		Not Applicable

4.0 Genetically Modified Organisms and Cell lines

4.1 Will genetic modifications be made to the microorganisms, biological agents, or cells described in Sections 1.0 and 2.0? YES If no, please proceed to Section 5.0

4.2 Will genetic modification(s) involving plasmids be done? YES

Bacteria Used for Cloning *	Plasmid(s) **	Source of Plasmid	Gene Transfected	Describe the change that results from transformation or transfection
E. coli DH5α	pET-28a	Novagen	No	Transformation to get His-tag fusion protein or GST-fusion protein
E. coli BL21	pGEX-4T3	GE Healthcare	No	

* Please attach a Material Data Sheet or equivalent if available.

** Please attach a plasmid map.

4.3 Will genetic modification(s) involving viral vectors be made? YES NO

Virus Used for Vector Construction	Vector(s) *	Source of Vector	Gene(s) Transduced	Describe the change that results from transduction

* Please attach a Material Safety Data Sheet or equivalent.

4.4 Will genetic sequences from the following be involved?

- ◆ HIV YES NO
- ◆ HTLV 1 or 2 or genes from any Level 1 or Level 2 pathogens YES NO
- ◆ SV 40 Large T antigen YES NO
- ◆ E1A oncogene YES NO
- ◆ Known oncogenes YES NO
- ◆ Other human or animal pathogen and or their toxins YES NO

4.5 Will virus be replication defective? YES NO

4.6 Will virus be infectious to humans or animals? YES NO

4.7 Will this be expected to increase the containment level required? YES NO

5.0 Human Gene Therapy Trials

5.1 Will human clinical trials be conducted involving a biological agent? YES NO
 (including but not limited to microorganisms, viruses, prions, parasites or pathogens of plant or animal origin)
 If no, please proceed to Section 6.0

5.2 If YES, please specify which biological agent will be used: _____
 Please attach a full description of the biological agent.

5.2 Will the biological agent be able to replicate in the host? YES NO

5.3 How will the biological agent be administered? _____

5.4 Please give the Health Care Facility where the clinical trial will be conducted: _____

5.5 Has human ethics approval been obtained? YES, number: _____ NO PENDING

6.0 Animal Experiments

6.1 Will live animals be used? YES NO If no, please proceed to section 7.0

6.2 Name of animal species to be used **C57/BL6 and Balb/C wild type mice.**

6.3 AUS protocol # **2008-033-01**

6.4 Will any of the agents listed in section 4.0 be used in live animals YES NO

6.5 Will the agent(s) be shed by the animal: YES NO

7.0 Use of Animal species with Zoonotic Hazards

7.1 Will any animals with zoonotic hazards or their organs, tissues, lavages or other body fluids including blood be used (see list below)? No If no, please proceed to section 8.0

7.2 Please specify the animal(s) used:

- ◆ Pound source dogs YES NO
- ◆ Pound source cats YES NO
- ◆ Cattle, sheep or goats YES, please specify species _____ NO
- ◆ Non-human primates YES, please specify species _____ NO
- ◆ Wild caught animals YES, please specify species & colony # _____ NO
- ◆ Birds YES, please specify species _____ NO
- ◆ Others (wild or domestic) YES, please specify _____ NO

8.0 Biological Toxins

8.1 Will toxins of biological origin be used? NO If no, please proceed to Section 9.0

8.2 If YES, please name the toxin(s) _____
Please attach information, such as a Material Safety Data Sheet, for the toxin(s) used.

8.3 What is the LD₅₀ (specify species) of the toxin _____

8.4 How much of the toxin is handled at one time*? _____

8.5 How much of the toxin is stored*? _____

8.6 Will any biological toxins be used in live animals? YES, Please provide details: _____ NO

*For information on biosecurity requirements, please see:
http://www.uwo.ca/humanresources/docandform/docs/healthandsafety/biosafety/Biosecurity_Requirements.pdf

9.0 Insects

9.1 Do you use insects? NO If no, please proceed to Section 10.0

9.2 If YES, please give the name of the species. _____

9.3 What is the origin of the insect? _____

9.4 What is the life stage of the insect? _____

9.5 What is your intention? Initiate and maintain colony, give location: _____
 "One-time" use, give location: _____

9.6 Please describe the risk (if any) of escape and how this will be mitigated:

9.7 Do you use insects that require a permit from the CFIA permit? YES NO
If YES, Please attach the CFIA permit & describe any CFIA permit conditions:

10.0 Plants

- 10.1 Do you use plants? NO If no, please proceed to Section 11.0
- 10.2 If YES, please give the name of the species. _____
- 10.3 What is the origin of the plant? _____
- 10.4 What is the form of the plant (seed, seedling, plant, tree...)? _____
- 10.5 What is your intention? Grow and maintain a crop "One-time" use
- 10.6 Do you do any modifications to the plant? YES NO
If yes, please describe: _____

- 10.7 Please describe the risk (if any) of loss of the material from the lab and how this will be mitigated:

- 10.8 Is the CFIA permit attached? YES NO
If YES, Please attach the CFIA permit & describe any CFIA permit conditions:

11.0 Import Requirements

- 11.1 Will any of the above agents be imported? NO
If no, please proceed to Section 12.0
- 11.2 Has an Import Permit been obtained from HC for human pathogens? YES NO
- 11.3 Has an import permit been obtained from CFIA for animal or plant pathogens? YES NO
- 11.4 Has the import permit been sent to OHS? YES, please provide permit # _____ NO

12.0 Training Requirements for Personnel Named on Form

All personnel named on the above form who will be using any of the above named agents are required to attend the following training courses given by OHS:

- ◆ Biosafety
- ◆ Laboratory and Environmental/Waste Management Safety
- ◆ WHMIS (Western or equivalent)
- ◆ Employee Health and Safety Orientation

As the Principal Investigator, I have ensured that all of the personnel named on the form who will be using any of the biological agents in Sections 1.0 to 9.0 have been trained.

SIGNATURE _____



